Hygienic Quality and Nutritional Value of Attiéké from Local and Imported Cassava Dough Produced with Different Traditional Starters in Burkina Faso

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Abstract

Attiéké is an essentially flavour starchy food produced from fermented cassava root. The product is widely consumed in Burkina Faso. The objective of the present study was to investigate the biochemical and the microbiological characteristics of attiéké from Burkina Faso. The samples (36) were collected from six (6) producers. Cassava dough which is used for attiéké production contains from 70.67% ± 0.25% to 86.02% ± 0.2% as starch. The undesirable organic elements rate is about 1.28 % ± 0.14% to 26.46% ± 0.53%. The inorganic impurities rate is about 0 to 7 ± 0.1 mg/100g. Cassava dough acidity varies from 1.8 ± 0.2 to 12.4 ± 0.1; and its pH from 3.48 ± 0.01 to 4.20 ± 0.01. Lactic bacteria are the main microorganisms involved in cassava dough fermentation (5.17 to 9.30 log cfu/g). Yeasts and molds number is low (<6.53 log cfu/g). Attiéké moister is about 50.6% ± 0.00% to 55.12% ± 0.7% and its protein contents from 0.77 to 1.74 ± 0.13 g/100g. The content in lipid of attiéké shows an important variation and is about 0.15 to 3.28 ± 0.32 g/100g. Carbohydrates content varies from 36.6 ± 0.04 to 47.01 ± 0.1 g/100g and its ashes content is from 140 to 780 ± 20 mg/100g. Attiéké acidity and pH are less weak than cassava dough. They vary respectively from 0.92 % ± 0.05% to 4.08% ± 0.57% and from 3.7 to 4.4 ± 0.01. As a main energizer food, attiéké energizing value is from 161.95 to 215.26 Kcal/100g. All the attiéké analyzed was exempt of aflatoxin (B1, B2, G1, G2) and ochratoxin A. The local attiéké has higher acidity, fermented bacteria load, protein and minerals salt than the imported one. The process mastery is the mainly factor that determines nutritional and sanitary quality of attiéké.

1. Introduction

Cassava (*Manihot esculenta* CRANTZ) is an important root crop in Africa, Asia, South America and India, providing energy for about 500 million people [1]. Traditionally, cassava roots are processed by different methods varying from region to region, leading to many different products like “gari”, “tapioca”, “placali” and “attiéké”. Attiéké is an essentially flavour starchy food, produced from fermented cassava dough. Attiéké is widely consumed in Burkina Faso and its production is incoming generator activity especially for women [2]. To produce attiéké, cassava roots are peeled, cut into pieces, washed and grated. During grating, the cassava dough is mixed with a traditional prepared inoculum as starter culture called “magnan”. The inoculum is prepared by storing boiled cassava roots for three days in a jute bag. As shown by Amoa-Awua *et al.* [3], the utilization of inoculum for attiéké production reduces significantly the fermentation time compared to other cassava fermented products, such as gari or chickwangue prepared without inocula [4] [5].

The inoculated dough is fermented overnight in covered container. After fermentation, the dough obtained is put into bags and pressed for several hours to remove water. The pressed cake is taken from the bags and squeezed through a sieve to obtain granules that are partially sun-dried. Fibers and waste are then removed. The granules are steamed to produce attiéké, which is sold in small plastic bags as a ready-to-eat food [5]. People in Burkina Faso enjoy it as a staple food, accompanied with salt, raw onions, spices, oil and fried fish.

The fermentation of cassava roots is very important because it helps to eliminate cyanides and to preserve, soften and produce important organic acids for the organoleptic properties of attiéké [6]-[8]. Several researchers have studied the “Attiéké” microflora [5]-[10], but the biochemical characteristics of cassava dough and attiéké produced in Burkina still remain insufficient. Therefore, the aim of the present study was to characterize cassava dough produced with four (4) different types of inocula used to ferment grated cassava into attiéké and to examine the nutritional quality of the final products (attiéké).

2. Material and Methods

2.1. Sampling

Four different types of traditional starter culture called inocula and coded A, B, C and D as described below (Table 1) were used by six (6) experienced traditional cassava processors in Burkina Faso to ferment cassava dough for attiéké processing. The cassava dough got from the four different types of cassava dough and the attiéké obtained were sampled in duplicate by using a sterile stomacher bags placed in icebox with ices and transported in the laboratory of microbiology of Food Technology Department (DTA/IRSAT/CNRST) for analysis.

In total, six (6) samples of imported cassava dough from Côte d’Ivoir, six (6) samples of cassava dough fermented with roasted inoculums, four (4) samples of cassava dough fermented with blanched inoculums and two (2) samples of cassava dough fermented with soaked inoculums were collected for microbiological analyses.

<table>
<thead>
<tr>
<th>Table 1. Characteristics of the inocula used for cassava dough production.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cassava dough</td>
</tr>
<tr>
<td>----------------</td>
</tr>
<tr>
<td>Roasted (Inoculum A)</td>
</tr>
<tr>
<td>Blanched (Inoculum B)</td>
</tr>
<tr>
<td>Soaked (Inoculum C)</td>
</tr>
<tr>
<td>Imported cassava dough (Inoculum D)</td>
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</table>
as lactic acid bacteria, molds and yeasts numeration, impurities, starch, pH and total acidity. And 18 samples of attiéké obtained from these fermented cassava dough were also collected for proximate compound determination of moisture, total carbohydrates, proteins, lipids, ash, energy value, minerals contents and mycotoxins (Aflatoxin B₁, B₂, G₁, G₂ and ochratoxin A) analyses.

2.2. Measurement of pH and Determination of Acidity

Ten gram (10 g) of each sample were dissolved in 50 ml of sterile peptoned buffered water and mixed. The pH was directly measured with a numeric pH-meter (WTW multi line P4). For Total acidity, 10 g of each sample are mixed with 50 ml of distilled water in an erlenmeyer. 10 ml of the dilution were then titrated against 0.1 N KOH using phenolphthalein as indicator; the total acidity was then calculated as a percentage of lactic acid.

2.3. Enumeration of Lactic Acid Bacteria (LAB), Yeasts and Moulds

The preparation of samples and tenfold dilutions for inoculation outs agar plates were carried out according to ISO-6887 [12]. For all numerations, 10 g of the samples were homogenized in a stomacher bags with 90 ml of sterile peptoned buffered water. The Tenfold serials dilution was prepared and spread-plated for microorganisms count. Yeasts and Molds were cultivated on Sabouraud-Chloramphenicol Agar (Oxoid LTD, Basingstore, Hampshire, England) after incubated at 25°C for 4 - 5 days and counted according to ISO 7954 standards [13]. Lactic acid bacteria were cultivated on De Man, Rogosa and Sharpe Agar (MRS, Merck 10660, Merck, Darmstadt, Germany) incubated anaerobically in an anaerobic conditions (anaerobic jar) at 37°C, for 2 - 3 days and counted according to ISO 15214 standards [14].

2.4. Proximate Composition Determination

Proximate analysis of samples was conducted using the following conventional procedures described by the Association of Official Analytical Chemists [15]. Dry matter was determinate by drying at 105°C ± 2°C overnight; ash content by incineration at 550°C for 12 h, crude protein (N × 6.25) by the Kjeldahl method after acid digestion; and crude fat content by Soxhlet extraction using n-hexane. Total carbohydrate content was determined by the phenol sulphuric acid method according to Tollier and Robin [16] and the values were expressed in g/100 g of attiéké. The starch content was determined using the colorimetric method described by Jarvis and Walker [17]. The energy value was calculated using the method described by Merrill and Watt [18].

2.5. Mineral Analysis

The contents of the minerals (Ca, Mg, Fe, Zn, Na, K, P) were determined after digesting of 0.5 g of sample using the Atomic Absorption Spectrophotometric method as outlined in the Association of Official Analytical Chemists Approved method [15].

2.6. Aflatoxin (B₁, B₂, G₁, G₂) and Ochratoxin a Analyses Contents

The analytical method used was based on liquid extraction with purification (5 g of sodium chloride + 125 ml of methanol extraction solution), followed by analysis by high-performance liquid chromatography with fluorescence detector (HPLC/FLD) and a photochemical post-column reactor according to ISO 16050 [19].

2.7. Organic Impurities

For the determination of organic impurities 100 g of each sample were placed on a sieve (mesh=1.4 mm) and drained with water. The retained residue was dried at 105°C ± 2°C for about 10 min before weighing and expressed as the organic impurities.

2.8. Inorganic Impurities

For the determination of inorganic impurities, 100 g of each sample were shaken for 30 mn in a beaker containing water. The sediment obtained was dried at 105°C ± 2°C for 10 min before weighing and expressed as the inorganic impurities.
2.9. Statistical Analysis

All the analyses were performed in triplicate. The data analysis was conducted using EXCELL and STATA version 2010. For physico-chemical and biochemical parameters, Simple Statistic Analysis was used to get means and standard deviations.

3. Results

3.1. Chemical Properties and Microflora of the Cassava Dough Fermented with Different Types of Inocula

3.1.1. The Chemical Properties of the Fermented Cassava Dough

1) Characteristics of the inocula used for cassava dough fermentation

The characteristics of the inocula used for the fermentation of the cassava dough are as showed in the following Table 1.

2) pH and acidity

Cassava sour dough has a pH ranged from 3.48 to 6.30 and its acidity is from 1.8% to 12.4% as lactic acid (Figure 1). Imported sour dough has a pH which varies from 3.56 to 4.2, and an acidity ranged from 1.8% to 9.6% some imported cassava sour dough considered as bad quality according to the processes are the least acidic (from 1.8% to 2.4%). The sour dough fermented with soaked inoculum has a weak value of acidity (2.8% to 3.2%) compared to those fermented with the blanched one (4.6% to 6.5%), Figure 1. The fermented cassava dough from the roaster inoculum is the most acid (5.6% to 6.3%). The pH of these samples is more acid than that found by Sotomey [20] and Assanvo and al. [9].

3) Undesirable organic elements

The rate of undesirable organic elements varies from 1.2 ±0.14 to 26.46 ± 0.53 g/100g (Figure 1 and Figure 2). The organic impurities rate is relatively weak. It varies from 0 to 7 mg/100g for local cassava dough and from 0 to 1 mg/100g for the imported cassava dough.

![Figure 1. Properties of cassava dough according to the type of inoculum.](image1)

![Figure 2. Physical properties of cassava dough according to producers.](image2)
4) Undesirable inorganic elements

The results show a weak value of undesirable inorganic elements. It varies from 0 to 0.07% (Figure 1). The high value of inorganic elements is observed with Gaoua and some of Ouagadougou samples. The imported cassava dough has an undesirable inorganic rate varying from 0 to 0.01% (Figure 1).

3.1.2. Microflora of the Cassava Sour Dough from Different Types of Inocula

Lactic acid bacteria counts varied from 5.11 to 9.30 log cfu/g. Except two samples (6.2 and 6.5 log cfu/g), all the samples of local cassava fermented sour dough presented high values of lactic acid bacteria compared to the imported sour dough (Figure 3). Yeasts and molds showed maximal value, about 6.53 log cfu/g. whereas no yeast and mold was observed in four imported cassava fermented dough and two local cassava fermented dough.

3.1.3. Starch Content of Cassava Fermented Dough

The starch content in cassava dough varies from 68.57 ± 1.23 to 86.2 ± 0.25 g for 100g of dry matter (DM). For local cassava dough, the dried starter and the roasted starter have the weakest rates of starch (68.57 ± 1.23 and 70.67 ± 0.25).

3.2. Properties of Attiéké Obtained from the Different Fermented Dough

3.2.1. Moister Content, pH and Acidity

The moister content of attiéké varies from 50.19 ± 0.00 to 55.12% ± 0.17% with an average of 52.92% ± 1.62% (Figure 5). Then, the dry matter is about 44.88% to 49.81%. This result is less than that found by Sotomey [20]. The attiéké lactic acidity varies from 0.92 ± 0.05 to 4.08% ± 0.57% as lactic acid, and its pH from 3.7 to 4.5 ± 0.01 (Figure 4).

3.2.2. Macronutrients (Total Carbohydrates, Fat, Protein, Ashes) Content and Energy Value

The total carbohydrate content in attiéké varies from 80.35 ± 0.87 to 97.62 ± 0.01 g/100g (Figure 5) according
Additive values of attiéké biochemical properties.

to the dry matter (DM). Attiéké is then essentially an energizer food. The proteins content in attiéké varies from 0.77 to 1.74 ± 0.13 g for 100g and from 1.6 ± 0.02 to 3.78 ± 0.27 g/100g according to the dry matter (Figure 4). Lipids content in attiéké analyzed varies from 0.15 ± 0.01 to 3.28 ± 0.31 g/100g. According to the dry matter, it is from 0.33 ± 0.02 to 7.16% ± 0.62% (Figure 4). Total ashes of attiéké vary from 140 to 780 ± 20 mg/100g at- tiéké. In relation to the dry matter, ashes rate varies from 310 ± 10 to 1610 ± 40 mg/100g (Figure 4). The ener-
gizing values of the attiéké vary from 156.46 to 215.26 Kcal/100g. Attiéké is an energizing food which content in protein and lipids is weak.

3.3. Mineral Contents in Attiéké

The minerals content of attiéké in relation with the dry matter are: phosphorus, from 150 to 524 mg/100g; pota-
sium, from 120 to 445mg/100g; sodium from 217 to 885 mg/100g; calcium, from 42 to 116 mg/100g; magn e-
sium, from 15 to 42 mg/100g; iron, from 7.05 to 14.29 mg/100g and zinc, from 0.18 to 0.73 mg/100g as shows in Table 2.

3.4. Aflatoxins and Ochratoxin a Contamination

Any attiéké sample tested reveals a contamination with aflatoxin (B1, B2, G1, and G2) or with ochratoxin A. Then, either this food has not been contaminated by the toxigenic microorganisms or attiéké analyzed have some properties which do not hallow the growth of those microorganisms.

4. Discussion

4.1. Chemical and Physicals and Microbiological Properties of Cassava Dough and Attiéké

The imported cassava fermented dough has generally the highest content in acidity. This could be explained by the species of microorganisms associate in cassava dough fermentation [9]. There is a significant difference of cassava fermented dough acidity among producers and also from one production to another. And, according to the nature of the inoculum used, there is also a significant difference of the cassava dough acidity and it micro-
bial charge.

The major part of sample has similar values of acidity to the Côte d’Ivoire norm for attiéké [21]. Sotomey [20] had found for the Beninese’s attiéké, an acidity rate of 2.12 ± 0.21 and a pH of 5.03 ± 0.24. The difference of acidity among producers and productions observed could be explained by fermentation process mastery, trans-
portation and storage conditions and also the nature of the inoculum [6] [22] [23]. Traoré [24] also proved that cassava variety influence it acidity value. The low acidity value of the bad cassava dough is in correlation with its weak load of fermented microorganisms. Acidity then mainly depends on fermentation process. The produc-
ers regulate the acidity of cassava dough to have acceptable value of attiéké acidity. The cassava dough acidity is then evaluated by tasting. This subjective evaluation is also changing from one day to another.

The impurities (organic and inorganic) constitute the third choice criteria of attiéké according to the consumers. Its frequency rate is about 36.9% [25]. Diancoumba and Sawadogo [26] identified peeling and washing as
Table 2. Minerals content of imported and local attiéké (mg/100g).

<table>
<thead>
<tr>
<th>Mineral</th>
<th>Soaked(a)</th>
<th>Roasted(b)</th>
<th>Blanched(c)</th>
<th>Roasted(b)</th>
<th>Soaked(d)</th>
<th>Imported(1)</th>
<th>Imported(2)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Phosphorus (P)</td>
<td>329</td>
<td>509</td>
<td>524</td>
<td>150</td>
<td>748</td>
<td>254</td>
<td>194</td>
</tr>
<tr>
<td>Potassium (K)</td>
<td>265</td>
<td>377</td>
<td>360</td>
<td>120</td>
<td>445</td>
<td>223</td>
<td>154</td>
</tr>
<tr>
<td>Calcium (Ca)</td>
<td>86</td>
<td>82</td>
<td>95</td>
<td>42</td>
<td>116</td>
<td>48</td>
<td>54</td>
</tr>
<tr>
<td>Magnesium (Mg)</td>
<td>30</td>
<td>34</td>
<td>36</td>
<td>15</td>
<td>42</td>
<td>20</td>
<td>23</td>
</tr>
<tr>
<td>Sodium (Na)</td>
<td>440</td>
<td>641</td>
<td>567</td>
<td>217</td>
<td>885</td>
<td>355</td>
<td>238</td>
</tr>
<tr>
<td>Iron (Fe)</td>
<td>7.93</td>
<td>7.05</td>
<td>12.36</td>
<td>7.17</td>
<td>8.96</td>
<td>11.85</td>
<td>14.29</td>
</tr>
<tr>
<td>Zinc (Zn)</td>
<td>0.73</td>
<td>0.18</td>
<td>0.31</td>
<td>0.29</td>
<td>0.62</td>
<td>0.52</td>
<td>0.59</td>
</tr>
</tbody>
</table>

\(a\): local sample from Ouagadougou; \(b\): local sample from Orodara; \(c\): local sample from Gaoua.

well as the burying underground of cassava dough as being the main process operation that govern the impurities rate. Indeed, during the peeling of the roots, the producers don't remove the central cylinder like Benin’s producers [20] which increase the organic impurities value. The weak value of inorganic impurities showed an important reduction comparatively to previous results [27]. Sotomey [20] found a similar value in average and the Côte d’Ivoire norm recommends some values ranged from 75% to 85%. Microorganisms counts of this study are less than those of the previous works [9] [11] [28].

4.2. Attiéké Nutritional Values

Starch is the main component of the total carbohydrates of attiéké. It is an energizing food which had to be balanced with other foods [29]. Sahoré and Nemlin [30] found content in proteins of 1.75% ± 0.01% and 1.70% ± 0.00% respectively from soft and bitter cassava variety, and the Côte d’Ivoire norm defines a value of 1% to 2% as the normal protein rate according to the dry matter. Sotomey [20] had found protein content of 1.87% ± 0.23% according to the dry matter. The weak value of protein content in attiéké is firstly due to the weakness content of cassava in protein [31]-[33]. Many factors can explain the protein content of attiéké: the variety of cassava used [30], soil composition, cultural technics and the process as well. The increase of proteins content in attiéké therefore depends on yeasts (unicellular protein organism) in cassava dough [34] [35]. Djoulde et al. [34] demonstrated that *Lactobacillus plantarum* and *Rhizopus oryzaeuse* as starter contribute to increase about 10% ± 2% of protein content comparatively with the traditional fermentation method.

The content in lipids of attiéké is low and contributes a little to attiéké energizing value. Sotomey [20] has found lipids content of 0.8 ± 0.09 g/100g in attiéké in relation to the dry matter. And, Sahoré and Nemlin [30] found lipids content of 1.25% ± 0.03% from soft cassava variety attiéké and 1.40 ± 0.05 g/100g from bitter cassava attiéké. Attiéké color and gluing factor depends on the nature and the quantity of oil added during the transformation process. About 0.5 l of oil is added for 55 Kg of cassava dough. The highest content in lipids of attiéké analyzed is 22 times higher than the lowest one.

Thus, attiéké macronutrients values showed an important variation which does not facilitate a proper estimation of its daily contribution to human body needs. The fluctuation of the average values is as shown in the following figure (Figure 8).

The ashes values found are similar to those of Sotomey [20] who had found on average, ashes content of 0.68 ± 0.09 g/100g. The Côte d’Ivoire norm [21] requires ashes values less than 1.4 g/100g of dry matter. Total mineral salts content of attiéké is appreciable. There is an interrelationship between the content in ashes and the origin of the attiéké (cassava). So, on average, the attiéké of Gaoua has the strongest content (0.77 ± 0.01 g/100g) followed by those of Ouagadougou (0.47 ± 0.1 g/100g) and then, the imported one (0.32 ± 0.00 g/100g) and lastly, the attiéké of Orodara (0.2 ± 0.03 g/100g). Those results could be explained by cassava variety used for attiéké production or soil content in mineral in the regions where cassava in cultivated. Processing also influences ashes content in attiéké. In fact, Ouagadougou producers used the same variety of cassava for their production but they do not have the same value of ashes content. Favier [36], Diallo et al. [7] already put out the
impact of processing on the nourishing value of the final product. According to Favier [36] the peeling operation unit only could cause the loss of 37% of cassava ashes.

The analyzed attiébé samples contained neither ochratoxins nor aflatoxins. Yandju et al. [37], found toxigenic moulds involved in the softening of cassava roots during low-moisture fermentation. But, Kastner et al. [41] did not found any contamination of aflatoxins among Côte d’Ivoire traditional inoculum. Only trace amounts of up to 0.2 mg/kg ochratoxins were present in some samples. The main toxigenic moulds are *A. parasiticus, A. flavus, A. ochraceus, A. carbonarius, P. verrucosum, P. nordicum* [39]-[41].

### 4.3. Impact of Transformation Process on Attiébé Contents

The processes influence significantly attiébé major components. The washing process influences starch and then carbohydrates content of attiébé. Sahoré and Nemlin [30] had found a variation of 13% on the total carbohydrates content according to the process. In average, the content in proteins of the attiébé produced in Ouagadougou is relatively higher than both the imported one and the one from Gaoua and Orodara. And, besides Ouagadougou producers who use the same cassava variety there is an important variation of the protein content in cassava (Figure 6).

This fluctuation is also observed with the imported dough attiébé. The variation among producers (Figure 6) and from one production to another (Figure 7), points out the impact of the processing on protein content in attiébé. Sahoré and Nemlin [30] showed that from the raw material to the final product (attiéké), the reduction of the content in protein is about 3 to 5%.

As showed in Figure 8 it appears that any producer manages to maintain a steady value of lipids content in attiébé from one production to another. The content in lipids increases the energizing value of attiébé but also presents a risk of degradation of the nutritional value during the storage. There is an important variation of moister also among producers and from one production to another. The mainly processing that influence attiébé content in moister are pressing, drying and cooking. This fluctuation is due to the fact that the processing is still traditional [40].

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**Figure 6.** Variation of biochemical parameters according to producers.

**Figure 7.** Variation of the biochemical parameters according to productions.
Many productions process influence attié ké component and therefore its nutritional value. According to the coefficient of variation, the parameters that are influenced the most by processes are impurities, lipids, ashes, protein, starch and carbohydrates as it is showed in the following picture (Figure 8).

5. Conclusions

The load of microorganisms associated in fermentation (lactic bacteria, yeasts and mildews) and the acidity of cassava dough are more important in local products than in the imported one. Local attié ké also has higher values in proteins and minerals salt. Attié ké is essentially an energizing food with a weak value of protein, and lipids. Attié ké chemical and physical components and the species of microorganisms involved in its fermentation did not allow the development of toxigenic microorganisms which produced ochratoxin or aflatoxin.

The variation of attié ké contents is tribute to processing, cassava variety (protein, carbohydrates, and minerals salt) soil content in minerals as well as farmers’ practices (protein, minerals salt, carbohydrates). Production process influences the content in starch and in total carbohydrates (peeling and washing), proteins (nature, quantity of inoculum and time of fermentation), acidity (washing, kind and quantity of inoculum and time of fermentation), lipids (oil addition), total minerals salt (peelings), and in impurities rate (good practices of production).

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References


