

Application of H-Point Standard Addition Method and Multivariate Calibration Methods to the Simultaneous Kinetic-Potentiometric Determination of Cerium(IV) and Dichoromate

Mohammad Ali Karimi^{1,2}*, Mohammad Hossein Mashhadizadeh³, Mohammad Mazloum-Ardakani⁴, Fatemeh Rahavian²

 ¹Department of Chemistry & Nanoscience and Nanotechnology Research Laboratory (NNRL), Faculty of Sciences, Payame Noor University (PNU), Sirjan, Iran
 ²Department of Chemistry, Faculty of Sciences, Payame Noor University (PNU), Ardakan, Iran
 ³Department of Chemistry, Tarbiat Moallem University of Tehran, Tehran, Iran
 ⁴Department of Chemistry, Faculty of Sciences, Yazd University, Yazd, Iran
 E-mail: ma_karimi43@yahoo.com & m_karimi@pnu.ac.ir Received June 27, 2010; revised July 20, 2010; accepted July 26, 2010

Abstract

A kinetic-potentiometric method for simultaneous determination of Cerium(IV) and dichoromate $(Cr_2O_7^{2^-})$ by H-point standard addition method (HPSAM), partial least squares (PLS) and principal component regression (PCR) using fluoride ion-selective electrode (FISE) is described. In this work, the difference between the rate of the oxidation reaction of Fe(II) to Fe(III) in the presence of Ce⁴⁺ and Cr₂O₇²⁻ is based of the method. The rate of consume fluoride ion for making complex is detected with a FISE. The results show that simultaneous determination of Ce⁴⁺ and Cr₂O₇²⁻ can be done in their concentration ranges of 1.0-30.0 and 0.1-20.0 µg/mL, respectively. The total relative standard error for applying the PLS and PCR methods on 8 synthetic samples was 2.97 and 3.19, respectively in the concentration ranges of 1.0-30.0 µg/mL of Ce⁴⁺ and 0.1-20.0 µg/mL of Cr₂O₇²⁻. In order for the selectivity of the method to be assessed, we evaluated the effects of certain foreign ions upon the reaction rate and assessed the selectivity of the method. The proposed methods (HPSAM, PLS and PCR) were evaluated using a set of synthetic sample mixtures and then applied for simultaneous determination of Ce⁴⁺ and Cr₂O₇²⁻ in different water samples.

Keywords: Multivariate Calibration, HPSAM, Simultaneous Kintic-Potentiometry, Dichromate, Cerium(IV)

1. Introduction

Dichromate $(Cr_2O_7^{2-})$ and cerium(IV) are strong oxidants in chemistry that are used widely as oxidizing agent in diverse chemical reactions in the laboratory and industry for the synthesis of many different kinds of chemical compounds. Dichromate is used as a common inorganic chemical reagent, most commonly used as an oxidising agent in various laboratory and industrial applications [1,2]. It also used to oxidize alcohols, determination ethanol, cleaning laboratory glassware of organic contaminants [3]. Cerium(IV) is also a common inorganic chemical reagent and industrially important and is used in nuclear reactors, alloys with nickel and chromium, microwave devices, lasers, agriculture and miscellaneous [4-6]. In analytical chemistry, standardized aqueous solutions of $Cr_2O_7^{2-}$ and Ce^{4+} are sometimes used as oxidizing titrants for redox titrations. Therefore, determination of these oxidants is very important. Several methods have been reported for the determination of them such as spectrophotometry [7,8], spectrofluorometry [9] and electroanalytical techniques [10,11]. In relation to the simultaneous analysis of $Cr_2O_7^{2-}$ and Ce^{4+} , Masin et al. have been reported a titrimetric method for the simultaneous determination of Ce^{4+} , MnO_4^{-} and $Cr_2O_7^{2-}$ using phosphate ion as precipitant of Ce^{4+} and Fe^{2+} as titrant [12]. To the best our knowledge, there is not any report for simultaneous determination of $Cr_2O_7^{2-}$ and Ce^{4+} using HPSAM and chemometrics methods.

In recent years the usage of chemometrics methods in electroanalytical chemistry, as in other areas of analytical chemistry, has received considerable attention as these methods can help us with extraction of more information from experimental data. Applications of HPSAM and chemometrics methods have been frequently reported for the calibration of overlapped voltammetric signals [13-16]. In the field of potentiometry, several methods have been reported based on flow injection system and titration using PLS, ANN and Kalman filter as modeling methods [17-21]. We are reported the first application of PLS and PCR multivariate calibration methods and HPSAM to the simultaneous kinetic-potentiometric determination of binary mixtures of hydrazine and its derivatives [22,23] and binary mixture of levodopa and carbidopa drugs [24]. The methods were based on the differences observed in the production rate of chloride ions in reaction of these species with N-chlorosuccinimide. The reaction rate of production of chloride ion was monitored by a chloride ion-selective electrode. Recently, we also reported the application of HPSAM, PLS and PCR methods for the simultaneous determination of binary mixtures of Fe(III) and Al(III) and ternary mixtures of Fe(III), Al(III) and Zr(IV) [25,26]. This method was based on the complex forming reaction of these metallic ions with fluoride ion that has a differential rate at certain reaction conditions. Therefore, the rate of fluoride-ion reaction with Fe(III), Al(III) and Zr(IV) was monitored by an fluoride ion-selective electrode (FISE).

This paper reports the first application of HPSAM, PCR and PLS to the simultaneous determination of oxidants binary mixtures such as $Cr_2O_7^{2-}$ and Ce^{4+} using potentiometric technique. The methods are based on the difference observed in the oxidation reaction rate of Fe(II) to Fe(III) in the presence of Ce^{4+} and $Cr_2O_7^{2-}$ as oxidants and complexing reaction between Fe(III) with fluoride ion at certain reaction conditions. The very fast response of the fluoride ion selective electrode (FISE) and its Nernstian behavior with respect to fluoride ions in acidic solutions indicated that this electrode might be employed effectively in kinetic studies of reactions involving changes in the fluoride ion concentration [25,26]. Therefore, rate of the complexing reaction of fluoride ion with Fe(III) was monitored by a FISE.

2. Experimental

2.1. Apparatus and Software

A solid-state Fluoride-selective electrode (Metrohm Model 6.0502.150) was used in conjunction with a double junction Ag/AgCl reference electrode (Metrohm Model 6.0726.100), whose outer compartment was filled with a saturated KCl solution. The Metrohm Model 780 potentiometer, attached to a Pentium (IV) computer, was

used for recording the kinetic potentiometric data. All measurements were carried out in a thermostated ($25.0 \pm 0.2^{\circ}$ C), double-walled reaction cell with continuous magnetic stirring. The electrode was stored in 1×10^{-3} mol/L potassium fluoride solution when not in use. For pH measurements, a Metrohm Model 780 pH meter with a combination glass electrode was used. PLS and PCR analyses was performed using a MATLAB 7.0 software.

2.2. Materials and Reagents

All chemicals were of analytical reagent grade and doubly distilled water was used throughout. A stock solution of iron (1000 μ g mL⁻¹) was prepared by dissolving 0.524 g of iron (II) sulfate (FeSO₄·7H₂O) in water and diluted to 100 mL. Stock solutions Ce⁴⁺and Cr₂O₇²⁻ were prepared in 100-mL flasks by dissolving 0.2885 g of cerium sulfate tetrahydrate and 0.1369 g of potassium dichromate in water and diluting with water to the mark. Cerium(IV) sulfate and potassium dichromate and salts of Fe(II) and fluoride were purchased from Merck (Germany). Acetate buffer solution (0.05 mol/L, pH 3.0) was prepared using acetic acid and NaOH solutions and adjusting its pH with a pH meter.

2.3. Procedure

Twenty five milliliters of double distilled water, 2.0 mL of buffer solution, 1.0 mL of standard fluoride solution (0.1 mol/L) and 1.0 mL of $5 \times 10^{-3} \text{ mol/L}$ of iron (II) solution were added to the thermostated $(25.0 \pm 0.2^{\circ}C)$ reaction cell. Five milliliter of the standard or sample solution of $Cr_2O_7^{2-}$, Ce^{4+} or a mixture of them were injected into the cell quickly, and after the stabilization of the potential (about 30 s), all data were recorded. The potential changes versus time were recorded at the time intervals of 1.0 s. Synthetic samples containing different concentration ratios of $Cr_2O_7^{2^-}$ and Ce^{4+} were prepared and standard additions of Cr₂O₇²⁻ were made. Simultaneous determination of $Cr_2O_7^{2-}$ and Ce^{4+} was conducted by recording the potential changes for each solution from 10 to 500 s. After each run the cell was emptied and washed twice with doubly distilled water.

Using the standard analyte solutions, we can construct a calibration graph of $(10^{\Delta E/S}-1)$ versus concentration (fixed-time method) [22-27], where ΔE is the potential variation in a selected time interval Δt and S is the slope of the fluoride electrode response, which is determined periodically by successive additions of micro-amounts of 100 µL of 1.0×10^{-5} -1.0 mol/L of NaF standard solutions in 25 mL of water mixed with 2 mL of buffer solution.

The simultaneous determination of $Cr_2O_7^{2-}$ and Ce^{4+} standard solutions with HPSAM was performed by measuring the potential changes (ΔE) at 60 and 80 s after

initiation of the reaction for each sample solution. Then plots of HPSAM of $(10^{\Delta E/S}-1)$ versus added concentration of Ce⁴⁺ were constructed for mixtures of Cr₂O₇²⁻ and Ce⁴⁺. Simultaneous determination of Cr₂O₇²⁻ and Ce⁴⁺ with PLS and PCR methods was performed by recording the potential for each solution from 10 to 500 s.

3. Results and Discussion

It is require finding the system that shows different kinetic behavior for the reaction with $Cr_2O_7^{2-}$ and Ce^{4+} . It is well-known that the rate of oxidation of Fe²⁺ with $Cr_2O_7^{2-}$ is much higher than that with Ce^{4+} [12]. Therefore, we could use from this different kinetic behavior for simultaneous determination of $Cr_2O_7^{2-}$ and Ce^{4+} . The concept for the simultaneous analysis of $Cr_2O_7^{2-}$ and Ce^{4+-} in this work is based on the difference in their oxidizing power. Preliminary studies showed that iron(II) ion as reagent at presence of fluoride ion using FISE is suitable for our purpose. Upon the addition of the oxidant (Ce^{4+} and $Cr_2O_7^{2-}$) into solution of Fe²⁺ by Ce^{4+} and $Cr_2O_7^{2-}$ take place as follows:

$$Ce^{4+} + Fe^{2+} + 2F^{-} \rightarrow [FeF_2]^{+} + Ce^{4+}$$
 (1)

$$Cr_2O_7^{2-} + 2 Fe^{2+} + 4 F + 14 H^+ \rightarrow 2 [FeF_2]^{3+} + 2 Cr^{3+} + 7 H_2O$$
 (2)

When using this ions and FISE for monitoring difference in the reaction rate, the linear range and differences of reaction rate for both two species ($Cr_2O_7^{2-}$ and Ce^{4+}) were suitable. In order to simultaneous kinetic potentiometric determination of $Cr_2O_7^{2-}$ and Ce^{4+} by HPSAM, PCR and PLS a series of experiments were conducted to establish the optimum system to achieve maximum sensitivity. Therefore, all experimental parameters affecting the reaction rate of Fe²⁺ with Ce⁴⁺ and Cr₂O₇²⁻ (response time, concentration of F⁻ and Fe²⁺, pH, etc) were carefully optimized.

3.1. Study of the Electrode Characteristics

The very fast response of FISE and its Nernstian behavior toward fluoride ions in acidic solutions indicates that this electrode might be employed effectively in the kinetic studies of reactions involving changes in the fluoride ion concentration [25,26]. The characteristics of the fluoride-selective electrode in the HNO₃-H₃BO₃-NaOH buffer were studied. In order to evaluate the operating characteristics of the FISE at pH < 5, calibration graphs were constructed for sodium fluoride in the concentration range of 1.0×10^{-5} - 1.0×10^{-1} mol/L at pHs 4.0, 3.0, 2.5 and 2.0. The slope was found to be 56.9 mV/decade and remained almost constant to 0.2 mV over 6 months of usage in this system at pH 3.0.

The effect of F⁻ concentration over the ranges of 1.0 \times 10^{-5} -1.0 × 10^{-1} mol/L fluoride ion on the linear range of calibration graph and reaction rate with Fe(II) was investigated (Figure 1). The results also indicate that the concentration of F⁻ has a great effect on the linear range and the change potential value. When the concentration of F⁻ is low, a gradual slope in the calibration graph is realized while a high concentration of F⁻ produces is high a steep slope in the calibration graph. So the fluoride concentration must be in excess, but by increasing the fluoride concentration, the potential change is decreased and the sensitivity is lower. Since, maximum differences in kinetic behaviour of Fe(III) (resulted from oxidation reaction of Fe^{2+} to Fe^{3+} by Ce^{4+} and $Cr_2O_7^{2-}$) and was observed in concentration of 1.0×10^{-3} mol L⁻¹ fluoride and both species also had larger values of potential change (ΔE) in this concentration. Therefore, a concentration of 1.0×10^{-3} mol/L fluoride was selected as the optimum concentration for further studies.

3.3. Effect of Fe²⁺ Concentration

The effect of Fe²⁺ concentration over the ranges of 1.0×10^{-5} - 1.0×10^{-1} mol/L Fe²⁺ ion on the reaction rate of Fe²⁺ with Ce⁴⁺ and Cr₂O₇²⁻ and linear range of calibration graph was investigated (**Figure 2**). The results have shown that the increase of Fe²⁺ concentration, up to 5.0×10^{-3} mol/L, causes an increase in the reaction rate of Fe²⁺ with both Ce⁴⁺ and Cr₂O₇²⁻ and the potential change, but decreased at higher concentrations. There- fore, a concentration of 5.0×10^{-3} mol/L Fe²⁺ ion was selected as the optimum concentration for further studies.

3.4. Effect of pH

Acidity of the solution influences potential response of



Figure 1. Effect of F^- concentration on the reaction of F^- and Fe^{2+} with 5.0 µg/mL of Ce^{4+} (**n**) and 10.0 µg/mL of $Cr_2O_7^{2-}$ (**•**). Conditions: 5×10^{-3} M Fe²⁺, pH 3.0, 25 °C.



Figure 2. Effect of Fe^{2+} concentration on the reaction of F^- and Fe^{2+} with 5.0 µg/mL of Ce^{4+} (**n**) and 10.0 µg/mL of $Cr_2O_7^{2-}$ (**4**). Conditions: 1×10^{-3} M F⁻, pH 3.0, 25 °C.

FISE, the oxidation reaction rate of Fe²⁺ with Ce⁴⁺ and Cr₂O₇²⁻ and complextion reaction rate of F⁻ with Fe³⁺. The results show that maximum difference in kinetic behavior of Fe³⁺ was observed at pH 3.0 (**Figure 3**). In addition, Fe³⁺ had larger values of potential change (ΔE) in this pH. Above pH 3.0, the potential change decreased evidently due to the occurrence of the hydrolysis reaction competing with the complexation between fluoride and Fe³⁺, and under pH 3.0, the potential change decreased too, probably owing to the formation of hydrogen fluoride, to which the fluoride electrode is insensitive. Thus, pH of 3.0 was selected as the optimum pH for further studies.

3.5. Composition Effect of Ground Buffer Solution

The change of potential value (ΔE) for reaction of Fe²⁺ with Ce⁴⁺ and Cr₂O₇²⁻ in the presence of certain amount fluoride ion in different acidic solutions was investigated (**Table 1**). The results have shown that in the solution of HNO₃-H₃BO₃ (pH 3.0), $\Delta E_{\rm Fe}^{3+}$ has larger values. According to obtained results, the 0.1 mol L⁻¹ boric acid-0.1 mol/L nitric acid- mixed solution (pH 3.0) containing 1.0×10^{-3} mol/L fluoride was chosen as the ground buffer solutions.

3.6. Temperature Effect of Reaction Rate

The temperature of solution evidently affects the reaction rate of the kinetic reaction. But higher temperatures do not have a positive effect on the reaction of Fe^{2+} with Ce^{4+} and $Cr_2O_7^{2-}$ and complexing reaction of Fe^{3+} with fluoride. Therefore, the temperature of solution was kept at $25 \pm 0.2^{\circ}C$ by thermostatic water bath in all of the measurements.

3.7. Potential-Time Behavior

The potential-time behavior of reactions of Fe²⁺ with

 Ce^{4+} and $Cr_2O_7^{2-}$ in the presence of F⁻ under the optimized conditions is shown in Figure 1. Figure 2 shows typical reaction curves for the reaction of Fe^{2+} with Ce^{4+} and $Cr_2O_7^{2-}$ at different concentrations. As can be seen in **Figures 4** and **5**, the reaction of $Cr_2O_7^{2-}$ is faster Ce⁴⁺ and was almost completed in 60 s after initial reaction but the reaction of $Ce^{4^{\ddagger}}$ was completed almost in 500 s. This difference in the reaction rates allowed us to design the HPSAM, PCR and PLS methods for simultaneous determination of Ce^{4+} and $Cr_2O_7^{2-}$. Characteristics of calibration graphs for the determination of Ce^{4+} and $Cr_2O_7^{2-}$, under the optimum conditions, are also given in Table 2. Therefore, above difference mentioned in the reaction rates allowed us to design the HPSAM, PCR and PLS a method for simultaneous determination of Ce⁴⁺ and $Cr_2O_7^{2-}$, in the concentration ranges of 1.0-30.0 and 0.1-20.0 µg/mL, respectively.

3.8. Requirements for Applying HPSAM

The basis of using HPSAM for treating kinetic data under the conditions that the reaction of one component is



Figure 3. Effect of pH on the reaction of F^- and Fe^{2+} with 5.0 µg/mL of Ce^{4+} (**n**) and 10.0 µg/mL of $Cr_2O_7^{2-}$ (*****). Conditions: 1×10^{-3} M F^- , 5×10^{-3} M Fe^{2+} , $25 \,^{\circ}C$.



Figure 4. Potential-time curves for the reaction of F^- and Fe^{2+} with 10.0 µg/mL of $Cr_2O_7^{2-}(a)$, 5.0 µg/mL of Ce^{4+} (b) and mixture of them (c).

At

Table 1. The values of ΔE for reaction of 10 µg/mL of Fe²⁺ with Ce(IV) and Cr₂O₇²⁻ in the presence of 1.0 × 10⁻³ M F⁻ ion in different acid solutions (pH = 3.0).

	H ₃ PO ₄ -H ₃ BO ₃	KCI-HCI	HNO ₃ - CH ₃ COOH	HNO ₃ -H ₃ BO ₃
$\Delta E_{\rm Fe}^{3+}$ (for $\rm Cr_2O_7^{2-})/mv$	2.3	7.5	8.5	13.0
$\Delta E_{\rm Fe}^{3+}$ (for ${\rm Ce}^{4+}/{\rm mv}$	1.8	5.5	6.7	10.4

Table 2. Characteristics of calibration graphs for the determination of Ce^{4+} and $Cr_2O_7^{2-}$.

Species	Linear range	Slope	Intercept	Correlation coefficient	Detection limit ^(a)
	$(\mu g/mL)$	$(mL/\mu g)$			$(\mu g/mL)$
Ce ⁴⁺	1.0-30.0	0.0681	0.0177	0.9995 (n = 10)	0.450
Cr ₂ O ₇ ²⁻	0.1-20.0	0.1119	0.2975	0.9994 (n = 10)	0.086





Figure 5. Typical potential-time curves for the reaction of F^- and Fe^{+2} with $Ce^{4+}(a)$ and $Cr_2O_7^{2-}(b)$ at different concentrations ($\mu g/mL$).

completed, while that of other component is not com-

Copyright © 2010 SciRes.

of the calibration lines. Considering a binary mixture of Ce^{4+} and $Cr_2O_7^{2-}$, for example, assume that the amount of R ($10^{\Delta E/S}$ -1) of the oxidation in the reaction of Fe²⁺ with Ce⁴⁺ and then complexation in the reaction of Fe³⁺ with F⁻ at time variables t_1 and t_2 are P_i and Q_i , respectively, while those for the $Cr_2O_7^{2-}$ -Fe²⁺-F⁻ reaction under the same conditions are Pand Q, respectively (**Figure 6**). They are equal in this case.

The following equations show the relation between them:

For Ce⁴⁺: $Q_i = P_i + m_i t_j$ ($t_1 \le t_j \le t_2$; i = 0, 1, ..., n) (1) For Cr₂O₇²⁻: $Q = P + m t_j$ (m = 0) (2) where subscripts *i* and *j* denote different solutions for *n* additions of Ce⁴⁺ concentration prepared to apply to HPSAM and the time comprising the t_1 - t_2 range, respectively.

Thus, the overall amounts of $(10^{\Delta E/S}-1)$ (or *R*) of the Ce⁴⁺-Cr₂O₇²⁻ mixture are:

$$At t_1 Rt_1 = P + P_i (3)$$

$$t_2 \qquad Rt_2 = Q + Q_i \qquad (4)$$

Simultaneous kinetic determination of concentration of Ce⁴⁺ and Cr₂O₇²⁻ by HPSAM requires the selection of two times t_1 and t_2 . To select the appropriate times, the following principles were observed. At the two selected times t_1 and t_2 , the amount of *R* of the Ce⁴⁺ must be linear with the concentrations, and the amount of *R* for Fe³⁺ must remain constant even if the Cr₂O₇²⁻ concentrations are changed. The amount of *R* for the mixture of Ce⁴⁺ and Cr₂O₇²⁻ should be equal to the sum of individual *Rs* of the two compounds. In addition, the slope difference of the two straight lines obtained at both t_1 and t_2 must be

3.5 t₂ 2.5 Q'+Qi 10 $^{\Delta E/S}$ -1 P + P1.5 Qi P 0.5 Q F -0.5 0 50 100 150 200 250 Time / Sec

Figure 6. Plot of potential changes ($10^{\Delta E/S}$ -1) for the reaction of F⁻ and Fe²⁺ with 10.0 µg/mL of Ce⁴⁺ (a), 5.0 µg/mL of Cr₂O₇²⁻ (b) and mixture of them (c).

as large as possible to achieve good accuracy. Then known amounts of Ce^{4+} are successively added to the mixture and resulting potential changes are measured at the two times and expressed:

$$R_{t1} = (10^{\Delta E(t1)/S} - 1)_{t1} = P_0 + P + M_{t1}C_i$$
(5)

$$R_{t2} = (10^{\Delta E(t2)/S} - 1)_{t2} = Q_0 + Q + M_{t2}C_i$$
(6)

where $\Delta E(t_1)$ and $\Delta E(t_2)$ are the potential changes measured at t_1 and t_2 , respectively. P_0 and Q_0 are the amounts of R for Ce⁴⁺ at a sample at t_1 and t_2 , respectively. P and Q are the amounts of R for Cr₂O₇²⁻ at t_1 and t_2 , respectively (**Figure 7**).

 M_{t1} and M_{t2} are the slopes of the standard addition calibration lines at t_1 and t_2 , respectively. C_i is the added Ce⁴⁺ concentration. The two obtained straight lines intersect at the so-called H-point (- C_H , R_H), which at point H (**Figure 7**), since $R_{t1} = R_{t2}$, $H(-C_H, R_H) \approx (-C_{\text{Ceric}}, R_{Di$ $chromate})$ from Equations (1) and (2) we have:

$$P_0 + P + M_{t1}(-C_H) = Q_0 + Q + M_{t2}(-C_H)$$
(7)

$$-C_{H} = [(Q - P) + (Q_{0} - P_{0})]/(M_{t1} - M_{t2})$$
(8)

as species $Cr_2O_7^{2-}$ is assumed not to evolve over the considered range of time, Q = P

and

$$C_H = (Q_0 - P_0)/(M_{t1} - M_{t2})$$
(9)

which is equivalent to the existing $C_{\text{Ceric}} (=P_0 / M_{t1} = Q_0 / M_{t2})$. Combining this with Equation (5) yields $R_H = P$. The overall equation for the potential at the H-point is simply represented as:

$$Q = P = R_H = R_{Fe} \tag{10}$$

The intersection of the straight lines (Equations (5) and (6) directly yields the unknown Ce⁴⁺ concentration (C_{Ceric}) and the *R* for Cr₂O₇²⁻ species ($R_{Dichromate}$) corresponding to t_1 and t_2 in the original samples, as the two times were chosen in such a way that the later species had the same *R* at both times. This analytical signal enables us to calculate the concentration of Cr₂O₇²⁻ from a calibration curve.

Since Ce⁴⁺ is selected as the analyte, it is possible to select several pairs of time ranges which present the same *R* for Cr₂O₇²⁻. Some of the selected time pairs were 60-80, 80-110, 150-200, 250-300 and 350-420 s. Greater time increments caused higher sensitivity and steeper slopes of the two time axes, as shown previously by Campins-Falco *et al.* [29]. Also, the accuracy of determinations was affected by the slope increments of H-point plots. However, the time pair that gives the greatest slope increment, lower error, and shortest analysis time was selected. For this reason, the time pair of 60-80 s as the most suitable times was employed.

A summary of the results obtained for various analyte concentrations is given in **Tables 3** and **4**. The concentration was calculated directly by solving a system of

equations of two straight lines. $Cr_2O_7^{2-}$ concentrations were calculated in each test solution by the calibration method with a single standard and ordinate value of *R*.

3.9. Multivariate Calibration

Multivariate calibration consists of establishment of a relationship between matrices of chemical data. These methods are based on a first calibration step in which a mathematical model is built using a chemical data set



Figure 7. Plot of HPSAM for simultaneous determination of a mixture of $Cr_2O_7^{2-}$ (15 µg/mL) and Ce^{4+} (25 µg/mL).

Table 3. Results of several experiments for analysis of Ce^{4+} and $Cr_2O_7^{2-}$ mixtures in different concentration ratios using HPSAM (T = 25°C).

R-C equation	r	Spiked (µg/mL)		Found (µg/mL)	
R C equation	1	Ce ⁴⁺	${\rm Cr_2O_7}^{2-}$	Ce^{4+}	$Cr_2O_7^{2-}$
$\overline{R_{80}} = 0.0322C_i + 0.4665$	0.9994	5.0	2.0	4.90 ± 0.10	2.12 ± 0.08
$R_{60} = 0.0509C_i + 0.5069$	0.9990				
$R_{80} = 0.0209C_i + 0.3056$	0.9990	3.0	3.0	3.14 ± 0.11	3.10 ± 0.10
$R_{60} = 0.0286C_i + 0.3334$	0.9999				
$R_{80} = 0.0164Ci + 0.5153$	0.9991	5.0	10.0	5.16 ± 0.13	9.85 ± 0.18
$R_{60} = 0.0239Ci + 0.5880$	0.9990				
$R_{80} = 0.0057Ci + 0.9900$	0.9930	13.0	15.0	13.20 ± 0.2	15.32 ± 0.21
$R_{60} = 0.0153Ci + 1.1312$	0.9970				
$R_{80} = 0.0052Ci + 1.5450$	0.9999	20.0	10.0	$\begin{array}{c} 19.76 \pm \\ 0.28 \end{array}$	10.12 ± 0.15
$R_{60} = 0.0155Ci + 1.6895$	0.9930				
$R_{80} = 0.0101Ci + 1.9061$	0.9950	25.0	15.0	24.70 ± 0.24	15.38 ± 0.28
$R_{60} = 0.0058Ci + 1.8365$	0.9995				
$R_{80} = 0.0014Ci + 0.9954$	0.9920	15.0	5.0	15.27 ± 0.23	5.34 ± 0.15
$R_{60} = 0.0107Ci + 1.0391$	0.9992				

Table 4. Results of six replicate experiments for analysis of Ce^{4+} and $Cr_2O_7^{2-}$ mixture using HPSAM (T = 25°C).

P.C. aquation		Spiked	l (µg/mL)	Found (µg/mL)		
<i>R</i> - <i>C</i> equation	r	Ce ⁴⁺	$Cr_2O_7^{2-}$	Ce ⁴⁺	$Cr_2O_7^{2-}$	
$\overline{R_{80} = 0.1090C_i} + 1.0720$	0.9902	13	15	12.98(99.8)	15.51(103.4)	
$R_{60} = 0.0046C_i + 0.9736$	0.9900					
$R_{80} = 0.0094C_i + 1.0276$	0.9980	13	15	12.79(98.3)	14.75(98.3)	
$R_{60} = 0.0040C_i + 0.9479$	0.9999					
$R_{80} = 0.0153C_i + 1.1312$	0.9980	13	15	13.10(100. 7)	14.80(98.6)	
$R_{60} = 0.0057C_i + 0.9900$	0.9933					
$R_{80} = 0.0104C_i + 1.0300$	0.9990	13	15	12.70(97.7)	14.40(96.0)	
$R_{60} = 0.0056C_i + 0.9611$	0.9930					
$R_{80} = 0.0154C_i$ + 1.1488	0.9930	13	15	13.30(102. 3)	14.80(98.6)	
$R_{60} = 0.0056C_i + 1.0034$	0.9940					
$R_{80} = 0.0156C_i + 1.0520$	0.9990	13	15	12.60(96.9)	15.0(100.0)	
$R_{60} = 0.0055C_i + 0.9567$	0.9940					
Mean				13.01	14.87	
SD				0.28	0.39	
RSD (%)				2.16	2.60	

(e.g., potential values) and a concentration matrix data set [30-34]. The calibration is followed by a prediction set in which this model is used to estimate unknown concentrations of a mixture from kinetic profile. Multivariate calibration methods are being successfully applied to the multi-component kinetic determination to overcome some of the drawbacks of classical methods. The theories and applications of chemometrics methods such as PCR and PLS, to the analysis of multi-component mixtures, have been discussed by several workers [31-35]. PCR and PLS modeling are powerful multivariate statistical tools, which are successfully applied to the quantitative analysis of spectrochemical and electrochemical data [33-36]. The first step in the simultaneous determination of the species by PCR and PLS methodologies involves the construction of calibration matrix for the binary mixture of Ce^{4+} and $Cr_2O_7^{2-}$. For constructing the calibration set, factorial design was applied to five levels in order to extract a great deal of quantitative information, using only a few experimental trials. In this research, a synthetic set of 34 solutions, including different concentrations of Ce^{4+} and $Cr_2O_7^{2-}$, was prepared. A collection of 26 solutions was selected as the calibration set (Table 5) and the other 8 solutions were used as the prediction set (Table 6). Their composition was randomly designed to obtain more information from the calibration procedure. Changes in the solution potential were recorded during a time period of 500 seconds.

To select the number of factors in the PCR and PLS algorithm, as a cross-validation method, leaving out one sample method was employed [37]. The prediction error was calculated for each species of the prediction set. This error was expressed as the prediction residual error sum of squares (PRESS):

$$PRESS = \sum_{i=1}^{m} \left(\hat{\mathbf{C}}_{i} - \mathbf{C}_{i} \right)^{2}$$
(11)

Where m is the total number of calibration sample, \hat{C}_i represents the estimated concentration while C_i is the reference concentration for the *i*th sample left out of the

Table 5. Calibration set for constructing PCR and PLS methods in determination of Ce^{4+} and $Cr_2O_7^{2-}$ (µg/mL).

Sample	Ce ⁴⁺	$Cr_2O_7^{2-}$	Sample	Ce ⁴⁺	$Cr_2O_7^{2-}$
1	5.0	2.0	14	22.0	15.0
2	5.0	6.0	15	22.0	18.0
3	5.0	12.0	16	1.0	0.5
4	5.0	20.0	17	1.0	6.0
5	10.0	10.0	18	1.0	4.0
6	10.0	15.0	19	25.0	17.0
7	10.0	19.0	20	25.0	19.0
8	13.0	17.0	21	25.0	20.0
9	13.0	20.0	22	18.0	7.0
10	3.0	3.0	23	18.0	15.0
11	3.0	9.0	24	15.0	5.0
12	3.0	17.0	25	15.0	9.0
13	22.0	14.0			

Table 6. Prediction set for constructing PLS and PCR methods in determination of Ce^{4+} and $Cr_2O_7^{2-}$.

Solution	Synthetic		Predicted (µg/mL)					
Solution	(με	g/mL)	PL	.S ^a	РС	CR ^a		
	Ce4+	Cr ₂ O ₇ ²⁻	Ce ⁴⁺	Cr ₂ O ₇ ²⁻	Ce ⁴⁺	Cr ₂ O ₇ ²⁻		
1	13.0	15.0	13.40(103.0)	15.80(105.3)	13.70(105.3)	15.60(104.0)		
2	3.0	13.0	2.80(93.3)	13.40(103.0)	2.78(92.6)	13.00(100.0)		
3	5.0	10.0	5.10(102.0)	10.20(102.0)	4.80(96.0)	9.90(99.0)		
4	13.0	18.0	13.50(103.8)	18.50(102.7)	13.50(103.8)	17.80(98.8)		
5	13.0	16.0	12.50(96.1)	15.40(96.2)	12.60(96.9)	15.30(95.6)		
6	22.0	16.0	22.30(101.3)	15.70(98.1)	22.60(102.7)	15.40(96.3)		
7	10.0	13.0	9.60(96.0)	12.70(97.7)	9.80(98.0)	13.10(100.7)		
8	3.0	5.0	3.20(106.6)	5.30(106.0)	3.10(103.3)	4.90(98.0)		
Mean recovery			100.2	101.3	99.8	99.1		
RMSEP			2.97	3.40	3.03	2.90		
RSEP(%)			3.19		2.97			

^a Predicted mean (recovery percent)

calibration during the cross validation. **Figure 8** shows a plot of PRESS against the number of factors for a mixture of components. To find out minimum factors, we also used the F-statistics to carry out the significant determination [33]. The optimal number of factors, for the two components, was obtained as 2 for both PCR and PLS.

For evaluating the predictive ability of a multivariate calibration model, the root mean square error of prediction (RMSEP), relative standard error of prediction (RSEP) and squares of correlation coefficient (R^2), which is an indication of the quality fit of all the date to a straight line, can be used as follows [33,37]:

$$RMSEP = \left(\sum_{i=1}^{N} \left(\hat{C}_{i} - C_{i}\right)^{2} / n\right)^{\frac{1}{2}}$$
(12)

$$RSEP(\%_{0}) = \left(\sum_{i=1}^{N} \left(\hat{C}_{i} - C_{i}\right)^{2} / \sum_{i=1}^{N} \left(C_{i}\right)^{2}\right)^{\frac{1}{2}} \times 100$$
(13)

$$R^{2} = \sum_{i=1}^{N} \left(\hat{C}_{i} - C' \right)^{2} / \sum_{j=1}^{N} \left(C_{i} - C' \right)^{2}$$
(14)

where \hat{C}_i represents the estimated concentration, C_i and n are the actual analyte concentration and the number of samples, respectively.

Table 7 shows the values of RSEP, RMSEP and R^2 for each component using PLS and PCR. It is shown that the obtained values, for the statistical parameters, are almost the same for both PLS and PCR methods.

3.10. Interference Study

The study of interference ions was performed by a standard mixture solution containing 10.0 µg/mL of each Ce⁴⁺ and Cr₂O₇²⁻ and a certain amount of foreign ions. The following excesses of ions do not interfere (*i.e.*, caused a relative error of less than 5%): more than a 1000-fold (largest amount tested) amount of K⁺, Zn²⁺, Cu²⁺, Bi³⁺, As³⁺, Cd²⁺, Mg²⁺, Be²⁺, Cl⁻, NO₃⁻, BO₃⁻³⁻, C₂O₄²⁻ a 100-fold amount of Mn²⁺, Ni²⁺, Co²⁺, pb²⁺, Cr³⁺, Ca²⁺; a 10-fold amount of SO₄⁻²⁻, PO₄⁻³⁻, Hg²⁺ and a 1-fold amount of Al³⁺, Fe³⁺, Zr⁴⁺, Ti⁴⁺, I. As was to be expected, Al^{3+} , Fe^{3+} , Zr^{4+} , Ti^{4+} have a interference effect because complex forming reaction of these metallic ions with fluoride ion. The interference of iodide ion was also because oxidation reaction with Ce⁴⁺ and Cr₂O₇²⁻ as oxidants.

3.11. Application

To evaluate the analytical applicability of the proposed methods (PCR, PLS and HPSAM), we spiked known amounts of both Ce^{4+} and $Cr_2O_7^{2-}$ into some water samples. The proposed methods were applied to determine analytes simultaneously and satisfactory results were obtained. The results are given in **Table 8**. The results show that the proposed methods could accurately determine the concentration of these oxidants mixture under investigation in real water samples, and there is no significant difference between the results of PCR, PLS and HPSAM for their simultaneous determination.

 Table 7. Statistical parameters calculated for the prediction set using PLS and PCR models.

	RSE	P (%)	RMSEP		R^2	
Component	PLS	PCR	PLS	PCR	PLS	PCR
Ce ⁴⁺	2.97	3.03	0.35	0.36	0.9980	0.9990
$Cr_2O_7^{2-}$	3.40	2.90	0.64	0.40	0.9993	0.9988



Figure 8. Plot of PRESS against the numbers of factors PLS(♦) and PCR(■).

Sample	Snike	d (ug/mI)	Found (µg/mL)						
Sample	ample Spiked (µg/mL)		HPS	HPSAM		PLS		CR	
	Ce^{4+}	$Cr_2O_7^{2-}$	Ce ⁴⁺	$Cr_2O_7^{2-}$	Ce ⁴⁺	$Cr_2O_7^{2-}$	Ce ⁴⁺	$Cr_2O_7^{2-}$	
1	4.0	0.5	4.20 ± 0.12	0.51 ± 0.10	4.10 ± 0.11	0.46 ± 0.05	3.97 ± 0.20	0.48 ± 0.02	
	10.0	10.0	10.20 ± 0.24	10.40 ± 0.30	10.10 ± 0.22	9.88 ± 0.20	9.79 ± 0.14	9.90 ± 0.10	
2	3.0	15.0	2.90 ± 0.06	15.40 ± 0.25	2.97 ± 0.05	14.80 ± 0.28	3.10 ± 0.10	15.20 ± 0.20	
	7.0	15.0	7.20 ± 0.21	15.30 ± 0.33	7.14 ± 0.18	14.88 ± 0.15	6.85 ± 0.13	14.90 ± 0.11	
3	25.0	4.0	24.60 ± 0.36	4.10 ± 0.10	24.8 ± 0.33	3.88 ± 0.09	25.4 ± 0.28	4.16 ± 0.15	
_	18.0	20.0	17.80 ± 0.28	19.30 ± 0.20	17.65 ± 0.37	19.10 ± 0.12	18.20 ± 0.13	19.8 ± 0.21	

4. Conclusions

This work as the first application of PCR, PLS and HPSAM in the simultaneous determination of the binary mixture of Ce^{4+} and $Cr_2O_7^{2-}$ shows the ability and excellent performance of ISEs as detectors not only for individually determination of produced or consumed ions, but also in the simultaneous kinetic-potentiometric analysis. In addition, this paper has also demonstrated that the ability and advantages of the HPSAM and chemometrics methods such as PCR and PLS, ISEs and kinetic methods produce a very attractive and excellent technique for the analysis of multi-component oxidant mixtures. Other chemometrics approaches like ANN, ISEs (flouride, bromide, iodide, etc) and other kinetic reactions in which the rate of production or consumption of the corresponding ion is different can also be used. Our team has obtained good results for the simultaneous determination of other species using HPSAM, chemometrics methods, different ISEs and various reaction systems and our complete results will be presented for publication in the future.

5. Acknowledgements

The authors would like to express their appreciations to Professor Afsaneh Safavi and Professor Hamid Abdollahi for their valuable discussion and useful suggestions. This research was supported by the Payame Noor Universities of Ardakan and Sirjan.

6. References

- K. Basavaiah and B. C. Somashekar, "Quantitation of Ranitidine in Pharmaceuticals by Titrimetry and Spectrophotometry Using Potassium Dichromate as the Oxidimetric Reagent," *Journal of the Iranian Chemical Society*, Vol. 4, No. 1, 2007, pp. 78-88.
- [2] O. Z. Zhai, "Catalytic Kinetic Spectrophotometric Determination of Trace Amounts of Ethylenediaminetetraacetic Acid Based on its Catalytic Effect on the Reaction of Rhodamine B and Potassium Dichromate," *Instrumentation Science and Technology*, Vol. 29, No. 38, 2010, pp. 72-82.
- [3] J. C. Dacons, H. G. Adolph and M. J. Kamlet, "Selective Solvent-Free Oxidation of Alcohols with Potassium Dichromate," *Tetrahedron Letters*, Vol. 43, No. 49, 2002, pp. 8843-8844.
- [4] J. Sumaoka, W. Chen, Y. Kitamura, T. Tomita, J. Yoshida and M. Komiyama, "Application of Cerium(IV)/ EDTA Complex for Future Biotechnology," *Journal of Alloys and Compounds*, Vol. 408-412, 2006, pp. 391-395.
- [5] I. D. Nickson, C. Boxall, A. Jackson and G. O. H. Whillock, "A Spectrophotometric Study of Cerium IV and Chromium VI Species in Nuclear Fuel Reprocessing Process Streams," *IOP Conference Series: Materials*

Science and Engineering, Vol. 9, No. 1, 2010.

- [6] H. J. Vieira and O. Fatibello-Filho, "Indirect Flow Injection Determination of N-acetyl-L-cysteine Using Cerium (IV) and Ferroin," *Química Nova*, Vol. 28, No. 5, 2005, pp. 797-800.
- [7] Y. Liu and P. Wang, "Kinetic Spectrophotometric Method for the Determination of Cerium(IV) with Naphthol Green B," *Rare Metals*, Vol. 28, No. 1, 2009, pp. 5-8.
- [8] A. M. Stoyanova, "Catalytic Spectrophotometric Determination of Chromium", *Turkish Journal of Chemistry*, Vol. 29, No. 4, 2005, pp. 367-375.
- [9] M. M. Karim, S. H. Lee, Y. S. Kim, H. S. Bae and S. B. Hong, "Fluorimetric Determination of Cerium(IV) with Ascorbic Acid," *Journal of Fluorescence*, Vol. 16, No. 1, 2006, pp. 17-22.
- [10] M. Mazloum-Ardakani, M. Salavati-Niasari, A. Dastanpour, "Determination of Chromate and Dichromate by Highly Selective Coated-Wire Electrode Based on Bis (Acetylacetonato)Copper(II)," *Bulletin of Electrochemistry*, Vol. 20, No. 5, 2004, pp. 193-197.
- [11] H. Rajantie and D. E. Williams, "Electrochemical Titration of Thiosulfate, Sulfite, Dichromate and Permanganate Using Dual Microband Electrodes," *Analyst*, Vol. 126, No. 1, 2001, pp. 86-90.
- [12] V. Mašín and J. Doležal, "Simultaneous Determination of Ce⁴⁺, MnO₄⁻ and Cr₂O₇²⁻ without Separation," *Fresenius' Journal of Analytical Chemistry*, Vol. 301, No. 1, 1980, p. 25.
- [13] E. Shams, H. Abdollahi, M. Yekehtaz and R. Hajian, "H-Point Standard Addition Method in the Analysis by Differential Pulse Anodic Stripping Voltammetry. Simultaneous Determination of Lead and Tin," *Talanta*, Vol. 63, No. 2, 2004, pp. 359-364.
- [14] E. Shams, H. Abdollahi and R. Hajian, "Simultaneous Determination of Copper and Bismuth by Anodic Stripping Voltammetry Using H-Point Standard Addition Method with Simultaneous Addition of Analytes," *Electroanalysis*, Vol. 17, No. 17, 2005, pp. 1589-1594.
- [15] K. Zarei, M. Atabati and M. Karami, "H-Point Standard Addition Method Applied to Simultaneous Kinetic Determination of Antimony(III) and Antimony(V) by Adsorptive Linear Sweep Voltammetry," *Journal of Hazardous Materials*, Vol. 179, No. 1-3, 2010, pp. 840-844.
- [16] F. Ahmadi and F. Bakhshandeh-Saraskanrood, "Simultaneous Determination of Ultra Trace of Uranium and Cadmium by Adsorptive Cathodic Stripping Voltammetry Using H-Point Standard Addition Method," *Electroanalysis*, Vol. 22, No. 11, 2010, pp. 1207-1216.
- [17] J. Mortensen, A. Legin, A. Ipatov, A. Rudnitskaya, Y. Vlasov and K. Hjuler, "A Flow-Injection System Based on Chalcogenide Glass Sensors for Determination of Heavy Metals," *Analytica Chimica Acta*, Vol. 403, No. 1-2, 2000, pp. 273-277.
- [18] M. Slama, C. Zaborosch, D. Wienke and F. Spener, "Chemometrical Studies on Mixture Analysis with a Single Dynamic Microbial Sensor," *Sensors and Actuators B: Chemical*, Vol. 44, No. 1-3, 1997, pp. 286-290.

- [19] N. Garcia-Villar, J. Saurina and S. Hernández-Cassou, "Flow Injection Differential Potentiometric Determination of Lysine by Using a Lysine Biosensor," *Analytica Chimica Acta*, Vol. 477, No. 2, 2003, pp. 315-324.
- [20] M. Akhond, J. Tashkhourian and B. Hemmateenejad, "Simultaneous Determination of Ascorbic, Citric and Tartaric Acids by Potentiometric Titration with PLS Calibration," *Journal of Analytical Chemistry*, Vol. 61, No. 8, 2006, pp. 804-808.
- [21] Y. Z. Ye and Y. Luo, "Kinetic Potentiometry Simultaneous Determination of Iron(III) and Zirconium(IV) by Using the Kalman Filter," *Laboratory Robotics and Automation*, Vol. 10, No. 5, 1998, pp. 283-287.
- [22] M. A. Karimi, M. Mazloum-Ardakani, H. Abdollahi and F. Banifatemeh, "Application of H-Point Standard Addition Method and Partial Least Squares to the Simultaneous Kinetic-Potentiometric Determination of Hydrazine and Phenylhydrazine," *Analytical Science*, Vol. 24, No. 2, 2008, pp. 261-266.
- [23] M. A. Karimi, H. Abdollahi, H. Karami and F. Banifatemeh, "Simultaneous Kinetic-Potentiometric Determination of Hydrazine and Thiosemicarbazide by Partial Least Squares and Principle Component Regression Methods," *Journal of the Chinese Chemical Society*, Vol. 55, No. 1, 2008, pp. 129-136.
- [24] M. A. Karimi, M. H. Mashhadizadeh, M. Mazloum-Ardakani and N. Sahraei, "Simultaneous Kinetic-Potentiometric Determination of Levedopa and Carbidopa Using Multivariate Calibration Methods", *Journal of Food and Drug Analysis*, Vol. 16, No. 5, 2008, pp. 39-47.
- [25] M. A. Karimi, M. Mazloum Ardakani, R. Behjatmanesh Ardakani and M. R. Zand Monfared, "Simultaneous Kinetic-Potentiometric Determination of Fe³⁺ and Al³⁺ Using H-point Standard Addition and Partial Least Squares Methods," *Chemical Analysis (Warsaw)*, Vol. 54, No. 6, 2009, pp. 1321-1337.
- [26] M. A. Karimi, M. Mazloum-Ardakani, R. Behjatmanes Ardakani, M. H. Mashhadizadeh, M. R. Zand Monfared and M. Tadayon, "Chemometrics-Assisted Kinetic-Potentiometric Methods for Simultaneous Determination of Fe(III), Al(III), and Zr(IV) Using a Fluoride Ion-Selective Electrode," *Journal of AOAC International*, Vol. 93, No. 1, 2010, p. 327.
- [27] E. Athanasiou-Malaki, M. A. Koupparis and T.P. Hadjiioannou, "Kinetic-Potentiometric Study and Analytical Applications of Micellar-Catalysed Reactions of 1- Fluoro-2,4-Dinitrobenzene with Amino Compounds," *Analytica Chimica Acta*, Vol. 219, No. 2, 1989, pp. 295-307.
- [28] K. Srinivasan and G. A. Rechnitz, "Activity Measurements with a Fluoride-Selective Membrane Electrode," *Analytical Chemistry*, Vol. 40, No. 3, 1968, pp. 509-512.

- [29] F. Bosch-Reig, P. Campins-Falco, A. Sevillano-Cabeza, R. Herraez-Hernandez and C. Molins-Legua, "Development of the H-Point Standard Additions Method for Ultraviolet-Visible Spectroscopic Kinetic Analysis of Two-Component Systems," *Analytical Chemistry*, Vol. 63, No. 21, 1991, pp. 2424-2429.
- [30] M. Bahram, K. Farhadi, A. Afkhami, D. Shokatynia and F. Arjmand, "Simultaneous Kinetic Spectrophotometric Determination of Cu(II), Co(II) and Ni(II) Using Partial Least Squares (PLS) Regression," *Central European Journal of Chemistry*, Vol. 7, No. 3, 2009, pp. 375-381.
- [31] M.A. Karimi, M. Mazloum-Ardakani, R. Behjatmanesh-Ardakani, M. H. Mashhadizadeh and N. Zarea Zadeh, "Application of H-Point Standard Addition Method and Multivariate Calibration Methods to the Simultaneous Kinetic-Potentiometric Determination of Hydrogen Peroxide and Peracetic Acid," *Analytical & Bioanalytical Electrochemistry*, Vol. 1, No. 3, 2009, pp. 142-158.
- [32] M. Chamsaz, A. Safavi and J. Fadaee, "Simultaneous Kinetic-Spectrophotometric Determination of Carbidopa, Levodopa and Methyldopa in the Presence of Citrate with the Aid of Multivariate Calibration and Artificial Neural Networks," *Analytica Chimica Acta*, Vol. 603, No. 2, 2007, pp. 140-146.
- [33] M. Otto and W. Wegscheider, "Spectrophotometric Multicomponent Analysis Applied to Trace Metal," *Analytical Chemistry*, Vol. 57, No. 1, 1985, pp. 63-69.
- [34] M. A. Karimi, M. Mazloum-Ardakani, R. Behjatmaneh-Ardakani, M. R. Hormozi Nezhad and H. Amiryan, "Individual and Simultaneous Determinations of Phenothiazine Drugs Using PCR, PLS and (OSC)–PLS Multivariate Calibration Methods," *Journal of the Serbian Chemical Society*, Vol. 72, No. 2, 2008, pp. 233-247.
- [35] M. A. Karimi, M. Mazloum-Ardakani, O. Moradlou, R. Behjatmanesh-Ardakani and F. Banifatemeh, "Simultaneous Kinetic-Spectrophotometric Determination of Hydrazine and its Derivatives by Partial Least Squares and Principle Component Regression Methods," *Journal of the Chinese Chemical Society*, Vol. 54, No. 1, 2007, pp. 15-21.
- [36] D. M. Haaland and E. V. Thomas, "Partial Least-Squares Methods for Spectral Analyses. 1. Relation to other Quantitative Calibration Methods and the Extraction of Qualitative Information," *Analytical Chemistry*, Vol. 60, No. 11, 1988, pp. 1193-1202.
- [37] M. A. Karimi, M. Mazloum-Ardakani, M. H. Mashhadizadeh and F. Banifatemeh, "Simultaneous Kinetic Spectrophotometric Determination of Hydrazine and Isoniazid Using H-Point Standard Addition Method and Partial Least Squares Regression in Micellar Media," *Croatica Chemica Acta*, Vol. 84, No. 4, 2009, pp. 729-738.