Real Time Recrystallization Study of 1, 2 Dodecanediol on Highly Oriented Pyrolytic Graphite (HOPG) by Tapping Mode Atomic Force Microscopy

S. K. Dora
Nanomagnetics Instruments Pvt. Ltd., Ankara, Turkey
Email: shamrid285@gmail.com

Abstract
Real-time atomic force microscopy (AFM) imaging revealed for the first time, the detailed growth/dissolution mechanism of Lauryl glycol (LG) or 1, 2 dodecanediol molecules on HOPG at the nano-level after recrystallizing them from chloroform solution. At the early stage of recrystallization, parallelogram-like-structures having lengths of several microns and distinct widths (between ~ 100 - 400 nm) were observed. Growth/dissolution behavior of these parallelogram-like-structures as a function of time was investigated. While dissolution occurred along all three dimensions, growth was found to be strictly two dimensional. Both the growth and dissolution process were found to be logarithmic in nature. The average growth rates along their length and width were found to be 11 nm/min and 1.5 nm/min respectively. Average dissolution rate in percentage on HOPG surface was found to be ~ 0.078%/min. Based upon the recrystallization of LG molecules schematics are drawn for a better understanding of the recrystallization process.

Keywords
1, 2 Dodecanediol, Kinetics, Real Time Growth, Tapping Mode AFM

1. Introduction
1, 2-Alkanediols have been used in household and cosmetic products for a long time as solvents [1], cryoprotectors [2], moisturizers [3], and viscosity modifiers [1] [4]. Because of low toxicities and good antimicrobial activities of 1, 2-alkanediols, they have gradually been replaced for the traditional preservatives such as
parabens, potassium sorbate, sodium benzoate, and formaldehyde-releasing compounds in cosmetic products [1] [5]. Diols with short aliphatic chains have been known to exhibit antimicrobial activities, depending on their chain length and the position of the hydroxyl groups [1] [6] [7]. On the other hand, long chain alkane diols (–OH group at different positions) are used as biomarkers [8]. They are also an important constituent in plant cuticular waxes [9] [10] [11]. 1, 2 dodecanediol otherwise known as lauryl glycol (LG) is a 12-carbon chain alkane with –OH groups at 1, 2 position. It has a variety of potential uses. For instance, it has been used to prepare lipophilic diamines and amino alcohols [12] which show activity against tuberculosis bacteria. It is also used as a reducing agent in the preparation of Fe(II), Co(II) and Ni(II) nanoparticles [13]. Freital et al. have demonstrated the importance of LG in the Ketalisation of ethyl levulinate resulting in a new class of biodegradable surfactants [14]. LG can also be found as one of the constituents in many cosmetic products used for skin [15]. Wilbur Johnson Jr and coworkers demonstrated the dermal (skin) absorption rate was up to 40% for lauryl alcohol [15] on pig skin. The surface epithelium of the skin of animal is nonpolar in nature. Therefore it is interesting to understand how the nonpolar surfaces affect the absorption of LG molecules and understanding their detailed absorption mechanism will ultimately help in preparing better skin cosmetic products. In addition, understanding of adsorption phenomenon at nano-level may also shed new light on the efficacy of LG as a reducing agent. As a model substrate, HOPG was chosen for the study. HOPG is nonpolar and crystalline in nature and is one of the most widely used substrates for self-assembly studies for a wide variety of molecules [16] [17] [18] [19].

As a suitable technique to study the recrystallization process of LG molecules atomic force microscope (AFM) in tapping mode was employed. One of the biggest advantages of atomic force microscopy (AFM) is its ability to work in all environments (vacuum, in air, in liquids) compared to other surface characterization techniques e.g. electron microscopes. In the field of crystal growth this versatility translates into the application for investigating ex-situ and in-situ crystal growth, whereby ex-situ usually means that after separation from their growth environment crystals are examined by AFM in air, and in-situ generally means crystal growth from solution phase examined under liquid environment. AFM has been used extensively for studying growth characteristics of numerous different crystals both organic and inorganic [20] [21] [22] [23] molecules. Real time growth characteristics of crystals are also observed using an AFM [24] [25] [26]. In this work, we have examined the real time growth process of lauryl glycol on HOPG. Visualization of the process was observed in ex-situ AFM using tapping mode after recrystallizing them from a chloroform solution on to a HOPG substrate. To my knowledge, AFM has not yet been employed to study the real time growth process of any kind of diols, although self-assembled monolayer STM study of another diol (1, 12 dodecanediol) is reported in literature [27]. There is obviously a strong incentive to understand how diols behave as an absorption material on a nanoscale which will ultimately provide key insights
about their interaction with nonpolar substrates such as animal skin when used as constituents in cosmetics or their role as reducing agents.

2. Experiment

LG (C₁₂H₂₆O₂, racemic mixture) was purchased from Sigma and used without further purification. AFM measurements were carried out in tapping mode using an Agilent 5500 system under ambient conditions. PPP-NCH cantilevers from Nanosensors with a nominal resonant frequency of ~300 kHz were employed for real time observation of crystal growth process. Imaging conditions used were a scan size of 4.9 × 4.9 µm², a scan rate of 0.612 Hz, and an image size of 256 × 256 pixels. The set point was chosen close to the lowest limit possible and all measurements were performed at room temperature. Real time recrystallization process was observed by AFM, after applying a 10 µL droplet of appropriate concentration (exact concentrations used are described in the result section) of lauryl glycol dissolved in chloroform on to the HOPG surface. The chloroform took ca. 30 seconds to evaporate from the surface leaving the LG molecules attached to the HOPG substrate. The substrate was then mounted to the AFM sample plate. The first image acquisition started as soon as possible after the chloroform evaporation, typically after ~10 minute. AFM images were taken consecutively from the same substrate area over a period of several hours applying a constant scan rate of 0.612 Hz. Height and length measurements of the grown crystals were made with the program WSxM (Version 4.0 Beta 8.3; Nanotec Electronica, Madrid, Spain). In addition, commercially available Pico image software from Keysight technology was used to determine the surface coverage of LG molecules.

3. Results

This section describes details of recrystallization process of LG molecules on HOPG substrate. Concentration of LG molecules in chloroform was also varied to understand whether it has any effect on recrystallization process.

Time evolution of structural changes after recrystallizing 0.4 mg/ml LG molecules from chloroform on to a freshly cleaved HOPG surface is presented in a series of AFM images in Figure 1. Although images were recorded for over 20 hours, only 5 representative images at different time intervals are presented here. Whole image sequence as video can be found as supplementary material (Video 1). As shown in Figure 1(a) which is taken after ~14 min, recrystallization of LG resulted in the formation of parallelogram-like-structures of variable length, width and height. The terms length, width and height of these structures are defined in the schematic (see Figure 2(a)). The average width of these structures varies between ~100nm to ~400 nm. Width profile of two such structures (see Figure 1(g) and Figure 1(h)) marked by black and blue line in Figure 1(a) gives values of ~175 nm and ~220 nm respectively. The random variation of widths within ~100 - 400 nm range makes it practically impossible to draw in any statistics for their width variation. On the other hand, structures having widths of
Figure 1. Series of AFM images showing LG molecule recrystallization on HOPG at different time intervals (~14 - 1063 min) after applying 0.4 mg/ml wax solution (chloroform solvent) to the substrate. Figure 1(a) taken after ~14 minutes shows parallelogram-like-structures distributed over HOPG substrates. Figure 1(b)-(e) taken after ~29, ~170, ~575 and ~1063 min respectively demonstrate the details of growth and dissolution process of these structures (for details see texts in result section). Figure 1(f) shows the height profile across the black line in Figure 1(d) indicating each such layer is formed by the close packing of two vertically stacked LG molecules one over other. Figure 1(g) and Figure 1(h) show the widths across the black and blue line in Figure 1(a). Size = 4.9 × 4.9 µm², scan rate = 0.612 Hz, 256 lines/image.

<100 nm or >400 nm are not observed at all during the initial stage of experiment. One exception to these structures marked by black arrow in Figure 1(a) shows a width >800 nm. However, this is just one of case and was not observed in other areas of the HOPG surface at all. So, I believe this structure is formed by individual structures lying sidewise very close to each other or overlapping partially on top of each other making the width look larger. Length of these structures varies from few hundred nanometers to several microns. For example, the shortest (solid white circle) and longest structure (solid green circle) in Figure 1(a) have lengths of ~300 nm and ~2 µm respectively. Structures of longer length were also observed in other areas of HOPG surface (figure not shown) indicating that the length of these structures also vary randomly. Taking in to account heights of parallelograms lying directly on HOPG surface, it was found that their heights vary between ~14 - 24 nm. See for example, the height profiles in Figure 1(g), h having heights of ~16 and 24 nm. In many other areas of HOPG surface, the height of these parallelogram-like-structures was also found to be varying between ~14 - 24 nm. It is important to note that the height is
Figure 2. Schematics used to explain the texts in the result section for heights, widths and lengths of parallelogram-like-structures (a); to demonstrate the growth and dissolution directions (b); red arrows for growth and white arrows for dissolution and probable vertical arrangement of LG molecules within a single layer of the parallelogram-like-structure (c).

measured from the lowest point (assuming it to be the HOPG surface) to the highest point on these structures. Some of these structures also found to be overlapping on top of others. The heights of these structures with respect to the underlying structures were also found to be ~14 - 24 nm. Adding to these regular parallelogram-like-structures, irregular 3 dimensional (3D) structures (see white arrow in Figure 1(a)) are also observed. Next interesting observation is their arrangement on HOPG in a triangular fashion thus making angles of either 60° or 120° (see for example, the angles marked in Figure 1(a)). It also observed that some of the structures which grow over the underlying parallelogram-like-structures do not follow this peculiar triangular arrangement (see green arrow heads in Figure 1(a)).

Figure 1(b) which is taken after ~29 min shows considerable morphologica
changes on the HOPG surface. Few of the structures visible in Figure 1(a) disappeared while few others started to grow. For example, the one marked by yellow arrow head in Figure 1(a) dissolved completely, marked by red arrow head dissolved partially whereas the one marked by blue head has grown in Figure 1(b). In other words, dissolution and growth seems to have occurred simultaneously. Further, it is also found that the structures in direct contact with the HOPG substrate are dissolved at a faster rate compared to the ones which are not in direct contact with HOPG (can be viewed better in video 1). Growth was found to be strictly two dimensional in nature at this stage, which means these structures either grew along their length (black arrow in Figure 1(b)) or width (white arrow in Figure 1(b)) by the continuous accumulation of LG molecules. In addition, growth occurs only at the free end of these structures. The term free end refers to the end which is not restricted by the presence of any obstacle. For example, the structure marked by black arrow in Figure 1(b) has grown towards the black arrow whereas at its other end the growth is restricted by the presence of other parallelogram-like-structures. On the other hand, the average heights of remaining or continuously growing structures however remain unchanged at this stage which may indicate that molecules do not dissolve or gets added above/underneath these structures in the initial stage of recrystallization process. A word of caution here, since the dissolution process of some these structures at this stage was much faster than the AFM scanning speed employed, hence the intermediate stages of these processes were not possibly observed. This means it is difficult to draw any conclusion whether dissolution occurred in 2 or 3 dimensions at this stage.

Further topographical changes of the same surface recorded at about 3 hours (for exact timing readers are referred to image captions in Figure 1) are shown in Figure 1(c). Looking at the time interval of >2 hours between Figure 1(b) and Figure 1(c), it is safe to assume that the rate of dissolution and growth process of these structures has decreased considerably with passage of time. However, the recrystallization process has not ceased completely as it is found that the width and length of some of these structures continue to increase (black arrow in Figure 1(c)) and other structures continue to dissolve (yellow arrow in Figure 1(b) dissolved in Figure 1(c)). Even after almost 3 hours of continuous scanning, no change in heights of any of the available structures is observed indicating molecule accumulation/dissolution occurs only along/from length or width. The growth/dissolution process has decreased further and very minute changes are observed after waiting up to ~10 hours (see Figure 1(d)). However, at this stage, one of these structure which was growing as described in previous section, started to dissolve (black arrow head in Figure 1(d)). Taking a closer look revealed that the dissolution process at this stage occurred in a layer by layer fashion (see black arrow in Figure 1(d), also see the video 1 for a better understanding of the process) and hence the height also started to decrease at this point. This layer by layer etching process was not noticed in the earlier dissolution process described in the previous sections. Average height profile of one
such layer (black line in Figure 1(d)) gives a height of ~3.5 nm (see Figure 1(f)). At about 18 hours the growth has completely ceased indicating a saturation point has reached (Figure 1(e)). No further morphological changes were observed even if the experiments were carried out up to 21 hours (data not shown).

To further investigate whether increase in concentration of molecules has any effect on growth behavior, imaging was performed after applying 1mg/ml LG-solution in chloroform on to HOPG surface. A couple of images showing the initial and final stages of recrystallization process for 1 mg/ml LG in chloroform are shown in Figure 3. The entire image sequence as video is supplied as supplementary material (video 2). At higher concentration as expected, more parallelogram-like-structures were observed (Figure 3(a), ~14 min). Most of these structures lying directly on HOPG substrate are arranged in triangular fashion similar to those observed 0.4 mg/ml LG concentration whereas few others not in direct contact with HOPG substrate are oriented in random directions (see for example, black arrow marks in Figure 3(a)). Interestingly, even if number of parallelogram-like-structures are increased considerably at higher concentration however, their widths remain similar to those observed at 0.4 mg/ml concentration i.e., between ~100 - 400 nm. In addition, an increase in irregular three dimensional (3D) structures (marked by white arrow) also observed on top of these parallelogram-like-structures. With passage of time, dissolution/growth of some of these structures (including the 3D irregular structures) started to occur which exactly follow similar mechanism as discussed above for 0.4 mg/ml LG concentration. The recrystallization process is completely ceased at ~15 hours (Figure 3(b), for exact timing please refer to figure caption) and further waiting up to ~17 hours didn’t show any visible change on the surface (figure not shown). This time period of saturation of ~15 hours for 1mg/ml LG concentration is slightly lower than for 0.4 mg/ml LG concentration where saturation occurred at ~18 hours. It is also important to note that there exists some thermal

Figure 3. A couple of AFM images showing initial (a), (after ~14 min) and final (b), (after ~915 min) stages of LG molecules recrystallization on HOPG, after applying 1 mg/ml LG solution (chloroform solvent) to the substrate. The black and white arrows in Figure 3(a) show the parallelogram-like-structures which do not follow the HOPG surface symmetry and irregular 3D structures respectively. Size = 4.9 × 4.9 µm², scan rate = 0.612 Hz, 256 lines/image.
drift for such long measurement time which is clearly visible by the shifting of positions of similar structures in both Figure 1 and Figure 3.

In order to have greater insight about the kinetics of growth along their length and width, the changes in length and width of a couple of such structures for the entire image series shown in video 1 are plotted separately vs. time (see Figure 4). Growth along its length presented in Figure 4(a) is measured along the dotted black line shown in the inset in Figure 4(a). Similarly change in widths plotted in Figure 4(b) is measured along the dotted black lines shown in the inset in Figure 4(b). Utmost care is taken to measure the change in length and width of the structures at the exact location for all the images shown in video 1. In the final stages of the plot several length and width points were skipped (since they represent same length or width) and only points at longer time intervals are plotted. The skipping of individual length and width for intermediate images definitely has no effect on the nature of the plot. These plots clearly demonstrate a logarithmic behavior for increase in length as well as width of these structures which means, the rate is faster in the first couple of hours and slowly decreases until it is completely ceased. Assuming that the structure has a length of 0 nm in the beginning (time = 0 min) and a final length of ~2 µm (time = 175 min), as shown in the plot in Figure 4(a), the average growth rate was calculated to be roughly ~11 nm/minute (scan rate 0.612 Hz, 256 lines/image). The term “at the beginning” refers to the starting point of recrystallization which was not observed in the experiment as it normally takes few minutes for mounting the sample and first image acquisition in AFM experiment which was ~14 minutes in this particular experiment. Following the same principle, the average growth rate for width was measured to be ~1.5 nm/minute. The above measured values are just the average values for the entire process whereas the actual recrystallization process is irregular. This means growth rate is faster in first couple of hours and then slowed down until saturation has reached. It is important to emphasize here that, the main objective of these plots are to show the growth behavior of the structures rather than their error free measurement of individual length/width. From these plots it is also concluded that, length of these structures increased at a faster rate compared to their width.

Similarly, to understand the dissolution behavior of parallelogram-like-structures and their growth kinetics, their change in coverage on the HOPG surface is plotted with respect to time in Figure 5. Commercial Pico image software was used to determine the surface coverage of these structures for all the images shown in video 1. It is possible to separate structures based on their heights using the threshold option in Pico image and this option was used here to differentiate parallelogram-like-structures from the rest of the topography for all the images in video 1. Although a couple of parallelograms also grown with respect to time but their effect while measuring the dissolution coverage is minimal. In other words, it is safe to exclude the effect of grown structures on the overall characteristics of the dissolution curve as the number of parallelograms that got dissolved is much higher than the grown ones. Dissolution of these structures
Figure 4. Plots show increase in length (Figure 4(a)) for the structure marked by dotted black line in Figure 4(a) inset and widths (Figure 4(b)) for the structures marked by dotted black lines in Figure 4(b) inset with respect to time for all the images shown in video 1. Great care was taken to measure the length and widths at the exact location for all the images. A logarithmic change in growth rate both along length and width is observed for these structures.

followed a logarithmic decrease behavior which means dissolution occurred at a must faster rate in the first couple of hours before slowing down considerably reaching saturation point at ~10 hours. During surface coverage measurement
Figure 5. The plot shows the dissolution of parallelogram-like-structures (for all the images in video 1) which also follows a logarithmic behavior. Pico image post processing software’s threshold option was used to determine the individual surface coverage of all the images in video 1 at different time intervals.

Similar plots were also observed for other videos (data not shown). The average dissolution rate in percentage was measured to be 0.078%/min assuming the initial surface coverage (at time \( t = 0 \) min) to be 42.2% and final (saturated) coverage to be 14.2% (at time \( t = 357 \) min).

4. Discussion

Looking at the recrystallization process of these structures, it was observed that their growth occurred only along their length and width but dissolution occurred along all three axes. Both these processes are shown in the schematics in Figure 2(b) where red arrows indicate growth directions and white ones represent dissolution direction. This makes sense, as the free ends along length and width act as preferential nucleation sites for the incoming LG molecules due to their low energy barrier compared to the accumulation of molecules above or below the already formed parallelogram-like-structures which require considerable amount of energy to do so and hence energetically not favorable. Hence the growth followed a two dimensional behavior. During growth faster increase in length compared to width is attributed to the availability of small surface area along length compared to width. Assuming an equal number of molecules travel per minute to get accumulated along length and width, these molecules must distribute themselves equally along the large surface area available across the width (and hence increase in width at a given time is slower) compared to the small surface area across the length (and hence increase in length at a given time is faster). For dissolution, no such energy barrier exists and hence they can occur.
from all three directions which is indeed the case in the observed measurements. A layer height of \( \sim 3.5 \) nm observed during layer by layer dissolution (see Figure 1(d)) process. Assuming the length of LG molecule to be \( \sim 1.8 \) nm (12-carbon chain dodecane has a chain length of 1.8 nm [28]), it is proposed that each dissolved layer is formed initially by the vertical close packing of 2 LG molecules with their length projected vertically (see Figure 2(c)). To understand the orientation of terminal –OH groups a high resolution study is currently underway. Although dissolution occurred from the very beginning of AFM imaging, however, the layer by layer dissolution was observed only after several hours of experiment. This can be attributed to the logarithmic dissolution behavior, which means the faster dissolution processes in the beginning could not be captured with the slow AFM scanning rate. As the dissolution process gets slower, the applied AFM scanning rate became fast enough to capture individual layer dissolution process. Another important aspect of dissolution process is the systematic removal of layers starting from top to bottom in a single parallelogram-like-structure. AFM tip based surface etching [29] could not explain this phenomenon. First, if tip etching is responsible for this process then most of the structures which are grown over the other parallelogram-like-structures would get etched first but in the current case the structures attached directly to HOPG surface got dissolved first. Secondly, AFM tip etching being a slow process, the layer by layer etching would have been observed much earlier during scanning which is clearly not the case in these experiments. In addition, we have also employed tapping mode where lateral force responsible for tip based etching is practically nonexistent. Thus, one hypothesis to explain the dissolution from top layer would be the top layer is less stable due to its one sided binding to the layer underneath compared to other layers which bind themselves on both sides and hence dissolution occurred from top surface. What drives the diffusion of LG molecules to undergo dissolution/growth? Most probably the diffusion is driven by the solvent (chloroform) molecule which is further aided by the temperature. Although most of the chloroform got evaporated within 30 seconds, it is believed that some of the chloroform (solvent) molecules that are trapped between the LG molecules can help in the diffusion process. As the time progressed, these trapped solvent molecules also started to evaporate and hence the growth/ dissolution started to slow down until no further change on the HOPG surface is observed. Factors such as temperature assisted diffusion processes is well documented [30] [31] [32] but it is believed to have played a minor role in these experiments. The reason being with increase in temperature (for example, laser heating of cantilever), the rate of diffusion and hence the growth/dissolution process should increase but exactly the opposite trend i.e., a decrease in growth/dissolution rate with respect to time is observed here. This clearly supports that temperature has minor effect in the crystallization processes presented here.

The next interesting question comes to mind is what happened to the LG mass after it undergoes dissolution? It is safe to exclude that, the material is evapo-
rated from the surface because the flash point of LG molecules is \( \sim 134.3^\circ C \) which is considerably higher than the temperature at which experiments are performed. Just to recall, all the experiments are performed at room temperature (20°C - 24°C). It is also very unlikely that material just jumps outside the scanning area within few minutes of scanning that exact area. So, the only probable explanation would be the LG molecules dissolve to form a thin film on the HOPG surface. If that is the case, then why this parallelogram-thin film conversion process occurred? One probable explanation would be the heteroepitaxial growth on HOPG. To lower their energy, the vertical standing LG molecules will prefer to change their orientation which is aided by the exact carbon-carbon-bond matching between HOPG and LG molecules. It is worth mentioning that molecules similar to LG \( \text{i.e.} \), for 1, 12 dodecanediol, it is already demonstrated by using scanning tunneling microscopy (STM) that the carbon chain lies parallel to the HOPG surface [27]. Once the film started to form, this process is further enhanced by the nucleation theory which states that large islands will grow at the expense of small islands [33] (in this case the parallelogram-like-structures). If the given hypothesis works, then the question arises why some structures also grew simultaneously? This is explained as follows: the areas near these grown structures probably got saturated with a thin film and the incoming material from nearby areas has nowhere to go but to get added to the existing structures. However, this is merely a speculation and no evidence is available at his moment to explain this phenomenon. The ceasing of dissolution observed after several hours can be attributed to the formation of a saturated thin film on the HOPG surface and hence the remaining materials remain as parallelogram-like-structures. It is found that this inter-conversion of parallelogram-like-structures to thin film is a very slow process and often takes hours to reach equilibrium irrespective of concentration. However, this process is concentration dependent as it took less time for 1 mg/ml LG solution to reach saturation point (\( \sim 15 \) hours) compared to 0.4 mg/ml LG solution (\( \sim 18 \) hours). This makes sense, as availability of more number of LG molecules at higher concentration means more number of molecules can diffuse at a given time to form the thin film and hence film saturation can occur comparatively earlier.

The random variation of length of parallelogram-like-structures at the initial stage at different positions in Figure 1(a) can be attributed to the availability of LG molecules in their nearby capture zones when recrystallized from the chloroform solution. On the other hand, the distinct widths between 100 - 400 nm (not combining to give structures of higher width) at the initial stage must be due to the inherent property of their molecular architectures. The maximum width of \( \sim 500 \) nm of such structures at the saturation point must be related to the fact that, once HOPG surface gets saturated with the thin film the incoming LG molecules have nowhere to go but to get accumulated to the existing parallelogram-like-structures even though it is not energetically favorable.

5. Conclusion

To conclude, real time recrystallization of LG molecules was observed on a
HOPG surface for the first time at the molecular level. Recrystallization involved both growth and dissolution of parallelogram-like-structures following a logarithmic behavior. While growth occurred along two dimensions, dissolution was found to be 3 dimensional in nature. Saturation time period for recrystallization was found to be concentration dependent. It is believed, detailed understanding of such processes at the nanoscale on nonpolar substrates like HOPG will act as a model to understand their absorption behavior on skins which are also non-polar in nature and will create curiosity among cosmetologists to explore the absorption behavior of other skin cosmetic constituents by using AFM. Understanding the absorption phenomena of such constituents at the nano level will ultimately help designing more effective cosmetic products in future.

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References


**Appendix**

[https://www.dropbox.com/s/fiof1ivee8c3sgi/Video1.mov?dl=0](https://www.dropbox.com/s/fiof1ivee8c3sgi/Video1.mov?dl=0)

[https://www.dropbox.com/s/rtibad8l64pj37n/Video2.mov?dl=0](https://www.dropbox.com/s/rtibad8l64pj37n/Video2.mov?dl=0)

**Video 1.** LG molecule recrystallization on HOPG from 0.4 mg/ml chloroform LG solution. Only trace images are used for making videos. Size = 4.9 × 4.9 µm, scan rate = 0.612 Hz, 256 lines/image.

**Video 2.** LG molecule recrystallization on HOPG from 1 mg/ml chloroform LG solution. Only trace images are used for making videos. Size = 4.9 × 4.9 µm, scan rate = 0.612 Hz, 256 lines/image.
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