Effect of Recombinant Human Erythropoietin on Survivin Expression in Brain Tissues after Traumatic Brain Injury in Rats

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ABSTRACT

Objective: To explore the regulative effect on Survivin of r-HuEPO after traumatic brain injury (TBI) in rats, and understand the neuroprotection mechanisms of r-HuEPO. Methods: Seventy-eight adult Wistar rats were randomly divided into sham operation group (n = 6), TBI group (n = 36) and r-HuEPO group (n = 36). The experimental TBI model was created by Feeney’s method. Samples were obtained after injury for measuring apoptosis of cells by Epics XL Flow Cytometer. Immunochemical method was performed for inspection of expressions of Survivin and NF-κB proteins. Results: Compared to the sham group, the number of apoptotic cells and Survivin, NF-κB immunopositive cells was significantly increased in the injured brain after TBI (P < 0.01). R-HuEPO significantly increased the expression of survivin and NF-κB, but decreased the apoptotic rates. Conclusion: Increased expression of NF-κB by r-HuEPO may play important role in regulating Survivin level, inhibiting neuronal apoptosis in cortex and exerting protective function to neurons.

Keywords: Recombinant Human Erythropoietin (r-HuEPO); Brain Injury; Survivin; NF-κB; Apoptosis

1. Introduction

Survivin was one of the new-found genes of the apoptosis inhibitor family. The protein expression of Survivin increased significantly after traumatic brain injury (TBI), nevertheless, it does not express in normal differentiated mature tissue. Recombinant human erythropoietin (r-HuEPO) has neuroprotective effect after traumatic brain injury (TBI), however, the specific mechanism of it was still unclear [1,2]. This study is to explore the effect of r-HuEPO on NF-κB, Survivin and cell apoptosis after TBI in rat, in the mean time, illuminate the potential mechanism of neuroprotective effect of r-HuEPO.

2. Materials and Methods

2.1. Main Instruments and Reagents

Epics XL flow cytometer (United States Becman-Coulter), BI-2000 image analysis system (Cheng du Tai meng Technology Co., Ltd.), r-HuEPO (Bei Jing Fourth Ring biopharmaceutical Ltd., Lot # S20020045), NF-κB P65 monoclonal antibody, Survivin polyclonal antibody. Santa Cruz Products, SP Kit and the DAB reagent (purchased from the box in the Shan Jin Qiao Biotechnology Limited in Beijing).

2.2. Experimental Animal and Groups

Healthy male Wistar rats, SPF, weight (260 ± 20) g, (Provided by Faculty of Medicine Centre for Experimental Animals, Gansu provinces, as SCXK (GAN) 2004-007). Animals were randomly divided into sham-operated control group, brain injury group and r-HuEPO treatment group, which administrated with intraperitoneal injection of r-HuEPO (3000 IU/kg) after TBI.

The doses of recombinant human EPO administered in the majority of the animal studies cited are generally in the range of 1000-5000 U/kg, which is over 10 times higher than those currently used clinically in anemic patients with chronic renal failure [13]. In the sham-operated group collected samples one day after injury, other groups were collected samples more than 6 hour and 1, 2, 3, 5, 7 days after injury respectively. Six rats were used at each of the time points.

2.3. Animal Model Method

The experimental TBI model was created by Feeney’s method [3]. Anesthesia was performed with sodium pentobarbital (60 mg/kg) by intraperitoneal injection. The head was fixed in stereotactic apparatus along the midline incision scalp, periosteal stripping exposed to the...
right parietal bone. The born window of 5 mm in diameter was created by using dental drilling in the site of 1.5 mm behind the coronal slit and 2.5 mm beside the midline and the dural matter integrity was kept to. A weight of 40 grams fell freely to hit and make contusion of the right parietal lobe. Then the bone window was closed with the bone wax and scalp incision sutured. According to the sub-time points after injury, the animals were decapitated. In addition to sham-operated group is not against, other steps was the same. In r-HuEPO treatment groups, intraperitoneal injection of r-HuEPO (3000 IU/kg, referenced from clinical administration) was administrated immediately after TBI.

2.4. Immunohistochemical Detection
Expression of Survivin and NF-κB was detected for study, measured with digital image analysis. Injured brain tissue with a complete cortex was selected, and was sliced along the coronal continually with thickness of 10 um after embedding in paraffin conventionally. Observation of Survivin and NF-κB immune response was conducted with optical microscopy, and used BI-2000 image analysis system for quantitative image analysis. Optical density (OD) was used to evaluate Survivin and NF-κB immunohistochemical staining intensity, and compared to the group OD value. We measured the optical density of each immunopositive cell using BI-2000 to calculate a numeration of each candidate genes’ expression, where optical density representing immunopositive cells.

2.5. Measure Neuronal Apoptosis
The fresh injured brain tissue of 100-150 mg was selected, in PBS solution homogenated, tissue homogenated liquid was filtered with 400 holes screen, filtrate was located in centrifuge tube, treated by 1000 r/min for 5 min , washed twice with PBS , 70% ethanol-fixed, left on the condition of 4˚C. Centrifugal supernatant to the next day, washed once with PBS, 1 ml PI dye was plused (concentration of 50 μg/ml, 32.4 ml of normal saline, PI2.5 mg, Rnase 0.5 mg, Triton X-100-0.25 ml, 50 mg sodium citrate, and distilled water to 50 ml), At 4˚C the dark stain was performed for 30 min. Epics XL application of flow cytometry record argon plasma excitation wavelength of 488 nm and red fluorescence, count 10,000 samples each cell to cell apoptosis detection indicators as a percentage.

2.6. Statistics
SPSS10.0 statistical software was used for analysis and the data described with X ± S. One-way ANOVA groups were compared using the least significant difference method for every 2, P < 0.05 for statistical significance.

3. Results
3.1. Changes in the Rates of Neuronal Apoptosis
In all brain injured group the neuronal apoptosis rates were higher than the sham-operated group, reached the peak 2 days after injury, then gradually declined. But the rates still were high until 7 days after injury (P < 0.01). The rates in r-HuEPO treatment group, except for the time points within 6 hours and 7 days after injury, were significantly lower than the control group (P < 0.05) (see Table 1).

The apoptosis rates of TBI group and r-HuEPO treatment group were significantly higher than samples obtained from sham-operation group. * shows statistic analysis acquires P < 0.01 TBI group compare with sham-operation. However, r-HuEPO group expressed a significant different in apoptosis rate, compare with TBI group, which labeled by Δ.

3.2. Expression of Survivin Protein and NF-κB in the Different Groups
Survivin and NF-κB positive staining positioning in the cytoplasm and (or) membrane showed yellow color (see Figures 1 and 2). A small amount of expression of Survivin was observed 3 days after traumatic brain injury, reached at the peak at 7d after traumatic brain injury. Both TBI group and r-HuEPO group samples expressed Survivin at 3d, 5d, 7d, which indicated significantly (P < 0.01), compare to the sham-operation group. Expression of Survivin could be observed 1d after injury in r-HuEPO treatment group, reached peak at 5d. Compared with the brain injury groups 1d, 2d, 3d, 5d after injury, Survivin expression was significantly increased (P < 0.01) (see Figure 3). NF-κB was obviously expressed 6h after injury, peaked at 1d, then decreased gradually, also was significantly higher than that of sham-operated group after 7d (P < 0.01); NF-κB of brain injury in r-HuEPO treatment group at each time point expressed higher significantly than brain injury groups (P < 0.05) (see Figure 4).

Table 1. Apoptosis rates of injured brain of different groups (%).

<table>
<thead>
<tr>
<th>Groups</th>
<th>6h</th>
<th>1d</th>
<th>2d</th>
<th>3d</th>
<th>5d</th>
<th>7d</th>
</tr>
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<tbody>
<tr>
<td>Sham-operation</td>
<td>2.06 ± 0.53</td>
<td></td>
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<tr>
<td>TBI</td>
<td>10.96 ± 1.24*</td>
<td>15.90 ± 1.41*</td>
<td>22.76 ± 1.86*</td>
<td>17.12 ± 1.51*</td>
<td>16.42 ± 1.07*</td>
<td>14.48 ± 0.54*</td>
</tr>
<tr>
<td>r-HuEPO</td>
<td>9.46 ± 1.30</td>
<td>13.00 ± 0.73 Δ</td>
<td>19.14 ± 0.88 Δ</td>
<td>14.78 ± 0.83 Δ</td>
<td>12.78 ± 0.79 Δ</td>
<td>12.34 ± 1.22</td>
</tr>
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Figure 1. Immunohistochemical staining images provide the evidence of the expression of Survivin. Picture A, B, C, D shows Survivin proteins expressed (arrows) in the 1st, 3rd, 5th, 7th day after TBI of samples from TBI group. Picture E, F, G and H illustrate the Survivin expression (arrows) at same time point, which obtained from samples of r-HuEPO group, which revealing r-HuEPO may induct the expression of Survivin.

Figure 2. Expression of NF-κB protein in the different groups, illustrated via immunohistochemical staining. Image A, B, C and D refers to the expression of NF-κB at the 6h, 3rd, 5th and 7th day after TBI (arrows) (E-H). Section E, F, G and H expressed the observation of same time-point with TBI samples which administrated with r-HuEPO (arrows).

Figure 3. The effect of r-HuEPO on expression of Survivin protein at different time points after TBI in rats (Note: *P > 0.05, vs sham group; #P < 0.01, vs sham group; $P < 0.01, vs TBI group; @ > 0.05, vs TBI group).
4. Discussion

Traumatic brain injury is one of the common clinical fatal diseases and very harmful to human health [4,5], and its pathophysiological mechanism is very complicated. Recent studies has given the evidence that the neural cell apoptosis occurred in head injury can promote the secondary brain damage, therefore lead to poor prognosis of the patients [6]. Consequently, the anti-neuronal apoptosis is supposed to be as an important aspect of severe head injury treatment.

Many studies results confirmed that EPO is endogenous cytokines of the central nervous system, and play a neurotrophic and neuroprotective role through neurons and glial cells paracrine effecting on neuron EPO-R. Outside endogenous EPO in a condition of the brain injury can struggle against neuronal apoptosis through penetrating the brain blood barrier and protect the brain from damage [7,8]. Yatsiv, etc. [2] reported that EPO reduced apoptosis and inflammatory response significantly in rat closed brain injury, promoted the restoration of cognitive and move function. Kumral, etc. [9] study found that EPO could reduce apoptosis through inhibiting pro-apoptotic gene expression of bax and DP5, plays a role in protection of the brain. But both home and abroad reported rarely about whether EPO can increase the expression of Survivin neuroprotective. Author explored the mechanism of EPO through the observation the effect of r-HuEPO on nerve cells’ expression of Survivin and NF-κB after traumatic brain injury, provide evidence for clinical application.

Survivin is a new member of inhibitors of apoptosis protein family; it is isolated from the human genome library by effector cell protease receptor-1 cDNA and hybridization screening. It locates on the chromosome 17q25, which contains 142 amino acids. Survivin is not expressed in the normal differentiation of mature tissues. However, it has the high expression in most tumor tissues and after brain injury. Survivin plays the role of cell cycle regulation and anti-apoptotic [1,10]. NF-κB signal transduction pathways is an important signal control system that involved in cell apoptosis, its increase can promote massive anti-Apoptosis factor expression in brain, and participate in gene encoding the transcription of inflammatory molecules after brain injury [11].

This study found that Survivin expression appeared early in the brain after injury in the treatment group with r-HuEPO, and its peak of the expression was earlier than in the control group. Compared to the control group, 1d, 2d, 3d, 5d after injury, Survivin expression levels were significantly increased, and apoptosis rates was significantly decreased. In injury brain side at each time point, NF-κB expression were significantly increased in the treatment group with r-HuEPO than the control group, and kept continuing high level.

Previous work shows that binding of Epo to its receptor, the receptor forms homodimers and undergoes phosphorylation by physically associating and interacting with the tyrosine kinase JAK2 (Janus Kinase-2). Siren also showed that Epo activates JAK2 cascades which activate NF-κB [14]. Therefore, the EPO administration causes the nuclear translocation of NF-κB. This study consist with the previous and in addition, expands the pathway that the translocation of NF-κB and the increasing in NF-κB association with DNA raise up of the synthesis and secretion of the anti-apoptotic factor (Survivin), thereby blocking the process of cell apoptosis, playing a brain Protective role. However, activation of NF-κB can also promote the expression of pro-inflammatory cytokines, promote inflammation process after brain injury, thereby increasing secondary brain damage after the brain injury [12]. On the other hand, r-HuEPO also plays...
an important role in suppression of Caspase-3 expression. Author’s previous study indicates that Caspase-3 expressed within 6 hours of TBI, which directly mediates one of important pathway of neuroapoptosis, however r-HuEPO administration may significantly lower down the level of the Caspase-3 activation [15]. In the mean time, current understanding of r-HuEPO reveals it has strong relationship with variety of pro-inflammatory cytokines. Thereby, further research of r-HuEPO in the mechanism of which regulates and controls the pathway in apoptosis and antiapoptosis will be necessary, to maximize the beneficial role in curing brain injury need further studies.

REFERENCES