Effect of administration of phenylephrine immediately after low dose insemination on pregnancy rates in mares

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ABSTRACT

There is considerable pressure on equine veterinarians to achieve good pregnancy rates with very small doses of semen. Phenylephrine administration in the rabbit increased numbers of oviductal, uterine, and cervical sperm, myometrial contractions, and fertilized ova after low dose insemination. The use of phenylephrine to enhance uterine contractility and fertility has not been investigated in the mare. Thus, the objective of this study was to determine if phenylephrine administration would result in clinically acceptable pregnancy rates in mares bred by low dose insemination. The hypothesis (Ha) was that pregnancy rates would be significantly higher in mares receiving phenylephrine compared to saline controls.

Six pony mares and eight horse mares were enrolled in this study. Mares were inseminated within 24 hours of ovulation with 30 million progressively motile spermatozoa from a single fertile stallion. Immediately following insemination, mares were administered either phenylephrine (0.06 mg/kg) or 1mL of saline via IV injection. Pregnancy status was determined 14 days following ovulation via transrectal ultrasonography. Pregnancy rates in phenylephrine treated mares were 44% (4/9) while 22% (2/9) in saline-treated mares (P > 0.05).

Keywords: Artificial Insemination; Low Dose Insemination; Equine; Phenylephrine

1. INTRODUCTION

Low dose insemination provides the opportunity to increase the number of offspring a valuable stallion can produce in a season, and extends the availability of frozen semen from deceased or infertile stallions. In addition, the development of new advanced reproductive techniques, such as gender selection of equine sperm and the use of density gradients for centrifugation, places pressure on the equine industry to achieve acceptable pregnancy rates with very low numbers of sperm. It is known from work in horses as well as other species that only a fraction of the original inseminate arrives in the oviducts following natural service or standard artificial insemination [1-3]. Therefore, techniques that maximize the percentage of sperm that reach the oviduct may improve pregnancy rates when fewer sperm are used. Techniques that have been previously described in the mare to enhance sperm delivery to the oviduct include rectally guided deep horn insemination and hysteroscopic insemination [4-7]. As both of these are technically demanding and not widely available, there is a need for alternate methods of reducing the total number of sperm in a breeding dose while maintaining acceptable per cycle pregnancy rates. One such method would be the pharmacologic stimulation of sperm transport through enhancement of uterine contractility at the time of breeding.

In the mare, the exogenous administration of both prostaglandin E2 (PGE2) and oxytocin has been investigated to facilitate gamete transport with disappointing or inconsistent results [8,9]. In contrast, administration of alpha adrenergic agonists has resulted in increased fertility after low dose insemination in the rabbit and ewe [10-12].
Phenylephrine, an alpha adrenergic agonist, is relatively selective for α1 receptors over α2 receptors. It is commonly used in animals as a decongestant, mydriatic agent, and to diagnose Horner’s Syndrome. The primary effect of activation of α1 adrenoceptors is to increase smooth muscle tone, corresponding to an increase in intracellular calcium. In rabbit does, phenylephrine has been shown to increase frequency and amplitude of primary uterine contractions and the frequency of secondary contractions [10]. The administration of phenylephrine at the time of breeding resulted in the recovery of increased numbers of spermatozoa from the cervixes, uterus and oviducts of sacrificed does [10]. The same authors demonstrated that phenylephrine administration increased the frequency of uterine contractions in ewes, but did not find improved sperm recovery from the oviducts of sacrificed animals [11,12]. Fertility trials have not been performed in the ewe, but administration of phenylephrine at time of low-dose insemination has been shown to increase recovery of fertilized ova from rabbit does [10]. No previous work has been conducted investigating this aspect of phenylephrine administration in the mare.

Therefore, the objective of this study was to determine whether administration of phenylephrine immediately after insemination would result in clinically acceptable pregnancy rates in mares that had been inseminated with low numbers of sperm. We hypothesized that pregnancy rates would be significantly higher in mares that received phenylephrine compared to those that received a saline control.

2. MATERIALS AND METHODS

Six pony mares and eight Quarter Horse or Thoroughbred mares, as well as one Quarter Horse stallion, were used in this study during the months of June through August. Mares were three to 15 years of age and were housed on pasture at two university-owned farms in central North Carolina. Seven mares were housed at the Teaching Animal Unit in Raleigh NC, and seven mares were housed at the Equine Health Center at Southern Pines. The stallion was maintained in an individual pasture on the grounds of the Equine Health Center at Southern Pines. All procedures were in accordance with North Carolina State University’s Institutional Animal Care and Use Committee’s guidelines for the humane treatment of research animals (IACUC ID#10-048-O). All animals had a complete breeding soundness exam prior to enrollment in the study, including ultrasound exam of the uterus, speculum and digital exam of the vagina and cervix and a uterine biopsy. Mares were cycling regularly and had no evidence of reproductive complications on examination. At the onset of the study, mares were randomly divided into two separate groups, one receiving phenylephrine and one receiving saline. A cross-over study design was used in which treatment groups were switched for subsequent estrous cycles, resulting in a total of 28 cycles in the study. Transrectal palpation and ultrasonographic examinations of the reproductive tract were performed every other day for breeding management. Mares identified in early to mid diestrus were given Prostaglandin F2α (Lutalyse, Pfizer, 5 mg IM) to induce estrus. Late diestral mares were allowed to return to estrus without treatment. When mares were detected in estrus and exhibited a dominant follicle greater than 30 - 35 mm on transrectal ultrasound, 1500 - 2000 international units (IU) of human chorionic gonadotropin (hCG, Chorulon, Intervet Schering Plough Animal Health Corp), depending on body weight, were administered intravenously to induce ovulation. Twenty-four hours later, mares were inseminated with 30 million progressively motile spermatozoa extended in commercial semen extender (INRA 96, IMV Technologies) in the uterine body.

Semen was collected from a single fertile Quarter Horse stallion using a Colorado-model artificial vagina. Immediately following collection, laboratory analysis of raw and extended semen was performed, including photometric measurement of sperm concentration (Densimeter; Animal Reproduction Systems) and measurement of motility and morphology via light microscopy (Figure 1). The fresh semen was extended with INRA 96 (IMV Technologies) up to a maximum volume of 10 mL, with the goal of obtaining a final concentration of 25 million sperm/mL. Each insemination dose contained 30 million progressively motile spermatozoa and was protected from light until insemination. Seven mares housed onsite were inseminated with fresh extended semen, whereas semen was cooled for up to 8 hours in a commercial shipper (Equine Express II Cooled Semen Shipper; Exodus Breeders Corporation) prior to insemination of seven mares located in Raleigh.

Figure 1. Equine semen sample stained with Eosin-Nigrosin Stain for evaluation of morphologic characteristics.
Mares were restrained in stocks with their tails wrapped and tied to the side away from the vulva and perineum for artificial insemination. After removing feces from the rectum, the perineal area was washed with betadine scrub, rinsed with water, and dried with a paper towel to remove fecal contamination. Insemination of all mares was performed by a single individual. Wearing a sterile lubricated glove, an insemination pipette was passed through the vulvar lips into the vaginal vault. The insemination pipette was then guided through the external cervical os and cervix and advanced 1 to 2 cm into the uterine body before depositing the semen. Immediately following insemination, mares were given either phenylephrine (0.06 mg/kg) or 1 mL of saline via intravenous injection. On the following day, mares were examined via transrectal ultrasonography to detect ovulation. If ovulation had not occurred, mares were inseminated and treated a second time as described above. Mares that did not ovulate within 72 hours of the hCG administration or which had evidence of endometritis were excluded from this study. Pregnancy status was determined 14 days following ovulation via transrectal ultrasonographic examination to identify the presence of an embryonic vesicle (Figure 2). At this time, pregnant animals received Prostaglandin F2α (Lutalyse, Pfizer, 5 mg IM) to terminate the pregnancy. A priori analysis using the program GPower 3.1 (G*Power, Düsseldorf, Germany) was performed prior to the onset of the study. Based on previous work with the stallion enrolled in this study and the results of Hawk and coworkers, power analysis indicated the need for 14 animals, examined over 2 cycles. At the conclusion of the study, data were statistically examined for differences between groups, differences between the first and second cycle in the study, differences between farms, and differences between mares inseminated once or twice. The statistical software program Statistix 8.1 (Statistix®, Analytical Software Inc, Tallahassee FL) was utilized for all analyses and data were tested using Wilkoxon Rank Sum tests. Differences were considered significant at $P < 0.05$.

3. RESULTS

Out of a total of 28 possible cycles from 14 mares during the period of the study, 18 cycles qualified for inclusion in the analysis. Three cycles were excluded because the mares failed to ovulate within 72 hours of hCG administration, 4 cycles were excluded because mares ovulated unexpectedly before insemination, and 3 cycles were excluded due to ultrasonographic or cytologic evidence of endometritis. Of the 18 included cycles, the pregnancy rate was 44% (4/9) for phenylephrine treated cycles and 22% (2/9) for the saline-treated cycles ($P = 0.62$; Table 1). No differences were detected in pregnancy rates by location (3 pregnant mares in each location), cycle (4

Table 1. Pregnancy rates in mares after insemination with 30 million spermatozoa and treatment with phenylephrine or saline.

<table>
<thead>
<tr>
<th></th>
<th>Pregnant</th>
<th>Open</th>
<th>Total Estrous Cycles</th>
<th>Pregnancy Rates</th>
</tr>
</thead>
<tbody>
<tr>
<td>TX*</td>
<td>4</td>
<td>5</td>
<td>9</td>
<td>44%</td>
</tr>
<tr>
<td>CT*</td>
<td>2</td>
<td>7</td>
<td>9</td>
<td>22%</td>
</tr>
</tbody>
</table>

*Phenylephrine HCl (0.06mg/kg) injected intravenously immediately after insemination; †1 mL of Saline injected intravenously immediately after insemination.

Figures were detected during the first cycle, 2 during the second), insemination number (3 pregnant mares were inseminated once, 3 were inseminated twice).

4. DISCUSSION

Altering contractility may be a viable means of improving pregnancy rates in mares when suboptimal doses of sperm or compromised sperm are used for insemination. This is supported by studies in rabbits and sheep, which demonstrate that phenylephrine and other uterotonic agents enhance uterine contractility, sperm numbers at the site of fertilization and fertilization rates [10-12]. These studies also demonstrated, however, that any positive effect on pregnancy rates is likely dose-specific. In mares, convincing evidence also exists to suggest that uterine contractility is an important component of sperm transport [13-17]. Artificial insemination alone has been shown to cause an initial increase in the myoelectrical activity of the uterus, resulting in transport of sperm through the female tract [18]. A second increase in myoelectrical activity, beginning 4 hours post breeding and lasting up to 12 hours, results from an inflammatory reaction to the semen and is associated with semen clearance from the reproductive tract [18]. Current research in mares is limited by the difficulty of objectively quantifying uterine contractility in response to insemination or drug administration. Most techniques described
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REFERENCES


**LIST OF ABBREVIATIONS**

mg: milligram
IM: intramuscular
IU: intrauterine
hCG: human chorionic gonadotropin
mL: milliliter