Cough Modulation by Upper Airway Stimuli in Cat—Potential Clinical Application?

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Abstract

The modulation of mechanically induced tracheobronchial cough was tested by applying various stimuli and the elicitation of other airway protective behaviors in pentobarbital anesthetized cats. Capsaicin and histamine were injected in the nose, and mechanical nylon fiber and/or air puff stimulation was applied to the nose and nasopharynx. Reflex responses of cough, sneeze, aspiration reflex and expiration reflex were induced mechanically. Swallow was initiated by the injection of water into oropharynx. Subthreshold mechanical stimulation of nasopharyngeal and nasal mucosa, as well as water stimulation in the oropharynx and larynx, with no motor response, had no effect on rhythmic coughing. Cough responsiveness and excitability increased with capsaicin and air puff stimuli delivered to the nose. Vice versa, the number of cough responses was reduced and cough latency increased when aspiration reflexes (>1) occurred before the cough stimulus or within inter-cough intervals (passive E2 cough phase). The occurrence of swallows increased the cough latency as well. Cough inspiratory and/or expiratory motor drive was enhanced by the occurrence of expiration reflexes, swallows, and sneezes and also by aspiration reflex within the inspiratory phase of cough and by nasal air puff stimuli. Complex central interactions, ordering and sequencing of motor acts from the airways may result in the disruption of cough rhythmic sequence but also in the enhancement of cough. Our data confirm that number of peripheral stimuli and respiratory motor responses significantly alters cough performance. We propose developing and testing stimulation paradigms that modify coughing and could be employed in correcting of inappropriate or excessive coughing.

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1. Introduction

Airway protection and defense comprises the execution and coordination of several behaviors such as cough, apnea, sneeze, swallow, expiration and aspiration reflexes. In healthy airways, coordination of these responses results in the prevention and/or correction of aspiration into the lower airways/lungs [1].

Cough employs air compression and fast expiratory airflows to clean the airways. The effectiveness of cough is based on sufficient amount of air in lungs at the peak of coughing inspiration (I) and on forceful ballistic like activity of expiratory (E) pump muscles to execute the compression and expulsion [1] [2]. During the compression phase the expiratory muscles generate a force against the closed glottis, producing increased intra-airway pressure. The accumulation of energy, in the compressive phase of cough, enhances the expulsive expiration and increases cleaning efficacy of the airways [1].

Hyperfunctional cough (i.e. chronic cough) can have various serious consequences on the cardiovascular and respiratory systems as well as on a quality of life [1] [3]. Hypofunctional cough and/or dystussia also has serious consequences on the ability to keep airways clean with significantly increased chance for aspiration pneumonia and more serious problems [4].

Among the reasons leading to inappropriate cough response there are increased and reduced primary cough afferent drive, inadequate cough modulation by secondary afferent inputs, and various central issues including serious central disorders such as Parkinson’s disease, stroke etc. [5] [6]. Understanding of cough modulators may allow their employment in corrections of excessive or insufficient cough response improving pathological conditions of patients.

This paper summarizes and sorts our previous data, brings new results to light, and provides a theoretical base for pre-clinical studies targeting correction of inappropriate cough response in patients. We hypothesize that several procedures involving the stimulation of afferents additional to cough primary pathway may have potential to improve inappropriate cough performance in humans.

2. Methods

All our experiments were conducted on cats. Detailed description of experimental procedures has been provided in our previous reports [7]-[11]. Animal care as well as all procedures were performed in accordance with the Animal Welfare Guidelines of the Comenius University and the legislation for animal use and welfare of Slovak Republic and European Union (Directive 2010/63/UE). All protocols were approved by Ethic Commitee of Jessenius Faculty of Medicine.

Briefly, the cats (n = 101, from eight series of experiments) were anesthetized by pentobarbital sodium (Spofa, Slovak Republic; Morbital, Polfa; Pfannenschmidt GmbH, Germany) and allowed to breath spontaneously (frequently, depending on pO2, an oxygen enriched air was delivered). Cough was induced by mechanical probing of the trachea down to the carina by a nylon fiber or a soft polyethylene catheter that had been moved back and forth and rotated (1/s). Sneezing was induced by gentle punctate mechanical stimulation of the nasal septum with an approximately 1.5 cm long soft nylon fiber. Aspiration reflex (AspR) was induced mechanically (soft nylon loop or air puff) in the nasopharynx via nasal approach or pharyngostomy. Expiration reflex (ExpR) was induced by nylon fiber loop or soft polyethylene catheter probing to the glottis (inferior vocal folds). Swallow was induced by the instillation of water into the oropharyngeal region (Figure 1). Control cough trials were performed before and after stimulation protocol with additional stimuli applied.

The cough reflex was defined by a large augmenting burst of diaphragm and/or parasternal EMG activity immediately followed (and partially overlapped) by a burst of expiratory abdominal EMG activity [1] [10] [12] with corresponding inspiratory-expiratory (I-E) oscillations of esophageal pressure (EP) as noted in Figure 2. Sneeze was distinguished from the cough reflex by activation of the styloglossus muscle during the expulsive phase, which matches the occurrence of ballistic-like abdominal discharge [10] [13].

Capsaicin (25 µl, 50 µM, Sigma, dissolved in ethanol and diluted in buffered saline) and histamine (25 µl, 16
Figure 1. Schematics of stimulations, related stimulation sites and induced airway motor behaviors. Capsaicin initially inducing the sneeze reflex and histamine were injected to the nose, air-puff pressure pulses directed from the nasopharynx to the nose and mechanical continuous stimulations inducing and not-inducing the sneeze reflex were applied in the nose. Mechanical tactile stimulation and air puff (some at subthreshold pressure with no motor response) eliciting aspiration reflex were delivered to the nasopharynx. Injection of water in oropharynx elicited swallow. Presence of water in pharynx and larynx was without motor response. Individual tactile mechanical stimuli to the glottis induced expiration reflex. Cough was produced by repetitive mechanical stimulation in the lower trachea.

Figure 2. Water in oropharynx (water) induced swallows (depicted by a burst of geniohyoid muscle activity—GH and slight depression immediately followed by a burst of cricopharyngeus muscle = upper esophageal sphincter activity—UES) resulted in prolonged inter-cough intervals (latency to the following cough response) and increased cough motor drives. The number of cough efforts, however, was not significantly altered. Int, integrated activity (moving average); EP, esophageal pressure; ABD, abdominal muscle activity; PS, parasternal muscle activity.

mM, Biosynth, Riedel de Haen AG, Germany, diluted in buffered saline) challenges were performed by the instillation of solution into one nostril (lasting approximately 3 s) via a thin plastic catheter inserted through the nose 7 - 8 mm from the nares (Figure 1). Tracheal stimulation began 10 - 15 s after administration of capsaicin or histamine in the nose. Nasal mucosa was cleaned with saline after completion of cough trial. The nasal air puff stimuli were delivered via catheter directed towards nasal cavity from the nasopharynx.
Arterial blood pressure, respiratory rate, end-tidal CO₂ concentration (ETCO₂) and body temperature were monitored and maintained within physiological range for each of the experimental conditions.

3. Results

In all protocols, including those with additional probing of airways, mechanical stimulation of the trachea/bronchi reliably elicited repetitive cough. With the addition of mechanical stimulation, which resulted in a behavioral response (i.e. sneeze or AspR) and/or irritant stimulation to the nose, cough was significantly modulated [7]-[11]. Of note, sub-threshold mechanical stimulation either to the nasopharynx [7] [8] or nose (continuous stimulations) [10] or pharynx had no effect on any analyzed cough characteristics (Table 1).

Cough responsiveness (number of efforts/stimulation time) representing cough excitability was increased with the addition of capsaicin [11] or air puff stimulation to the nose (Table 1). Sneeze was elicited during the initial capsaicin administration to the nose in 77% (10/13) of cats [11], but the air puff did not result in any elicited behavior. Of note, cough was only induced following the initial capsaicin adaptation period (i.e. sneeze no longer elicited) [11].

Cough responsiveness was reduced when AspR (17.2 ± 2.4) occurred just prior to cough or AspR (3.3 per period) were induced within inter-cough intervals (quiescent cough E2 phase) [7]. The effects correspond to prolonged latency to the first and subsequent coughs (prolongation of cough quiescent E2 phase duration; Table 1) [7]. E2 phase duration was also prolonged with the presence of swallow, however, with swallow there was no effect on number of coughs (Table 1; Figure 2) [9].

Table 1. Summary of cough modulation by additional afferent input or behavior induction.

<table>
<thead>
<tr>
<th>Stim/event</th>
<th>CN</th>
<th>latency</th>
<th>I drive</th>
<th>I EP</th>
<th>E drive</th>
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<tr>
<td>nas mech</td>
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<td>nas Sn</td>
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<td>NS</td>
<td>NS</td>
<td>+84%*</td>
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<tr>
<td>nas caps</td>
<td>+17%*</td>
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<td>NS</td>
<td>+84%*</td>
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<td>nas hist</td>
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<tr>
<td>nas puff</td>
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<tr>
<td>AspR before</td>
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<td>+250%*</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
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<tr>
<td>AspR E2</td>
<td>-50%**</td>
<td>+183%*</td>
<td>NS</td>
<td>NS</td>
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<td>NS</td>
</tr>
<tr>
<td>AspR I</td>
<td>NS</td>
<td>NS</td>
<td>+72%**</td>
<td>+63%**</td>
<td>+44%*</td>
<td>+38%*</td>
</tr>
<tr>
<td>AspR E</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
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</tr>
<tr>
<td>ExpR E2</td>
<td>NS</td>
<td>NS</td>
<td>+76%**</td>
<td>+45%**</td>
<td>+199%*</td>
<td>+66%**</td>
</tr>
<tr>
<td>ExpR I</td>
<td>NS</td>
<td>-</td>
<td>+74%**</td>
<td>+29%*</td>
<td>+148%*</td>
<td>+108%**</td>
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<tr>
<td>ExpR E</td>
<td>NS</td>
<td>-</td>
<td>+62%*</td>
<td>+32%**</td>
<td>+275%*</td>
<td>+145%**</td>
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<td>orph Sw</td>
<td>NS</td>
<td>107%*</td>
<td>+22%*</td>
<td>+44%***</td>
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</table>

CN—cough number; latency—the duration from the stimulus to the beginning of the first cough, alternatively (for AspR E2 and orph Sw) inter-cough intervals = quiescent E2 phase of coughing; I drive—the diaphragm or parasternal muscle (for orph Sw) EMG moving average maximum; I EP—peak inspiratory esophageal pressure; E drive—the abdominal muscle EMG moving average maximum; E EP—peak expiratory esophageal pressure; nas-mech—nasal mechanical stimulation not inducing motor response (sneeze); nas Sn—nasal mechanical stimulation that induced sneeze reflex; nas caps—nasal stimulation with capsaicin; nas hist—nasal stimulation with histamin; nas puff—nasal stimulation with air puff directed outward (expiratory); nph puff—nasopharyngeal air puff stimuli not inducing aspiration reflex; AspR—aspiration reflex; before—before cough trial begun; E2 during quiescent E2 phase of cough; I—during cough inspiration; E—during inspiratory-expiratory transition or active E1 cough phase; ph+lar w—water stimulus within the pharynx and/or larynx when no swallow and/or expiration reflex has been induced; orph Sw—water stimulus in the oropharynx that induced swallow reflex; *p < 0.05; **p < 0.01; ***p < 0.001 compared to control coughing.
During combined trials swallow, sneeze and cough (rhythmic behaviors) always occurred in an ordered sequence without significant overlapping of their motor activities [9] [10]. During sneeze/cough trials each behavior is either cough or sneeze [10] and they would be grouped together (i.e. several sneezes in a row followed by several coughs in a row). Swallow occurred (95%) during the quiescent E2 phase of cough (termed phase restriction) [9]. Swallow also enhanced subsequent cough I and E components (Figure 2) [9]. Co-occurrence of cough and sneeze resulted in increased cough E component (I and E component of sneeze) [10] and nasal air puff stimuli increased cough E, as well.

AspR and ExpR (short and immediate non-rhythmic responses) can occur during any phase of cough. Figure 3 demonstrates AspR transient inhibition of cough E with no change to the spatio-temporal cough feature [8]. However, AspR in the I phase of cough [8] and ExpR in any position (Figure 4) during coughing improved both I and E component of cough.

When number of cough was reduced due to applied stimuli, prolonged latency, quiescent E2 cough phase and total cough cycle durations were observed. The only altered active cough time interval was the cough expiratory period when AspR were induced during cough I [8].

4. Discussion

The cough reflex is an important mechanism of airways defense [1] [4] [12]. Cough’s physiological significance is reinforced by its constancy under various and antagonistic conditions. Additionally, the present experimental data demonstrated that nasal stimulation, AspR, ExpR, sneezing and swallowing are able to enhance many parameters of cough performance [7]-[11] [14] [15]. These robust results prompt discussion of two major issues. First, potential mechanisms of cough modulation. These mechanisms could help to explain cough irregularities in many patient populations and allow for pharmacological interventions. Second, the use of these “experimental
paradigms” as therapeutic interventions where modulations might be an effective course of treatment for cough irregularities [14] [15].

Our laboratories systematically analyzed cough under various conditions including: stimulations of additional afferent pathways and initiation of additional motor behaviors during coughing on experimental animals over decades. Based on the high number of reported experimental data, we hypothesize two functional levels for cough control [16] [17]. First, the cough motor pattern is generated by the central pattern generator (CPG); and second, the occurrence, ordering and appropriate placement of the motor act (including the inhibition of any other behavior) is provided by the behavioral control assembly [16]. According to this theory the subthreshold stimuli did not pass the “decision making filter” and were not effective in modulating cough excitability. As noted in Table 1, with the exception of the nasal air puff, all subthreshold stimulation was not able to significantly modulate the measured cough parameters.

Additionally, we hypothesize that the mechanism of cough modulation, by other simultaneously executed behaviors, is alterations in synaptic drive at the level of the CPG. The neuronal network involved in cough production extends over multiple areas of brainstem [18] [19]. High number of these neurons are multifunctional (i.e. they participate in multiple behaviors) [20] [21]. Thus, there is a high probability that the execution of one motor behavior (e.g. swallow) directly shares excitability with elements of cough network [22] [23].

AspR and ExpR are able to overlay onto the cough motor pattern (Figure 3). The mechanism for how this is possible remains unknown. Interestingly, AspR within cough E1 phase was able to temporarily interrupt the active expiratory phase (Figure 3), but it did not change EMG amplitude or phase durations [8]. AspRs (>1) within passive expiratory phase (E2) or pre-cough reduced cough responsiveness but also did not alter the cough motor pattern. On the other hand, AspRs during the inspiratory phase of cough enhanced cough I and E components. We hypothesized that higher volume related feedback (i.e. pulmonary stretch receptor activation) can increase the excitability for cough [8]. However, there is no relationship of volume-timing during cough [24], there are limited correlations between I and E efforts/durations during cough [25], and the reduction of volume feedback increased I and E drive [26].

The up regulatory effect of ExpR on cough, regardless of cough phase, likely reflects increases in primary cough afferent drive [1]. It is consistent with increased cough response when higher intensity of stimulation (higher drive from receptors), larger area of stimulation, and multiple cough inducing sites including laryngeal and tracheal cough inducing areas are stimulated [27].

Capsaicin in the nose was able to modulate cough, while histamine did not have any significant effects [11]. This is likely due to the distribution of receptors and ion channels on related nasal sensory cells, target population of nasal sensory afferents and their connectivity to higher order neurons. Capsaicin, the agonist of TRPV1 receptors, is a powerful tussive agent. It induces cough responses from the larynx and tracheobronchial tree, and sneeze from the nasal mucosa [11] [28]. We expected little modulatory effect on cough from the sneeze responses that occurred immediately after application, because nasal capsaicin increased CN while sneeze reflexes during coughing increased cough expiratory efforts (Table 1) [11]. Histamine receptors H1-H4 are widely expressed in the nasal mucosa, however our experiments failed to show any significant modulation of cough response in cats. However, application of histamine to the nose of volunteers with allergic rhinitis enhances cough sensitivity. There are very likely inter-species differences in the response induced by histamine. Moreover, the anesthesia could cause the lack of the response in anaesthetized state in comparison with conscious subjects [29] [30].

The most significant feature of cough reduction was a longer latency to the first and/or subsequent coughs in the sequence. Increasing cough latency may represent practical technique for cough reduction in patient populations. Additionally, increased cough motor drive related to combinations of cough with swallow/sneeze could compensate for the trend of lowering cough number [15]. Such “compensation” could help increase airway protection by maintaining clear pharynx, while not recruiting cough [9] [17] [30]. This could represent behavioral target to reduce the sensation of needing to cough, without centrally suppressing the cough pattern generator with pharmaceutical agents.

There are other parallel afferent inputs and pathways modulating coughing besides those we summarized in this paper [31] [32]. Upper airways inflammatory diseases and gastro-esophageal reflux disease can up regulate cough [33] [34], while bronchopulmonary C-fiber activation [35], BP alterations [36] and perhaps irregular cough volumes stretch receptor activation [26] can decrease cough excitability and/or cough motor drive. We focused on those, which are easily applicable and inducible due to their potential employment for cough modula-
lation in correction of excessive or insufficient coughing.

Number of peripheral stimuli and motor responses from the airways significantly alters cough performance. Complex central interactions, ordering and sequencing of motor acts may result in the disruption of cough rhythmic sequence but also in the enhancement of cough. We propose developing and testing stimulation paradigms that modify coughing and could be employed in correcting of inappropriate or excessive coughing.

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