Natural killer cells responsiveness to physical exercise: A brief review

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ABSTRACT
Natural killer cells (NK) are a group of peripheral blood lymphocytes which display cytotoxic activity against a wide range of tumour cells. They are a consistent part of the inflammatory response that is activated when either internal or external injuries occur as they are able to synthesize perforins. An important role is played by NK cells in the host defence against tumours without expressing any antigen-binding receptor in their membrane which, however, distinguish T and B lymphocytes. NK activity appears early in the immune response, thus providing immediate protection during the time required for the activation and proliferation of cytotoxic T lymphocytes and for their differentiation into functional cells. Even though much research regarding the effects of aerobic training exercise on NK cell numbers and function, there appears to be much controversy regarding its effect. NK cells are rapidly mobilized into circulation in response to acute exercise, most likely by increased shear stress and catecholamine-induced down-regulation of adhesion molecule expression. However, tissue injury and inflammation which often accompanies strenuous exercise have been associated to post-exercise NK cell suppression. Scientific evidence indicates exercise-induced changes in NK cell redistribution and function should be strongly influenced by stress hormones including catecholamines, cortisol and prolactin as well as by soluble mediators such as cytokines and prostaglandins. The role of exercise therapy in cancer patients and survivors rehabilitation is becoming increasingly important as it is thought to modulate immunity and inflammation. However, more knowledge about the effects of exercise on immune function in these patients is needed.

Keywords: NK Cells; Aerobic Exercise; Anaerobic Exercise; Catecholamines; Prolactin

1. INTRODUCTION
Natural killer cells (NK) were first described in the 1970s as a group of peripheral blood lymphocytes (5% - 10%) which display cytotoxic activity against a wide range of tumour cells [1]. Thus, NK cells can be considered as part of the inflammatory response that is activated when either internal or external occur, since they are able to synthesize perforins [2], a family of pore-forming proteins of six esterases called granzymes A-F. NK cells play an important role in the host defence against tumours without expressing any antigen-receptor (i.e. the CD3 receptors). Therefore, NK cells lack the attribute of immunologic specificity and memory. The cytolytic activity of NK cells is enhanced by interferon (IFN)-gamma [3] and IL-2 [4,5] whereas prostaglandin E-2 down-regulates the function of NK cells [6]. In humans, Chédiak-Higashi syndrome is associated with an absence of NK cells [7] and an increased incidence of lymphomas. NK cells play an important role in the host defence against tumours without expressing any antigen-receptor in their membrane which, however, distinguish T and B lymphocytes (i.e. the CD3 receptors). Therefore, NK cells lack the attribute of immunologic specificity and memory. The cytolytic activity of NK cells is enhanced by interferon (IFN)-gamma [3] and IL-2 [4,5] whereas prostaglandin E-2 down-regulates the function of NK cells [6]. In humans, Chédiak-Higashi syndrome is associated with an absence of NK cells [7] and an increased incidence of lymphomas. NK cells are also able to lyse some virus-infected target cells [8]. In fact, this NK activity appears early in the immune response, thus providing immediate protection during the time required for the activation and proliferation of cytotoxic T lymphocytes and for their differentiation into functional cells. A case study has been described in which a young woman lacking NK cells suffered a life-threatening infection...
with cytomegalovirus [9]. NK cells express the p75 kD component of the IL-2 receptor. The IL-2 molecules enable activated NK cells to progress from G1-phase into the proliferative phase of the cell cycle. These cytokines stimulate antiviral activity in IL-2 activated NK cells, and this activation plays an important role in host defence during the first few days of many viral infections until a specific cytotoxic lymphocyte cell response develops. Human NK cells can be subdivided into different populations based on the relative expression of the surface markers CD16 and CD56. The two major subsets are CD56 (bright) CD16 (dim/−) and CD56 (dim) CD16 (+), respectively. The CD56 (bright) NK cells are numerically in the minority in peripheral blood but constitute the majority of NK cells in secondary lymphoid tissues. They are abundant cytokine producers but are only weakly cytotoxic before activation. Recent [10] data suggest that under certain conditions, they possess immunoregulatory properties, and that they are probably immediate precursors of CD56 (dim) NK cells. CD56 (bright) NK cell percentages are expanded or reduced in a certain number of diseases, but the significance of these variations is not yet clear. The cytotoxic activity of CD56 (dim) NK cells is significantly higher than that of CD56 (bright) cells and they contain much more perforin, granzymes and cytolytic granules [11]. The high expression level of CD16 makes them efficient mediators of antibody-dependent cellular cytotoxicity, whereas CD56 (bright) CD16 (dim) NK cells perform antibody-dependent cellular cytotoxicity only weakly and CD56 (bright) CD16−NK cells of course not at all [12]. Upon stimulation with cytokines such as IL-2 or IL-12, the cytotoxic activity of all NK cell subsets dramatically increases [13].

2. IMMUNE FUNCTION AND PHYSICAL EXERCISE

2.1. Generalities

Experimental and clinical evidence indicates that high intensity, long duration physical training is associated with adverse effects on immune function [14,15]. On the contrary, it has been proposed [16] that training schedules with moderate exercise may lower the risk of upper respiratory tract infection (URTI). As pointed out in the "Position statement Part one: Immune function and exercise", by Walsh et al. [17], the hypothesis of a U-shaped relationship between physical activity and resistance to disease makes intuitive sense, in accordance with the general belief that although regular, moderate doses of physical activity have beneficial effects on health, excessive amounts or intensities of physical activity have negative consequences. In the case of the immune system, Shek and Shephard [18] suggested that an excess of physical activity could induce an analogous to clinical sepsis, with tissue destruction from an excessive inflammatory reaction. Although initially conceived simply in the context of URTI [19], the concept of a U-shaped response has now been extended to cover the effects of physical activity upon a variety of clinical disturbances of immune function. In terms of cancer prevention and therapy, Thune and Furberg [20] utilized the metabolic exercise training (MET = 3.5 ml O2 per kg body weight per min) to estimate the energy cost of physical activities and suggested moderate activity (>4.5 MET) more than light activities (<4.5 MET) as a good physical activity to protect against overall cancer risk. Moreover, Mooren et al. [21] found that excessive exercise could cause apoptosis and Neubauer et al. [22] showed DNA damage in the same exercise condition. Certain autoimmune conditions also respond to carefully regulated physical activity programmes. In fact, Baslund et al. [23] showed a good effect of 8 wks of bicycle training on the immune system of patients with rheumatoid arthritis, although they pointed out that it would need to be clearly established whether benefit occurs through direct modulation of cell counts and cytokines, or through changes in the activity of transcription factors for pro-inflammatory cytokines. In general, increased levels of T and B lymphocyte subclasses have been found during acute exercise [14,24,25], while controversial data have been obtained in the activity of NK cells since they have been found to increase during exercise [26], subsequently decreasing at the end of the effort [27], remain unchanged [28,29], or be reduced [30]. A meta-analysis of 94 studies evaluating NK cell counts showed that there is a marked increase at the end of exercise, and a catecholamin-mediated demargination of cells has been considered as being responsible for this occurrence [31]. However, post-exercise down regulation in NK cell numbers and activity was seen frequently and this was attributed either to monocyte/granulocyte-induced rises in plasma prostaglandins and/or to cortisol and prolactin increasing blood levels.

2.2. NK Cells Responsiveness to Prolonged Exercise

Despite much research regarding the effects of exercise aerobic training on NK cell numbers and function [17], there appears to be much controversy regarding its effect. Fairey et al. [32] found increased natural killer cell cytotoxic (NKCC) activity with respect to non-training subjects in a large trial (25 females) of postmenopausal breast cancer survivors who trained on a cycle ergometer three times per week for 15 wks. On the contrary, Campbell et al. [33] investigated the effect of 12-month aerobic exercise, relative to stretching control,
on \textit{in vitro} immune function in a randomized, controlled trial of 115 post-menopausal, overweight, or obese sedentary women demonstrating that NKCC activity did not differ between groups. Hence, they concluded that the controlled trial showed no effect of aerobic exercise on \textit{in vitro} immune function, despite excellent retention, high adherence, and demonstrable efficacy of the exercise intervention. However, intense training has been shown to alter NK cell subsets and reduce NKCC [34,35]. Despite studies in animals having demonstrated that regular exercise can increase \textit{in vivo} NK cell cytotoxicity [36,37], it has been found that the specific contribution of these cells in mediating this exercise effect is still unclear [38].

2.3. NK Cells Responsiveness to Acute Exercise

There are many papers concerning the acute effects of exercise on circulating NK (CD3−CD16+CD56+) cells, and this due to their ease of study and large magnitude change in response to exercise [17]. NK cells are rapidly mobilized into the circulation in response to acute exercise, most likely by increased shear stress and catecholamine-induced down-regulation of adhesion molecule expression [39]. Hoffman-Goetz [40] found that during a few minutes of intense exercise the number of NK cells in peripheral blood results increased, and Shephard and Shek showed that the intensity rather than the duration of exercise was responsible for the rise in cell numbers [31]. As shown in a study by Gannon \textit{et al}. [41], NK cell cytotoxicity is a major functional measure of NK activity. They demonstrated that unstimulated NKCC was dependent on the intensity and duration of the exercise bout since, immediately after a single bout of moderate or exhaustive exercise, there is a 50% - 100% increase in human peripheral blood NKCC, and this concerns both young and old people [42]. The exercise-induced increase in NKCC is largely due to an increase in the absolute number and percentage of blood NK cells. However, NKCC expressed on a per cell basis does not appear to change much after acute exercise unless the bout is intense and prolonged, in which case NKCC can be depressed for several hours, possibly indicating an enhanced period of susceptibility to infection [43]. There appears to be a differential mobilization such that CD56 (bright) NK cells are less sensitive than CD56 (dim) [17]. This might indicate a reduced ability to defend against pathogens during acute exercise, as CD56 (bright) cells are more cytotoxic. However, the health significance of exercise induced changes in circulating NK cell subsets, like other leukocytes, has not been completely clarified [39]. Concerning the extent to which short-lasting, high-intensity exercise alters NK cell subsets, Suzui and colleagues [44] reported on the effects of brief, incremental exercise on these cell subsets. Nine males exercised on a cycle ergometer for 5 min at each of 4 increasing intensities (50%, 90%, 120%, and 140% of their individual ventilatory threshold), with blood samples drawn after every workload. The authors found that only the proportion of CD56 (dim) cells increased in response to exercise; the proportion of CD56 (bright) cells in the peripheral circulation did not change. However, because of an overall leukocytosis both CD56 (bright) and CD56 (dim) cell counts increased with increasing exercise intensity. In a successive study [45] the same authors confirmed their earlier findings by showing that in 6 males cycling for 30 min at 120% of their individual ventilatory threshold (~70% VO$_2\text{max}$) the proportion of CD56 (dim) cells, but not the proportion of CD56 (bright) cells, increased significantly. Based on these two studies, one can conclude that the redistribution of CD56 (bright) cells appears to be resistant to changes in exercise intensity. Almost all the experiments in which the behaviour of lymphocyte subclasses was observed during acute exercise concerned prolonged (60 min or more), sub-maximal muscle effort [46-48]. On the other hand, a significant increase in NK cells was seen about sixteen minutes after a head-up tilt test [49] and only about three minutes after the beginning of a physical exercise or a time delay from the evoking stimuli that is too short [50]. However, it could be of interest to understand the relationships among relative levels of reached work rates at each step of an incremental exercise, and the corresponding changes in both absolute NK cell quantity and NK cell percentage in the peripheral blood. The experiment by Del Giacco \textit{et al}. [51] provides much information on the above requirement. They evaluated the time course of NK cell numbers and relative concentration in peripheral blood at three points: pre-exercise, work-rate corresponding to oxygen consumption at anaerobic threshold (VO$_2$-AT), work-rate at maximum oxygen consumption (VO$_2$-max), in 8 healthy male subjects who performed a progressive cycle ergometer exercise up to exhaustion. It was observed that while VO$_2$ progressively increased up to VO$_2$-max, which was 28% higher than at the VO$_2$-AT, both NK cell numbers and relative concentration also increased progressively to reach the work-rate corresponding to VO$_2$-AT, but when the work-rate increased further up to its maximum, in correspondence of VO$_2$-max, the increases of absolute and relative NK cell values were only 9% or a non-statistically significant variation. They concluded that when the contribution from anaerobic sources for exercising muscles became relevant (\textit{i.e.} in correspondence of VO$_2$-AT) and progressively increased to reach the maximum workload, NK cells ceased to increase. A mismatch between the pro-
gressively increasing physical stress, occurring from VO₂AT to VO₂max, and the corresponding NK-dependent defence against micro-organisms should have happened when the exercise work rate overcame that corresponding to the VO₂AT [15, 31]. In the same study, to better investigate a possible inhibitory effect of supra-maximal exercise on the exercise-induced NK cells proliferation, Del Giacco and colleagues supposed that if the heavy exercise, whose strength was indicated by the value of the excess in the expired carbon dioxide (CO₂-exc) [52], induced NK cell suppression, then the increase in the percentage of these cells at VO₂max, with respect to VO₂AT, ought to be as lower as the higher was CO₂-exc.
To verify this hypothesis they tested the following general equation for a quadratic regression:
\[ NK = a + bCO₂_{exc} + cCO₂_{exc}^2 \]  (1)
where NK, which was expressed, in turn, as absolute (mm³) or percentage (%), represents the difference of these cell values at VO₂max and at VO₂AT, respectively. Applying the Eq. (1) to their data, del Giacco et al. obtained parabolas with parameter (b) which was a negative number and parameter (c) which was a positive number, or by one arm descending followed by the other arm ascending. This means that the higher the CO₂-exc at the end of the progressive exercise the lower the increase of the NK cells concentration at this point with respect to the value observed at VO₂AT. However, when CO₂-exc reached at VO₂max was very high, the direction of this relationship was inverted. The authors concluded that the CO₂-exc value corresponding to the parabolas vertex marks the transition at which NK cell inhibitory mechanisms stopped their prevalence on activator mechanisms. It must be considered that tissue injury and inflammation which often accompanies strenuous exercise have been associated to post-exercise NK cell suppression [53]. However, since it has been found that certain indices of oxidative damage to skeletal muscle appeared only at the end of heavy exercise [54, 55], it may be that a precocious down regulation of NK cells, due to these exercise induced muscle injuries, should be reached before the cessation of a strenuous, anaerobic exercise when lactate flow from the working muscles to blood becomes consistent (i.e. CO₂-exc increases). If this happens, an unprotected window, lasting the time in which the subject works over the anaerobic threshold, ought to be opened during which the athlete becomes susceptible to viral infections.

3. ENDOCRINE CONTROL OF NK CELLS ACTIVITY DURING EXERCISE

3.1. Generalities

Responses to stressful events, as is the physical exercise performed at work rates up the anaerobic threshold, are generally regarded as reactions of the organism to accommodate to or compensate for stress. This reaction is classically described as an activation of the symphato-adrenal system and the hypothalamic-pituitary-adrenocortical axis. Recently, in men who cycled at 40, 60, 80 and 100% of their power output at VO₂max in successive time blocks of 10 minutes each up to exhaustion, De Vries et al. [56] showed that increases in catecholamines reflected the relative work load, increases in prolactin occurred only after exercise reached an intensity of 80% VO₂max whereas increases in cortisol were found just after exhaustion. These results suggest that activation of stress hormones occurs at different time points, supporting the notion that these hormones have different roles in preparing the organism for physical activity and recovery, i.e. workload-related adaptation on the one hand and protection against disturbed homeostasis and leukocytes redistribution in the circulation on the other. When physical exercise reaches a workload exceeding that corresponding to the anaerobic threshold, it typically results in muscle soreness and injury, and inflammatory response to the muscle injury is initiated, characterized by the movement of fluid, plasma proteins, and leukocytes, including NK cells, into the injured area [46]. An exaggerated response in the inflammatory cascade, which could compromise the actual request of mechanical work from exercising muscles, is prevented via several pathways including the activation of the hypothalamic-pituitary system which exerts a strong antiinflammatory action [57]. Nevertheless, if anaerobic stress becomes too high, i.e. the CO₂-exc at VO₂max reaches very high values, NK cell concentration achieved at the maximum work-rate could tend to slow its descent with respect to the VO₂AT value and, in agreement to the Eq. (1), it starts to increase again [17]. In light of the opposite pro-inflammatory/anti-inflammatory mechanisms which take place when strenuous exercise is performed, such a pro-inflammatory/protective mechanism against infection could gradually prevail against the antiinflammatory/performance-preserving mechanism when the subject’s anaerobic power becomes too high. A lot of scientific evidence indicates exercise-induced changes in NK cell redistribution and function ought to be strongly influenced by stress hormones including catecholamines, cortisol and prolactin as well as by soluble mediators such as cytokines and prostaglandins.

3.2. Catecholamines

Catecholamine effects on NK cell activity and redistribution have been extensively studied [36, 58-60], and it has been found that NK cells express both adrenergic beta-1 and beta-2 receptors in their membrane [61].
However, mainly beta 2 subtype are found on the membrane of these cells [36,59]. Klokker et al. [60] found that both beta 1 and beta 2 adrenergic receptor blockade suppress NK cell increase due to head-up tilt. The effects of catecholamines could include the recruitment of NK cells from the spleen or other sites to the circulating pool due to the reduced adhesion of NK cells to endothelial cells [36,62]. In fact, local infiltration of epinephrine to reduce bleeding in patients undergoing a surgical intervention under general anaesthesia produced an instantaneous increase in both NK cell circulating number and activity [63]. In rats The number of blood NK cells doubled within 10 min of metaproterenol administration and returned to baseline levels within 1 h. By this time, metaproterenol suppressed blood NK activity in a dose-dependent manner. Two beta-adrenergic antagonists, propranolol, which crosses the blood-brain barrier, and nalidol, which does not, blocked this suppression [64]. Considering humans, plasma catecholamines were determined in peripheral blood of subjects resting in supine position by Knudsen et al. [65] and a negative correlation was found between epinephrine and the percentage of CD3−CD56+ cells. The Authors suggested that epinephrine plays a dual effect on NK cells: an acute effect by which NK cells are mobilised from depots and a chronic effect which reduced the number of these cells in peripheral blood. The same conclusion was supported by the observations reported by Nagao et al. [62] who suggested that moderate exercise induced a mobilisation of NK cells in men due to down-modulation of NK cell surface adhesion molecules by catecholamines but, in contrast to this, NK cells rapidly decreased below baseline levels after the exercise. Moreover, the increase in epinephrine due to 60 min of bicycle ergometer at 75% of VO2max and the increase in epinephrine in the same subjects due to its infusion up to reach plasma concentrations comparable with those seen during exercise, induced a comparable increase in NK (CD16+) cells concentration in the peripheral blood which, however, dropped below basal levels 2 h afterward. Even though a positive relationship between an exercise induced increase in NK cell numbers and activity and corresponding rises in blood catecholamines seems to be demonstrated [62,66,67], Mackinnon [26] pointed out that the effects of exercise-dependent increasing blood catecholamines on NK cells number and concentration depend on differences in muscle group activation, intensity and duration of exercise. In fact, in subjects of the above experiment, 60 min bicycle exercise at 80% of the VO2max induced a significantly higher increase in catecholamines and in peripheral blood NK cells than 60 min back-muscle training up to 29% of VO2max. Both moderate exercise (45 min at 60% VO2max) in women and intense exercise (80% VO2max up to exhaustion) in men showed a workload related increase in NK cells associated with a correspondingly high increase in plasma catecholamines [67]. All the above mentioned observations suggest a catecholamine-dependent increase in both concentration and percentage of NK cells when workload increases progressively up to reach the VO2AT. However, despite catecholamines still increase up to reach VO2max, it has been found that NK cells ceases to increase further [51]. To explain this fact, some other changes occurring in hormone plasma levels and soluble mediators, when anaerobic mechanisms are recruited, must be considered. Prostaglandin E2 (PGE2), Cortisol and prolactin are all known for their inhibitory action on exercise-induced increases in NK cell numbers and activity, and each of them might be a candidate to play such a role in the anaerobic-dependent smoothing of the NK versus workload relationship.

### 3.3. Prostaglandins

Much evidence supports a marked effect of PGE2 on NK cell biology [68]. Recent research [69] has reported that NK cells express all PGE2 EP receptors and that PGE2 acts on NK cells through the EP2 and/or EP4 receptor subtypes, which are known to be powerful activators of the adenylate cyclase system. The adenylate cyclase system is involved in inhibiting killing by NK cells and inducing the CD94/NKG2A inhibitory NK receptor following PGE2 signaling [70], and for this PGE2 has an inhibitory effect on NK cell function. It can be deduced that the EP2/EP4 receptors have emerged as pivotal regulators of NK cell activity, and targeting these receptors may prevent NK inhibition by PGE2. Recently, Uchida et al. [71] compared four different intensities of a bench press exercise for PGE2 concentrations in the blood of 35 healthy men. They found that, with respect to non-exercising subjects, Serum PGE2 concentration increased markedly (P < 0.05) after exercise consisting in a workload of 100% of maximum. These observations support the possibility that PGE2 could well explicate their inhibitory activity on NK cells when physical exercise reaches very high workloads.

### 3.4. Cortisol

It is well known that sustained exercise induces an increase in blood cortisol levels, and Gatti et al. [72] showed that, in vitro, pre-incubation for 20 h of peripheral blood mononuclear cells with 1 × 10^−8 to 1 × 10^−7 cortisol resulted in a significant decrease of NK cell activity. However, it has been observed that cortisol begins to accumulate in the blood after a delay of several minutes from the start of exercise and it peaks at a large stage during exercise or shortly afterwards, depending on the type and intensity of activity that is undertaken [56].

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During acute time-limited exercise stress, as is an incremental exercise up to exhaustion, previous experiments showed that blood cortisol did not increase [25,56] or increased slightly as it is unlikely that such a modest increment alone can account for the magnitude of the exercise induced relative immunodepression [14] shown when workload passes from VO$_{2\text{-AT}}$ to VO$_{2\text{max}}$. Singh et al. [73], before a high-intensity exercise, randomly gave dexamethasone, hydrocortisone and placebo to healthy subjects and immediately after the exercise they observed that increases in CD56 lymphocyte subsets were comparable in all three conditions. This observation concords with an exclusion of such an inhibitory role of endogenous corticosteroids in the lack of NK cell increase when the anaerobic threshold has been overcome during an incremental exercise. Furthermore, during the course of 4 h immobilization stress in pigs, a negative correlation between blood cortisol and NK cell cytotoxicity was seen only at the late phase of the stress [74]. It is possible that the delayed cortisol response to exercise stress may be initiated by a drop in blood glucose levels as well as against tissue damage due to vigorous and prolonged muscular effort [56].

### 3.5. Prolactin

De Meirleir et al. [75] and Luger et al. [76] showed that subjects exercising at a workload lower than that corresponding to their anaerobic threshold did not increase their plasma prolactin levels, but this hormone rose promptly and significantly in the plasma when workload reached and overcame anaerobic threshold. On the other hand, Melis et al. [77], in elite windsurfers simulating windsurfing movements with a specific laboratory ergometer, found that after 30 min of heavy exercise blood levels of prolactin were significantly reduced. These athletes also reported to have not reached any sensation of fatigue during the test, and this concords with the idea that to increase plasma levels of prolactin such psycho-physical stressors are need. Chambers et al. [78] were able to identify in rat NK cells the expression of receptors for prolactin by a combination of reverse transcription/polymerase chain reaction and Southern blotting. Moreover, subjects with pathological hyperprolactinemia showed a reduced function of NK cells which was reversed by bromocriptine treatment, and a prolactin-induced low active binding to target cells together with a reduction in active killing and recycling capacity in NK cells was proposed [79]. It has been found that when exercise workload overcame VO$_{2\text{AT}}$ the increase in blood lactate may coincide with a wide increase in blood prolactin levels [76]. This, in turn, may interact with NK cell surface receptors for prolactin limiting further increases in NK cell count in the peripheral blood. An exaggerated response in the inflammatory cascade, which might compromise the actual request of mechanical work from exercising muscles, could be prevented via the activation of the hypothalamic-pituitary system from which prolactin can be released into the blood. Prolactin, in turn, may reduce NK cell contribution in the inflammatory response to strenuous exercise, thus contributing to maintain the highest work output from recruited muscles.

### 4. NK CELLS AND PRO-INFLAMMATORY/ANTI-INFLAMMATORY ADVERSE MECHANISMS

Although many previous findings [12,14,15,80-83] support the increase in circulating NK cell count during progressive exercise as well as the rapid post-exercise NK cell recovery, however, the experiment by Del Giacco et al. [51] showed that the number of NK cells at the maximum workload did not further increase with respect to their number counted at VO$_{2\text{AT}}$. This fact probably meant that when the contribution from anaerobic sources for exercising muscles became relevant (i.e. in correspondence of the VO$_{2\text{AT}}$) and progressively increased up to reach the maximum workload, NK cells ceased to increase. A mismatch between the progressively increasing physical stress, occurring from VO$_{2\text{AT}}$ to VO$_{2\text{max}}$, and the corresponding NK-dependent defence against micro-organisms [15,84] should have happened when exercise work rate overcame the VO$_{2\text{AT}}$ and progressively approached the VO$_{2\text{max}}$. Interestingly, the applied regression analyses in the experiments of Del Giacco et al. [51] indicate that when the value of NK cell count at VO$_{2\text{max}}$ was subtracted from that at VO$_{2\text{AT}}$, the lower the difference was, the higher the corresponding excess of carbon dioxide in the expired air (CO$_2$-exc) was. Considering that recent experiments have confirmed that the higher CO$_2$-exc is the higher the rate of lactate production in the blood is [85], thus a higher engagement in the lactate metabolism could induce a greater limitation to an increase in the NK cell concentration and percentage in the peripheral blood leukocytes. The above considerations suggest that a progressively higher lactate metabolism involvement could induce a limitation to further increase the NK cell count and their relative percentage in peripheral blood leukocytes. However, data from the Eq. (1) concerning the quadratic regression obtained by Del Giacco et al. [51] suggested that this trend could be reversed in correspondence of very high CO$_2$-exc values (i.e. values overcoming those corresponding to the...
parabola vertex). This aspect agrees with the possibility that a pinpoint threshold for CO$_2$-exc should occur, above which a shift from inhibition to activation of NK cells activity is induced. In the light of the pro-inflammatory/anti-inflammatory adverse mechanisms which take place when a strenuous exercise is being performed, such a pro-inflammatory/protective mechanism against infection tends to progressively recover with respect to the anti-inflammatory/performance-preserving mechanism. Nevertheless, this hypothesis must be confirmed by further experiments in which NK cell numbers and percentage in the peripheral blood reached at critical time points in subjects working over the anaerobic threshold, as are those comprised between VO$_2$-AT and VO$_2$max, are compared with corresponding blood levels of prolactin, cortisol, catecholamines and prostaglandin E-2.

5. NK CELLS, EXERCISE AND TUMOURS

Translating all the above considerations on possible interactions among NK cells, physical exercise and tumour proliferation/inhibition, a recent and very important study by Inbar et al. [86] on rats, to which a dose of CRNK-16 leukemia cells was administered, showed that when they were subjected to intermittent forced swim stress mortality rates were increased. However, prolonged $\beta$-adrenergic blockade increased baseline survival rates possibly by blocking tumour-related levels of catecholamines and prostaglandins. The authors found that the physical stressor transiently suppressed NK activity against CRNK-16 lines on a per NK basis. They concluded that environmental stress, like strenuous exercise, promoted leukemia progression in rats, potentially through suppressing the cell mediated immunity mainly played by NK cells. The role of exercise therapy in the rehabilitation of cancer patients and survivors is becoming increasingly important as it is thought to modulate immunity and inflammation. However, more knowledge about the effects of exercise on immune function in these patients is needed. A systemic review was recently carried out by Kruijsen-Jaarsma et al. [86] and they found that various immune parameters improved after exercise. However, knowledge of the effects of exercise on immune function in cancer patients is still limited. In this systematic review, the authors found that natural killer cytotoxicity increased after exercise in cancer patients, along with lymphocyte proliferation and granulocyte cell counts. The number of leukocytes, lymphocytes, natural killer cells, T lymphocytes, C-reactive protein, and pro- and anti-inflammatory mediators remained stable in response to exercise. In agreement with Kruijsen-Jaarsma et al., it can be concluded that additional research is needed to gain insight into the mechanism linking exercise and immune function of NK cells in different populations, as well as to better understand the association between this immune parameter and clinical outcomes.

6. ACKNOWLEDGEMENTS

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