Processes Governing the Retention of Phosphorus and Nitrogen in Nyashishi Wetland

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Abstract

One of the important functions of the wetland is the retention of catchment nutrients and improving lacustrine water quality. This study analyzed how much nutrients were retained in the Nyashishi wetland, southern part of Lake Victoria, and went further to analyze the processes which were responsible in the reduction of nutrients. Three major processes were analyzed in this study, namely nutrients uptake by three macrophyte species (Eichhornia crassipes, Cyperus papyrus and Typha domingensis) dominating the Nyashishi wetland, nitrification and denitrification processes. The study demonstrated high nutrients retention especially phosphorus nutrients of which up to 98% were retained. In some occasions, particularly for nitrogen nutrients, there was 0% retention especially during wet season. In some other instances, the inflow exceeded the outflow meaning that, more nutrients were generated by the wetland itself. Among the three processes analyzed, nutrients uptaken by macrophytes were more efficient in reducing nutrients in wetland water. Biological nitrification and denitrification which are believed to be the major pathway for ammonia removal in both natural and constructed wetlands were less important in this study. Among the macrophyte species, Eichhornia crassipes demonstrated higher uptake rate than the other two species possibly due to its high turnover rate. This ability can be exploited in removing excess nutrients from runoff by frequent harvesting of the mature plants.

Keywords

Phosphorus; Nitrogen; Nitrification; Denitrification; Nutrients Uptake
1. Introduction

Wetlands constitute an important part of the catchment area of Lake Victoria, and perform a number of beneficial roles that have direct consequences on the lake ecosystems [1]. One of the most important functions is the retention of catchment nutrients, such as nitrogen and phosphorus, thus maintaining and improving the lacustrine water quality [2]. Through the uptake of nutrients by wetland sediments and plants, and by sedimentation of nutrient-rich particulate matter, wetlands can act as buffers against Lake Eutrophication [1] [3]. Another identified fate of nutrients once they enter the wetland is loss through degassing, and direct flow through the wetlands [2].

Wetland macrophytes play an important role in removing nutrients through incorporation of nitrogen and phosphorus in plant tissue to produce plant biomass. If wetland macrophytes are sustainably harvested, this could form a permanent removal mechanism for nutrients and other substances incorporated in the plant tissue. Several macrophytes, such as Water hyacinth (Eichhornia crassipes), water lettuce (Pistia stratiotes) and duckweed (Lemnaceae) [4] have been reported for the efficiency removal of nutrients.

Particularly, nitrogen removal in the wetland is influenced by several factors, such as the type of macrophytes present in the wetland, sediment layers and the speed of incoming water. Transformation pathways are nitrification and denitrification, decomposition and mineralization of organic nitrogen. Biological nitrification, which is facilitated by nitrifiers, such as Nitrosomonas, Nitrobira, Nitrosococcus and Nitrobacter followed by denitrification is believed to be the major pathway for ammonia removal in both natural and constructed wetlands [5].

Scanty information is available on the removal and transformation pathways of nitrogen and phosphorus in tropical natural wetlands [6] [7]. In the last decades, the human population around Lake Victoria has increased considerably. The population has grown from 4.6 million in 1932 to 27.7 million in 1995 and was estimated at 38 million in 2010 [8]. Currently, Mwanza region supports about 4 million people, with a population growth rate of 3.4%. Such growth leads to increases in domestic and industrial waste and sewage water, firewood burning and agricultural land use [9]. Soil erosion and runoff are the consequences of the growing demand for agricultural land use, leading to increased nutrient input into the lake [9]. Atmospheric deposition and land runoff are responsible for approximately 90% and 94% of the phosphorus and nitrogen input into the lake [10]. Pollution loading to Lake Victoria due to urban waste water runoff was estimated to be 719 tons/year of total nitrogen and 292 tons/year of total phosphorus; out of which 324 tons/year of total nitrogen and 208 tons/year of total phosphorus are from industrial loading [11]. This exerts substantial pressures on already inadequate sewage treatment systems. Nyashishi River alone has been reported to transport up to 62 tons/year total nitrogen and 11 tons/year of total phosphorus [11]. This has caused substantial phosphorus and nitrogen being washed into the lake. Consequently, eutrophication is increasingly continuing to become a significant issue in Lake Victoria [12] [13]. Thus, the presence of functional wetlands is a vital means of reducing the amount of phosphorus and nitrogen deposited in the Lake. The current study was designed to examine the removal capacity of the Nyashishi wetland, and the major processes governing the removal of nitrogen and phosphorus in the natural Nyashishi wetland. This information is necessary in order to fully utilize the potential of this wetland in maintaining the water quality of Lake Victoria.

2. Materials and Methods

2.1. Study Area

Nyashishi wetland is located on the western side of Lake Victoria some 15 km from Mwanza City. The area is characterized by a double maxima rainfall, with heavy rains occurring during October to December. The wetland is traversed by the Nyashishi River, which is an intermittent river draining stormy waters and domestic waste water into Lake Victoria. The wetland is lined with multiple macrophyte species such as Cyperus papyrus community and Typha domingensis with small patches of Phragmites mauritianus and mixed Cyperus-typha Figure 1. Samples for this study were drawn from the water inlet section (designated as SSW), which was mainly the river before entering the wetland, the within wetland section (Swamp), and the outlet which was located at the river mouth (OW). The distance from SSW to the outlet point was about 4 km. Water samples for nutrients analysis were collected from the inlet (SSW) and outlet sites (OW), while the nitrification and denitrification experiments were conducted within the wetland (Swamp), (see Figure 1).

2.2. Determination of Nutrients Retention in the Wetland

Water samples for nutrients analysis were collected using a 2 litres Van Dorn opaque plastic water sampler.
Water samples were kept into 1 litre plastic bottles that were pre-cleaned with phosphorus-free detergent, stored in cool boxes with ice and transported to the water quality laboratory in Mwanza for processing. In the laboratory, turbidity was measured using a turbidimeter, while sub samples for soluble reactive phosphorus analysis were filtered through GF/C and analysed using the ammonium molybdate method (APHA, 2005). Kjeldhal method was used to determine organic nitrogen. Total phosphorus was done by hydrolysis of unfiltered sample using potassium persulphate in order to oxidize all P compounds to PO₄-P followed by Ammonium molybdate method determination (APHA, 2005). The ammonium-nitrogen (NH₄-N) analysis was based on the formation of indophenol blue. Nitrate (NO₃-N) was determined using the Sodium-salycilate method [14] Calorimetric determinations were carried out using a UV-2001 spectrophotometer (Hitachi Ltd, Tokyo, Japan). Percent nutrients removed were calculated according to Equation (1).

\[
\text{% removed} = \left( \frac{C_{\text{in}} - C_{\text{out}}}{C_{\text{in}}} \right) \times 100
\]

where \(C_{\text{in}}\) is the concentration at the inlet (µg/l), \(C_{\text{out}}\) is the concentration at the outlet (µg/l).
2.3. Nutrients Uptake by Wetland Macrophyte

Three plant species which were dominant in the Nyashishi wetland (*Eichhornia crassipes*, *Cyperus papyrus* and *Typha domingensis*) were selected for the nutrient uptake experiment. The experiment consisted of 12 replicates and was run for 28 days. Initial concentrations of NO₃-N, NH₄-N and SRP, were determined at the beginning of the experiment. Subsequent samplings were done after two days and after every seven days in order to determine the uptake rate of nutrients. The nutrients uptake rate by the plants were calculated as the difference between measured concentration values and the previous concentration values over the experiment period.

2.4. Denitrification Experiments

Denitrification rates were determined at one sampling point, the swamp, which was in the middle of the wetland. Plastic containers of 1 litre capacity were half filled with nearly anoxic water. 20 mg/l concentration of 1-Allyl-2-thiourea (ATU) was added to the samples to inhibit the nitrification processes. Containers with anoxic water samples were sealed and then inserted into water-filled holes at the site. The experiment was conducted for 24 hours. Denitrification and nitrification were carried out according to Zimmo [4] except that; plastic containers were inserted in sediments instead of PVC’s. Denitrification rate was determined by considering the decrease in NO₃-N concentration over the 24 hours incubation period. The nitrification rates were determined by considering the increase in NO₃-N concentration over 24 hour’s incubation period. Then after, the containers with samples were transferred to the laboratory for final NO₃-N analyses according to APHA (2005).

2.5. Nitrification Experiments

The setup for nitrification experiment was similar to that of denitrification. The only difference between the two setups is that, samples in this experiment were incubated without the nitrification inhibitor 1-Allyl-2-thiourea (ATU) so as to allow the nitrification process to occur along with denitrification. The nitrification rates were determined by considering the increase in NO₃-N concentration over 24 hour’s incubation period.

2.6. Data Analysis

Percentage reductions in different physico-chemical parameters were compared using a student T-test to determine seasonal variation (*i.e.* wet vs. dry season). Comparison for nutrients uptake among the different macrophyte species was performed using a One-Way Analysis of Variance (1-way ANOVA). The data were first tested for homogeneity of variance using a Levene’s test, followed by a log transformation prior to analysis whenever homogeneity of variance was not observed. Non parametric test was used when the data were still not normally distributed even after transformation. Statistical significance was measured at $\alpha = 0.05$. All analyses were performed using SPSS version 17.

3. Results

3.1. Electrical Conductivity

The electrical conductivity (EC) at station SSW ranged from 122.4 ± 4.1 µS/cm to 1107 µS/cm and at station OW the EC values ranged from 109.23 ± µS/cm to 364.3 ± 3.6 µS/cm The mean (±SE) percentage reduction of EC was 47.9% ± 11.1% (range: 2.3% - 79.9%) in wet season and 74.5% ± 2.6% (range: 65.5% - 79.9%) in dry season respectively. The lowest percent reductions were observed in May 2008 (2.3%), but also low percentage values were recorded in April 2008 (13.4%) and 2009 (18.4%) respectively. The highest reduction was recorded in October and November – which was the onset of the rain season. There was a significant difference in mean electrical conductivity (EC) reductions between wet and dry seasons with the dry season having higher reductions than wet season ($t_{(7,33)} = 2.33$, $p = 0.049$, Table 1). Values in dry season were higher and comparable for the whole period (Table 1).

3.2. Turbidity

Turbidity at station SSW ranged from 12.0 ± 2.1 NTU to 156.0 ± 3.2 NTU and at station OW the turbidity values ranged from 2.7 ± 1.4 NTU to 9.3 ± 2.1 NTU. Turbidity decreased, from station SSW to OW by 73% to
Table 1. Percentage reduction in values of physical variables and nutrients in Nyashishi wetland during the wet and dry season: EC = Electrical conductivity, SRP = soluble reactive phosphorus, TP = Total phosphorus, NH$_3$-N = Ammonia nitrogen, TN = Total Nitrogen, NO$_3$=Nitrate.

<table>
<thead>
<tr>
<th>Months</th>
<th>EC (µS/cm)</th>
<th>Turb. (%)</th>
<th>SRP (µg/l)</th>
<th>NO$_3$ (µg/l)</th>
<th>TP (µg/l)</th>
<th>NH$_3$-N (µg/l)</th>
<th>TN (µg/l)</th>
</tr>
</thead>
<tbody>
<tr>
<td>JAN</td>
<td>67.05</td>
<td>94.85</td>
<td>72.88</td>
<td>34.09</td>
<td>80.76</td>
<td>33.38</td>
<td>64.77</td>
</tr>
<tr>
<td>APRIL</td>
<td>13.36</td>
<td>87.32</td>
<td>75.25</td>
<td>36.41</td>
<td>57.17</td>
<td>−100.00</td>
<td>64.38</td>
</tr>
<tr>
<td>MAY</td>
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<td>73.04</td>
<td>74.94</td>
<td>10.84</td>
<td>66.47</td>
<td>83.33</td>
<td>53.50</td>
</tr>
<tr>
<td>JAN</td>
<td>74.61</td>
<td>76.00</td>
<td>90.82</td>
<td>18.10</td>
<td>68.12</td>
<td>1.41</td>
<td>56.91</td>
</tr>
<tr>
<td>APRIL</td>
<td>18.35</td>
<td>95.09</td>
<td>89.54</td>
<td>56.75</td>
<td>76.85</td>
<td>75.12</td>
<td>44.68</td>
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<tr>
<td>MAY</td>
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<td>81.70</td>
<td>91.16</td>
<td>65.81</td>
<td>79.07</td>
<td>12.47</td>
<td>59.47</td>
</tr>
<tr>
<td>OCT</td>
<td>79.81</td>
<td>80.95</td>
<td>80.35</td>
<td>26.66</td>
<td>69.33</td>
<td>66.03</td>
<td>12.26</td>
</tr>
<tr>
<td>NOV</td>
<td>71.89</td>
<td>88.69</td>
<td>98.04</td>
<td>47.64</td>
<td>95.36</td>
<td>91.44</td>
<td>69.19</td>
</tr>
<tr>
<td>JUN</td>
<td>65.55</td>
<td>92.94</td>
<td>95.35</td>
<td>52.21</td>
<td>91.97</td>
<td>83.56</td>
<td>55.18</td>
</tr>
<tr>
<td>JULY</td>
<td>72.27</td>
<td>93.33</td>
<td>94.60</td>
<td>49.76</td>
<td>90.73</td>
<td>84.15</td>
<td>60.81</td>
</tr>
<tr>
<td>SEPT</td>
<td>76.81</td>
<td>92.21</td>
<td>93.65</td>
<td>46.32</td>
<td>89.31</td>
<td>84.74</td>
<td>66.97</td>
</tr>
<tr>
<td>JUN</td>
<td>78.17</td>
<td>85.71</td>
<td>74.28</td>
<td>23.91</td>
<td>54.22</td>
<td>46.06</td>
<td>−64.71</td>
</tr>
<tr>
<td>JULY</td>
<td>79.86</td>
<td>77.78</td>
<td>72.47</td>
<td>9.73</td>
<td>67.80</td>
<td>29.04</td>
<td>−37.45</td>
</tr>
</tbody>
</table>

93.6% and 50% to 87 % during the wet and dry seasons respectively. The mean reduction in turbidity did not differ significantly between the wet (mean ± SE = 84.7% ± 2.9%) and dry (88.4% ± 3.0%) seasons ($t_{(11)} = 0.84$, $p = 0.42$; Table 1), implying that turbidity reduction is not seasons dependent.

3.3. Soluble Reactive Phosphorus

Soluble reactive phosphorous ranged from 62.3 ± 2.8 µg/l to 806.96 ± 3.9 µg/l at station SSW and from 12.25 ± 1.6 µg/l to 47.46 ± 2.7 µg/l at OW station. The percentage reduction was in wet (84.1% ± 3.3%; range: 72.9% - 98.0%) and dry (86.1% ± 5.2%; range: 72.5% - 95.4%) seasons respectively. However, the differences between seasons were not significant ($t_{(9)} = 0.314$, $p = 0.761$; Table 1), probably implying that phosphorus sequestering is an all-time process in the wetland.

3.4. Total Phosphorus

Total phosphorus concentration ranged from 126.5 ± 3.1 µg/l to 1318.8 ± 4.1 µg/l at station SSW and from 38.79 ± 2.6 µg/l to 253.76 ± 3.4 µg/l at the OW station. The mean percentage reduction in Total Phosphorus was comparably similar between seasons ($t_{(11)} = 0.60$, $p = 0.56$; Table 1) mean for wet and dry seasons = 74.1% ± 4.1% and 78.8% ±7.6% respectively) where high and low values were recorded in both seasons. For example, while higher percent reduction in TP were recorded in November (91.44%), June (92%) and July (90.7%), the lowest reduction were found in April (57.2%) and June (54.2%) during the wet and dry seasons respectively.

3.5. Ammonia

Ammonia concentration ranged from below detection concentration to 324.7 ± 3.5 µg/l at station SSW and from 20.7 ± 2.9 µg/l to 119.1 ± 3.1 µg/l. There was a negative retention in ammonium concentration in April (100% release), followed by lowest reduction value of (1.4%) in January 2008, (12.5%) in May 2009 and (33.4%) in January 2009. Highest reduction in wet season was found in November (91.44%). Percentage reduction during dry season ranged from 29% (July) to 84.7% (September) (Table 1). Nevertheless, the differences between wet (60.6% ±23.6%) and dry (65.5% ±11.7%) season were insignificant ($t_{(9)} = 0.17$, $p = 0.87$).

3.6. Nitrate

The concentration of nitrate was 99.7 ± 3 µg/l to 333.4 ± 3.5µg/l at SSW station, and 21.35 ± 2.3 µg/l to 212 ±
3.7. Total Nitrogen
Total nitrogen ranged from 594.4 ± 3.8 µg/l to 1862.4 ± 3.9 µg/l at SSW station and the concentration at OW ranged from 374.0 ± 3.6 µg/l to 979.0 ± 4.0 µg/l. Total nitrogen reduction varied from 12.3% in October (2009) to 64.8% in January (2008) during wet season, and negative retentions of 64.7% and 37.5% were recorded during the dry season in June and July respectively (Table 1). However, mean values for wet (57.3% ± 3.1%) and dry (16.2% ± 27.8%) seasons were not significantly different (t(14) = 1.47, p = 0.32).

3.8. Nutrients Uptake by Wetland Macrophytes

3.8.1. Uptake of Soluble Reactive Phosphorus
The uptake of soluble reactive phosphorus by C. papyrus was observed starting from day 2. The concentration of SRP decreased from 215.15 ± 2.2 µg/l at the beginning of the experiment to 54.86 ± 5.92 µg/l on day 28. The average uptake rate of C. Papyrus was estimated at 5.72 ± 2.03 µg l⁻¹ d⁻¹ although the high uptake rates were observed at the beginning of the experiment and decreased at the end of the experiment (see Table 2). Uptake of SRP in E. crassipes treatment decreased from 215.15 ± 2.2 µg/l at the beginning of experiment to 13.91 ± 1.4 µg/l on day 28; and the average uptake rate of E. crassipes was estimated at 7.9 ± 0.12 µg l⁻¹ d⁻¹. Moreover, SRP concentration in T. domingensis treatment decreased from 215.15 ± 2.2 µg/l at the beginning of the experiment to 9.54 ± 3.37 µg/l on day 28; and the average uptake rate was estimated at 7.4 ± 0.07 µg l⁻¹ d⁻¹. The control treatment maintained higher SRP levels throughout the experiment period, decreasing from 215.15 ± 2.2 µg/l at the beginning of experiment to 86.73 ± 3.25 µg/l; and the average uptake rate was estimated at 4.59 ± 0.12 µg l⁻¹ d⁻¹. Although SRP concentrations kept decreasing over the experimental period, there were species-specific variations in the uptake rates of the nutrient (Figure 2(a); Table 2). For example, there was some phosphorus production in the E. crassipes treatment on the second day of the experiment (Table 2). A sharp decline in SRP in the E. crassipes and T. domingensis treatments which was observed in the first days of the experiment translate into high uptake rates. A more gradual decline in SRP, which was observed in the control and C. papyrus treatments translate into slow uptake rates (Figure 2(a)). Notwithstanding, a Kruskal-Wallis test showed no significant differences in phosphorus uptake among the three macrophyte species (KW = 1.220, p = 0.582).

3.8.2. Ammonium Uptake
Uptake of ammonia in C. papyrus treatment was observed from day 2. The concentration of ammonium decreased from 264.20 ± 0.21 µg/l at the beginning of the experiment to 41.83 µg/l on day 28. The estimated average uptake rate of ammonia in C. papyrus was 12.94 ± 3.42 µg l⁻¹ d⁻¹. The ammonia concentration in E. crassipes treatment showed a different trend. The concentration decreased from 264.20 ± 0.21 µg/l at the beginning of the experiment to 27.23 µg/l on day 14; and the estimated average uptake rate of ammonia up to day 14 was 13.16 ± 1.49 µg l⁻¹ d⁻¹. A gradual but significant increase in ammonium concentration was observed after the 14th day indicating the generation of ammonia in the treatment system. The T. domingensis treatment system showed a similar ammonia uptake trend to that of E. crassipes. The concentration decreased from 264.20 ± 0.21 µg/l at the beginning of the experiment to 32.8 µg/l on day 21, and increased again to 53.67 µg/l on day 28. Likewise, the control treatment also started to release ammonium towards the end of the experiment (Figure 2(b)). The water hyacinth (E. crassipes) showed the highest ammonium uptakes up to day 14 than the rest of the species.

Table 2. Mean (±SD) phosphorus uptake rates (µg l⁻¹ d⁻¹) by three macrophyte species in Nyashishi Wetland.

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Day 2</th>
<th>Day 7</th>
<th>Day 14</th>
<th>Day 21</th>
<th>Day 28</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>5.50 ± 1.51</td>
<td>8.51 ± 1.20</td>
<td>3.71 ± 0.90</td>
<td>0.71 ± 0.61</td>
<td>0.03 ± 0.52</td>
</tr>
<tr>
<td>E. Crassipes</td>
<td>1.58 ± 4.52</td>
<td>22.94 ± 3.13</td>
<td>1.73 ± 1.26</td>
<td>0.58 ± 0.70</td>
<td>0.25 ± 0.58</td>
</tr>
<tr>
<td>C. papyrus</td>
<td>11.93 ± 2.49</td>
<td>5.71 ± 1.87</td>
<td>3.29 ± 0.87</td>
<td>0.96 ± 1.12</td>
<td>0.69 ± 0.42</td>
</tr>
<tr>
<td>T. domingensis</td>
<td>6.67 ± 4.25</td>
<td>16.47 ± 3.13</td>
<td>4.18 ± 1.71</td>
<td>1.31 ± 0.10</td>
<td>0.17 ± 0.03</td>
</tr>
</tbody>
</table>
Table 3. Mean (±SE) ammonium uptake rates (µg l⁻¹ d⁻¹) in three macrophyte species from the Nyashishi Wetland.

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Day 2</th>
<th>Day 7</th>
<th>Day 14</th>
<th>Day 21</th>
<th>Day 28</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>13.40 ± 3.47</td>
<td>10.30 ± 2.19</td>
<td>3.30 ± 1.90</td>
<td>0.09 ± 0.20</td>
<td>1.46 ± 1.15</td>
</tr>
<tr>
<td>E. craspes</td>
<td>51.41 ± 2.03</td>
<td>14.28 ± 0.0</td>
<td>2.44 ± 0.71</td>
<td>−0.48 ± 0.32</td>
<td>−2.4 ± 1.08</td>
</tr>
<tr>
<td>C. Papyrus</td>
<td>12.94 ± 3.01</td>
<td>9.40 ± 2.24</td>
<td>6.98 ± 0.37</td>
<td>1.58 ± 1.04</td>
<td>0.01 ± 0.85</td>
</tr>
<tr>
<td>T. Domigensis</td>
<td>15.09 ± 2.76</td>
<td>11.19 ± 1.51</td>
<td>6.26 ± 1.31</td>
<td>1.67 ± 0.58</td>
<td>−0.74 ± 0.04</td>
</tr>
</tbody>
</table>

3.8.3. Nitrate Uptake
Nitrate concentration in E. crassipes treatment decreased from the initial concentration of 105.20 ± 0.0 µg/l to 63.53 ± 1.89 µg/l on the second day. The nitrate concentration in the E. crassipes treatment increased sharply to the highest (330.27 ± 8.53 µg/l) on the 21th day of the experiment (Figure 2(c)). Concentration of nitrate in C. papyrus treatment decreased from the initial concentration of 105.20 ± 0.0 µg/l to 69.80 ± 1.91 µg/l on the second day. After that, a significant increase was observed on the 21st day (136.23 ± 5.55 µg/l). Likewise, the concentration in T. domingensis treatment decreased from the initial concentration of 105.20 ± 0.0 µg/l to 70.98 ± 2.58 µg/l on the 2nd day. Then there was a significant increase to 261.34 ± 0.74 µg/l on day 21 (Figure 2(c)).

3.9. Nitrification and Denitrification in the Wetland Sediments
The denitrification and nitrification rates of the Nyashishi wetland are given in Table 4. Denitrification rate ranged between 0.02 ± 0.02 µg l⁻¹ d⁻¹ and 0.082 ± 0.01 µg l⁻¹ d⁻¹ with an average value of 0.059 ± 0.16 µg l⁻¹ d⁻¹. Nitrification was also very low and negative net nitrification rates were encountered. While the lowest denitrification value of 0.024 ± 0.02 µg l⁻¹ d⁻¹ was encountered in April 2008; the maximum (0.140 ± 0.21 µg l⁻¹ d⁻¹) and minimum (−0.043 ± 0.06 µg l⁻¹ d⁻¹) nitrification rates were noted in April 2008 and June 2009 respectively. In this study, denitrification rates showed no definite pattern over time, while nitrification showed a slight increase (Table 4).

3.10. Comparison of Nutrients Consumed by Macrophyte Species in the Treatments
Comparison was made among plant species to see which species exhibited higher uptake rates (Figure 3). Water hyacinth (Eichhornia crassipes) showed the highest reduction of nitrate-nitrogen which was available in the treatment containers (31%), followed by Cyperus papyrus (26%). Percentage reduction in Typha domingensis was comparable with reduction in the control containers (Figure 2). Water hyacinth still exhibited the highest percentage reduction for nitrite-nitrogen (43%), followed by Typha domingensis (33%), while Cyperus papyrus showed the lowest performance (24%). However, there was no reduction in the control containers. Likewise, Eichhornia craspes was the most efficient in phosphorus reduction as 54% of the available soluble reactive phosphorus was noticed, followed by Typha domingensis (37%) and the lowest reduction was observed in the Cyperus papyrus containers (7%). In the control treatments, only 2% was reduced. Similarly, the same trend was observed for ammonia-nitrogen whereby 40% reduction was recorded in Eichhornia craspes containers, followed by 22% in T. domigensis containers. The percentage reduction in Cyperus papyrus and control containers was comparable (Figure 3).

4. Discussions
The results indicate that, there was reduction in most of the parameters as water passed through the Nyashishi wetland, although the retention capacity was different according to seasons. The higher reduction of electrical conductivity in the dry season is caused by the low flushing of effluents in the wetland which normally increases due to runoff in the rain season. During the dry season there is low volume of water in the river, thus reduced water flow. This situation allows suspended matter (i.e. detritus) to settle to the bottom, improving nutrient removal mechanisms. Moreover, at the inlet point, there is pool of water which is used for animal watering espe-
Table 4. Mean (±SE) for four measurements of denitrification and nitrification rates (µg l⁻¹ d⁻¹) at station WW in Nyashishi wetland.

<table>
<thead>
<tr>
<th>Experiment</th>
<th>Denitrification rate</th>
<th>Nitrification rate</th>
</tr>
</thead>
<tbody>
<tr>
<td>Experiment 1</td>
<td>0.024 ± 0.02</td>
<td>−0.043 ± 0.06</td>
</tr>
<tr>
<td>Experiment 2</td>
<td>0.082 ± 0.01</td>
<td>−0.025 ± 0.01</td>
</tr>
<tr>
<td>Experiment 3</td>
<td>0.058 ± 0.02</td>
<td>−0.017 ± 0.00</td>
</tr>
<tr>
<td>Experiment 4</td>
<td>0.074 ± 0.02</td>
<td>0.14 ± 0.21</td>
</tr>
</tbody>
</table>

Figure 2. Change in (a) soluble reactive phosphorus (b) Ammonia and (c) nitrate concentration in three experimental macrophyte species. C = control; WH = water hyacinth (E. crassipes); P = Cyperus papyrus; T = Typha domingensis.

Spatially during the dry season. During the dry season the water flow diminishes to a standstill in some years, and due to higher evaporation, there is higher nutrients concentration. The decrease in water flow might have contributed to the higher reduction in electrical conductivity especially in November and December. These months in Mwanza region are the onset of the rainy season, so the water volume was still low.

The reduction in the levels of turbidity along the wetland was manifested in both seasons. The reason is due to huge swampy areas in the wetland made of several species of wetland macrophyte, such as *Cyperus papyrus* L. and *Phragmites mauritianus* (Kunth). The extensively interlaced but permeable root mat of papyrus effectively retained suspended organic particles which provided sufficient substrates for the proliferation of heterotrophic bacteria as also stated by [7]. In so doing it makes wetland systems to work as giant filters, by removing suspended solids, and dissolved nutrients pollutants from the water before it enters the lake [15]. Moreover, macrophyte reduces turbidity using their root hairs which have electrical charges that attract opposite charges of colloidal particles. When suspended solids with opposite charges pass through the root hairs, they adhere onto the roots. Consequently, they are slowly assimilated by the plant and microorganisms [16].

Phosphorus species analysed (SRP and TP) were retained in the wetland throught the year both in the wet and
dry season. This was probably due to attachment of colloidal particles to root hairs as well as uptake by macrophytes as also stated by [17]. The retention of phosphorus in the wetland was supported by laboratory experiments on nutrients uptake by wetland plants whereby; the decrease in phosphorus species was recorded in all experimental treatments. The high productivity of macrophytes enables substantial amount of nutrients to be stored in plant biomass.

Ammonia exhibited a negative retention especially in April during the peak of the rain season, and other retention was observed in September and November. The negative retention is explained by high rate of flushing due to the high flow rate of water through the wetland. Moreover, higher concentration of ammonia was recorded in the lake during the same season, suggesting that other chemical/biological activities were taking place in the system. But also, the increase might have been caused by decomposition of organic matter brought by runoff into the wetland which used up available oxygen in the sediment and thus nitrification process ceased. These findings are supported by [18] who stated that, at relatively low dissolved oxygen, nitrification of ammonia ceases and the absorptive capacity of the sediments is reduced. This causes a marked increase of the release of ammonium from the sediments.

Nutrients release from the wetland was also recorded for TN in June and July; this indicates that the wetland does not effectively retain TN. Similar findings were reported by [19] Their study demonstrated that, the wetland had a negative retention of total nitrogen, and this was associated with the large export of organic nitrogen produced in the wetland. Findings from Kirinya wetland in Uganda, showed a low retention of nitrogen such that; 75% of the nitrogen flowing into the wetland flowed out into the lake and only 25% was retained [15].

In the experiment on nutrients uptake by wetland plants, phosphorus decreased in all treatments, even in the controls without plants. The phosphorous removal was not only by the macrophyte, as it has been also observed by [20] and [21]. This suggests the importance of biological removal for sewage treatment in constructed and sedimentation ponds apart from chemical precipitation by alum or lime in conventional sewage treatment plants [20].

The nutrients uptake experiments demonstrated that, *E. Crassipes* was the most efficient plant in phosphorus reduction as 54% of the available soluble reactive phosphorus was uptaken, compared to 37% by *Typha domingensis* and only 7% of *Cyperus papyrus*. The higher uptake rate by *E. Crassipes* was due to relatively higher growth rate of the plant as compared to the other two plant species [22]. *E. Crassipes* can double within a short time; say 6 days (Mitchell, 1976). Moreover, *E. crassipes* has also extensive root system which has excellent filtration and high bacterial density. The root system provide surface onto which bacteria grow; provide media for filtration and adsorption of solids and in some cases release natural polymers that facilitate flocculation and sedimentation [23]. The other two species showed less absorption capacity because the submerged portions of their stems has less filtration and bacterial density than the roots of floating plants like *E. crassipes* [23].
The reduction in ammonium concentration was observed in all treatments including the control, from the observation it can be inferred that, apart from uptake by plants, a certain amount of ammonium may have been oxidized to other forms of N, otherwise, the difference would have been less in the containers with no plants. This conclusion is backed by the nitrate results which showed an increase in nitrate concentration in all treatments instead of removal.

Like other nutrients, uptake of both ammonium and nitrate were slightly higher for *E. crassipes* than in the other plant species, due to its high growth rate than the two species as described above. These findings are in agreement with the results by [24] who obtained higher values of phosphorus and nitrogen nutrients uptake in *E. crassipes* than *Pistia stratiotes* in the Pantanal, Brazil.

Although bacterial nitrification-denitrification is thought to be the dominant cause of nitrogen removal from the wetland soils, [25], generally in this study the rates were low. This is because, the amount of oxygen transferred to the wetlands through macrophyte roots and atmospheric diffusion is relatively small compared to the amount of oxygen required to oxidize ammonia due to the dense vegetation cover of papyrus which tends to limit atmospheric aeration and oxygen production by algal photosynthesis; hence less nitrate is available to undergo the denitrification process e.g. [7] [26]. Similarly, [15] concluded that, denitrification did not have a significant contribution in the nitrogen budget due to the minor importance of nitrate in the system. Concurrently, [19] also stated that, denitrification was of minor importance on the influence of nitrate in Nyashishi wetland.

5. Conclusion

According to the results of this study, plants nutrients uptake demonstrated to be the dominant nutrients removing mechanism in the Nyashishi wetland system. Among the wetland macrophytes, *E. Crassipes* was the most efficient in nutrients removal as compared to *Typha domingensis* and *Cyperus papyrus*. However, apart from the macrophytes, the nutrients uptake experiment showed that, microorganisms and algae also remove the nutrients. Moreover, it is important to note that, this “locking” of nutrients in aquatic macrophytes is only short-term because of the rapid turnover especially for water hyacinth. If plants are not harvested, the dead tissues decompose rapidly and release nutrients back into the water. Frequent harvesting of the biomass is therefore recommended and is necessary for complete removal of nutrients from the wetland system. Since the nutrients uptake is high despite the relatively short life of the plant, this is an advantage for wetland management. This ability can be exploited in removing excess nutrients from runoff by frequent harvesting of the mature plants.

References


