Assessment of Insulin Resistance and Insulin Growth Factor-1 in Egyptian Patients with Chronic Hepatitis C

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Abstract

Introduction: Insulin resistance (IR) is documented in patients with chronic hepatitis C (CHC), plays an important factor in disease progression and predicting poor response to treatment. In chronic liver disease, levels of insulin-like growth factor-1 (IGF-1) were correlated with IR. Aim: To evaluate IR and serum levels of IGF-1 in Egyptian patients with CHC after anti-viral therapy. Patients and Methods: Forty biopsy-proven, non-diabetic, CHC patients, who received combined IFN/ribavirin therapy, in addition to 10 healthy controls were studied. Serum levels of IGF-1, growth hormone (GH) were measured and HOMA-IR (homeostasis model assessment of IR), body mass index (BMI) were calculated. Baseline data, retrieved from patients’ files before initiation of therapy, together with response to antiviral therapy were analyzed with respect to the measured variables. Results: All patients possessed a significant higher HOMA-IR score (p = 0.02), GH levels (p < 0.001) and blood glucose levels (p < 0.001) than controls while significantly lower levels of IGF-1 were found in patients (p = 0.008) with no significant difference between responders and non-responders as regards the previously mentioned variables. HOMA-IR score was significantly correlated with GH levels, p = 0.003 and with IGF-1 levels in both responders and non-responders. Low IGF-1 levels were associated with increase in fibrosis stages and were significantly correlated with insulin (r = –0.499, p = 0.001). Conclusion: CHC patients exhibit an increased HOMA-IR score reflecting the existence of IR irrespective of treatment response. Low IGF-1 levels were associated with advanced stages of fibrosis and thus could contribute to the progression of hepatic fibrosis in CHC.

Keywords: Hepatitis C, Insulin Resistance, Interferon, Ribavirin, IGF-1

1. Introduction

Hepatitis C virus (HCV) induced insulin resistance (IR) has been related to steatosis development, fibrosis progression and non-response to interferon (INF)/ribavirin therapy [1]. HCV infected patients showed a higher prevalence of diabetes mellitus (DM) and IR than those infected with hepatitis B [2]. A prospective study of 500 HCV patients demonstrated a significant correlation between genotype 1 or 4 infection and IR, both in the presence and absence of DM [3]. The mechanism implied in IR is the enhanced production of tumor necrosis factor α (TNF-α) by the HCV core [4]. Moreover, TNF-α inhibits the function of insulin substrate 1 and 2 and decrease the expression of glucose transporter and lipoprotein lipase in peripheral tissues [5,6]. Increased liver iron accumulation and modification in the levels of adipocytokinemria can have an additional effect on insulin sensitivity in HCV infection [4].

IR can occur early in the course of chronic liver disease [7] independent of body weight, stage of liver disease, and presence or absence of overt DM [8,9]. Moreover, IR is an important predictor of sustained response to antiviral therapy [10], thus improving insulin sensitivity was suggested as a useful adjunct to therapy [11].

Chronic liver disease (CLD) is characterized by acquired growth hormone (GH) resistance, low concentrations of insulin-like growth factor 1 (IGF-1) with respect to normal or elevated GH levels [12]. IR can be induced by high amounts of GH [13] and was correlated with IGF-1 [14]. GH resistance is determined by several factors including malnutrition, impaired liver function, re-
duced expression of hepatic GH receptors and a possible role of TNF-α in blunting hepatic response to GH in patients with CHC. IGF-1 was significantly lower in CLD patients due to decreased hepatic production or modified bioavailability secondary to decrease binding proteins.

2. Aim

Assessment of the IR and serum levels of IGF-1 in Egyptian patients with chronic hepatitis C (CHC) after treatment with combined IFN/ribavirin therapy.

3. Patients and Methods

3.1. Patients

Forty non diabetic (fasting blood glucose level less than 6.0 mmol/litre), CHC patients, who received combined IFN/ribavirin therapy in the context of a project carried out by the Schistosomal Liver Unit, Cairo University in the period of 2005-2007 and attended at regular follow up, were invited to participate in the study. Patients were enrolled after obtaining an informed consent. They included 32 males and 8 females their age ranged from 28-60 years, mean ± SD: 43.28 ± 8.25.

The control group included 10 healthy non diabetic subjects with no history of CLD. They were 3 males and 7 females, mean ± SD age: 36.60 ± 9.22 years with normal liver biochemical profile, negative seromarkers for HBV and HCV and normal abdominal ultrasonography. Patients with baseline impaired glucose tolerance (diagnosed as blood glucose ≥ 6.1 mmol/L), diabetes mellitus and history of alcohol intake were excluded.

All patients and control underwent:

- Full history taking including a nutritional questionnaire concerning dietary restrictions, diet supplementations and weight changes.
- Full clinical examination and BMI was calculated as body weight in kilograms divided by height in m².
- Recent Investigations: liver functions, kidney functions and complete blood count.
- Sample collection: After an overnight fast, 10 ml of venous blood were withdrawn from each subject in the study then the blood was centrifuged at 3000 rpm for 15 min rapidly after clotting. Serum was separated with Pasteur pipette and divided as aliquots into Ependorrff tubes. One tube was used for determination of fasting blood sugar using the available commercials kit. The rest of the Ependorrff tubes were kept frozen at –80°C till used for assay of insulin, growth hormone and IGF-1, also this Ependorrff was used to detect HCV infection by reverse transcriptase-polymerase chain reaction (RT-PCR).
- Fasting blood sugar and fasting serum insulin was determined by radioimmunoassay (RIA) [16].
- IR was calculated by the Homeostasis Model Assessment of Insulin resistance (HOMA-IR), using the following equation: fasting insulin (μU/ml) × fasting glucose (mmol/L)/22.5 [17].
- Serum IGF-1 and serum GH measured by RIA using reagent purchased from Nichols Institute (San Juan Capistrano, CA) [18,19].
- Recent qualitative RT-PCR for HCV detection was done for patients only. RNA was obtained using the Qiagen extraction protocol (QIAamp Viral RNA Mini kit, Qiagen, Germany) and sensitive in-house nested RT-PCR standardized [20] using primers specific for the 5’ noncoding region of the HCV genome was employed [21].
- Abdominal ultrasound.

Baseline data before the initiation of therapy that was retrieved from their files included:

- History of infection with schistosomiasis, body weight change during treatment and afterwards.
- Type and duration of IFN received: Patients received either recombinant standard IFNα-2a 3 million units taken three times per week (22 patients) or 20-kDa linear pegylated (PEG) IFNα-2a derived from Hansenula Polymorpha expression system taken once weekly 160 microgram (18 patients) SC, in addition to oral ribavirin taken daily in a dose of 1000 - 1200 mg (according to body weight).
- METAVIR scoring system [22] for assessment of liver biopsy. All of the recruited patients had activity ≤ A2 and 75% had fibrosis ≤ F2 with minimal or mild steatosis (<33%).
- Response to treatment, sustained virological response (SVR) defined as the absence of detectable HCV RNA in serum at the end of treatment and 6 months later was achieved in 10 patients, 18 patients were non responders and 12 relapers (HCV RNA becomes undetectable on treatment and is detected again after discontinuation of treatment).

3.2. Statistical Analysis

Results were expressed as mean ± standard deviation (SD) or number (%). Comparison between the mean values of the two groups was done using unpaired student t test while multiple group comparison was performed using ANOVA with post hoc using LSD test. Comparison between categorical data [n (%)] was done using Chi square test. Correlation between parameters was performed using Spearman’s rank correlation coefficient.
SPSS computer program (version 11 windows) was used for data analysis. p-value less than 0.05 was considered significant; less than 0.01 was considered highly significant and less than 0.001 was considered extremely significant.

4. Results

All patients expressed a significant higher HOMA-IR score (p-value = 0.02), GH levels (p-value < 0.001) and blood glucose levels (p-value < 0.001) than the controls; while mean levels of IGF-1 were significantly lower in CHC patients; p-value = 0.008 (Table 1). BMI did not show any statistically significant difference between recruited patients and control. Impaired glucose tolerance (blood glucose ≥ 6.1 mmol/L) occurred in 9 treated patients, 6 of them were non responders. No dietary restriction or significant weight changes was reported during treatment or in the period afterwards in any of the recruited patients, only mild weight gain was reported by the majority of responders. No significant difference was found on comparing age, sex, BMI, HOMA-IR score, IGF-1, insulin, activity and fibrosis score and steatosis grades between responders and non-responders. Impaired glucose tolerance was noted among non responders compared to responders, but with no significant difference. GH levels were significantly different between responders and non-responders, p-value 0.01 (Table 2).

Correlations studies were described as below:
- In control group: HOMA score had a significant positive correlation with both insulin (r = 0.979, p-value = 0.01) and BMI (r = 0.794, p-value < 0.01), but there was no significant correlation with age, sex, GH levels, IGF-1 levels. Moreover, IGF-1 was not correlated with age, sex, BMI, GH, insulin.
- In all 40 patients (whether responders or non responders), HOMA-IR score was significantly correlated with GH and insulin levels (r = 0.451, p = 0.003, Figure 1), r = 0.852, p = 0.01 respectively. However, age, sex, BMI, IGF-1, stages of fibrosis and achievement of SVR had no significant association with IR.
- HOMA-IR score was significantly correlated with GH (r = 0.566, p = 0.001) in non responders, and with IGF-1 in non-responders and responders (r = −0.381, p = 0.038, Figure 2) (r = −0.77, p = 0.009, Figure 3) respectively.
- IGF-1 was significantly correlated with insulin, r = −0.499, p = 0.001 (Figure 4), decreased with advanced stages of fibrosis, r = −0.270, p = 0.08 and had no significant association with age, sex, BMI and GH.

Table 1. Comparison of BMI and laboratory results between CHC patients and controls.

<table>
<thead>
<tr>
<th>Variables</th>
<th>Control (n = 10)</th>
<th>Patients (n = 40)</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>BMI &lt; 25</td>
<td>25.86 ± 4.1</td>
<td>28.82 ± 5.14</td>
<td>0.1 (NS)</td>
</tr>
<tr>
<td>GH (5 - 7 ng/mL)</td>
<td>7.77 ± 0.95</td>
<td>12.44 ± 3.83</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>IGF-1 (ng/mL)</td>
<td>195.12 ± 6.85</td>
<td>180.45 ± 16.25</td>
<td>0.008</td>
</tr>
<tr>
<td>Serum glucose</td>
<td>4.01 ± 0.32</td>
<td>5.43 ± 1.30</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>(3.9 - 6.1 mmol/L)</td>
<td>11.24 ± 0.95</td>
<td>13.69 ± 5.86</td>
<td>0.19 (NS)</td>
</tr>
<tr>
<td>HOMA-IR (≤2.5)</td>
<td>2.01 ± 0.30</td>
<td>3.45 ± 1.99</td>
<td>0.028</td>
</tr>
</tbody>
</table>

Values were expressed as mean ± SD or number (%). p-value NS = not significant, p < 0.05 = significant (S), p < 0.01 = highly significant, p < 0.001 = extremely significant.

Table 2. Clinical and laboratory data of the studied patients groups.

<table>
<thead>
<tr>
<th>Variable</th>
<th>Responders (10)</th>
<th>Non-responders and relapsers (30)</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age: mean ± SD</td>
<td>44.00 ± 6.91</td>
<td>43.03 ± 8.75</td>
<td>0.87 (NS)</td>
</tr>
<tr>
<td>Sex: M/F n (%)</td>
<td>9/1 (90/10)</td>
<td>23/7 (76.7/23.3)</td>
<td>0.36 (NS)</td>
</tr>
<tr>
<td>BMI</td>
<td>28.05 ± 4.08</td>
<td>29.07 ± 5.48</td>
<td>0.88 (NS)</td>
</tr>
<tr>
<td>GH (5 - 7 ng/mL)</td>
<td>13.92 ± 2.91</td>
<td>11.95 ± 4.01</td>
<td>0.01 (S)</td>
</tr>
<tr>
<td>IGF-1 (ng/mL)</td>
<td>183.40 ± 18.83</td>
<td>179.46 ± 15.52</td>
<td>0.37 (NS)</td>
</tr>
<tr>
<td>Serum glucose</td>
<td>5.43 ± 1.61</td>
<td>5.42 ± 1.21</td>
<td>0.79 (NS)</td>
</tr>
<tr>
<td>(3.9 - 6.1 mmol/L)</td>
<td>3 (30%)</td>
<td>6 (20%)</td>
<td>0.31 (NS)</td>
</tr>
<tr>
<td>Insulin (µU/mL)</td>
<td>14.38 ± 6.08</td>
<td>13.46 ± 5.87</td>
<td>0.96 (NS)</td>
</tr>
<tr>
<td>HOMA-IR (≤2.5)</td>
<td>3.68 ± 2.23</td>
<td>3.37 ± 1.94</td>
<td>0.9 (NS)</td>
</tr>
</tbody>
</table>

Values were expressed as mean ± SD or number (%). Comparison between the mean values of different parameters was performed using Mann Whitney U test.
1. Discussion

Chronic hepatitis C has been proposed as a metabolic disease and insulin sensitivity as a predictive factor for liver fibrosis [23]. Increased prevalence of type 2 DM, the development of hepatic steatosis, a more rapid progression of hepatic disease and reduction in the SVR.

- BMI showed a significant positive correlation with the degree of steatosis (r = 0.436, p = 0.018) which in turn was negatively correlated with the achievement of SVR (r = –0.428, p-value 0.02). Moreover, achievement of SVR was significantly correlated with GH levels (r = 0.405, p-value = 0.009) while age, sex, serum IGF-1 and fibrosis score had no significant association.

5. Discussion

Chronic hepatitis C has been proposed as a metabolic disease and insulin sensitivity as a predictive factor for liver fibrosis [23]. Increased prevalence of type 2 DM, the development of hepatic steatosis, a more rapid progression of hepatic disease and reduction in the SVR.
have been associated with IR in CHC patients [7].

In the present study, CHC treated patients, whether responders or non-responders, demonstrated significantly higher HOMA-IR and GH levels than healthy controls while levels of IGF-1 were significantly lower. Similarly previous studies reported an HCV induced IR. Moucari et al., reported that IR is a specific feature of HCV genotypes 1 and 4 even in non-diabetic patients and is associated with significant fibrosis that is independent from steatosis [3]. IR may occur at an early stage of liver disease as evidenced by patients with 0 - 1 hepatic fibrosis having higher levels of HOMA scores [8]. Hyperinsulinemia, in HCV patients, due to diminished hepatic insulin degradation rate can lead to a false increase of the HOMA-IR [24]. Furthermore, HCV infection by itself can lead to IR as HCV core protein induces hepatic steatosis and interferes with the insulin-signaling pathways [25].

Achieving sustained response depends on several factors such as age, baseline viral load, genotype, fibrosis and baseline glucose concentration [26]. IR is one of the most important host factors in the prediction of response in non diabetic HCV patients and represents a common denominator to the majority of features associated with difficult to treat patients. The risk of developing DM and impaired fasting glucose (IFG) may increase in patients with severe steatosis, mild weight gain reported in the majority of sustained responders, and to the high BMI of the participants compared to other studies. It is worth mentioning that few patients developed impaired fasting glucose tolerance since their last follow up date which could be due to the effect of IFN therapy that might enhance underlying autoimmunity against beta cells leading to overt type 1 DM [31,32]. The lack of correlation between IR and BMI could be due to the other variables affecting IR such as fibrosis stage, steatosis, GH and IGF-1 levels. Moreover, the number of responders in comparison to non-responders also affected the statistical analysis. On the contrary to our results, several investigators have demonstrated that obesity and steatosis were related to IR in chronic HCV infection [7,8] and that weight loss could improve IR in these patients [33]. Furthermore, the correlation of IR with steatosis severity, BMI [34] could explain the elevated prevalence of impaired fasting glucose [35] and DM in CHC patients compared to the general population.

In the present study, there was a lack of correlation between HOMA-IR and fibrosis stages in the recruited patients. The low IGF-1 levels that might reflect severe chronic liver disease were significantly correlated with HOMA-IR in responders and non-responders. Previous data illustrated the contribution of IR to the progression of liver fibrosis [8,36-38]. IR was considered a major independent determinant of fibrosis regardless of the genotype and the severity of liver damage [39]. Mean HOMA index increases with the stage of fibrosis [7] and could help to differentiate stages of fibrosis. The presence of steatosis is responsible for causing progressive fibrosis in addition to the role of intrahepatic inflammation [40,41].

In the current study, HOMA-IR score failed to show a significant difference between responders and non-responders. However, both SVR and HOMA-IR were associated with increased GH levels while HOMA-IR was significantly correlated with the low IGF-1 levels. The low IGF-1 levels were negatively correlated with serum insulin, reflected an advanced fibrosis score and were able to display the presence of GH resistance in presence of high GH levels.

Similar to our results, Plöckinger et al., [42] reported high GH levels in treated HCV patients with low IGF-1 concentrations indicating the existence of GH resistance of hepatocytes and that high amounts of GH can lead to IR [13].These GH levels following therapy may be due to a direct drug effect or related to the suppression of viral load. Furthermore, IGF-1 gene expression was down regulated in progressive chronic liver disease and accordingly serum IGF-1 level decreases [43].

Our results are in general agreement with other studies. Most researchers documented GH resistance in HCV even in the phase preceding cirrhosis [12] as evident by low IGF-1 levels in the presence of normal or increased GH secretion. About 90% of circulating IGF-1 originates in the liver. In liver fibrosis, disturbed synthesis of IGF-1 reflects the severity of the clinical stage and could repre-
sent a good, noninvasive marker of liver fibrosis. This is in accordance with our study where IGF-1 levels were associated with advanced stages of fibrosis. Others causes contributing to low IGF-1 in adults are malnutrition and diabetes mellitus [44] and it is worth mentioning that none of our patients were diabetic and none of the participants showed clinical evidence of protein-calorie malnutrition or signs of vitamin deficiency. IGF-1 was found to decrease serum insulin [45]. We were able to demonstrate a similar negative correlation between IGF-1 and insulin.

Our results concerning the HOMA-IR are in partial agreement with previously published data which could be due to different population studied as most Egyptian HCV patients carry genotype 4 [46] and also the absence of baseline HOMA-IR figures before therapy to be compared with scores taken during and after therapy.

Genotypes were found to behave differently concerning IR, steatosis, fibrosis and response to treatment. In genotype 1, IR has been associated with steatosis development and fibrosis progression [47] and SVR rate was twice in patients with HOMA ≤ 2 compared to those with HOMA ≥ 2 [48]. While in genotype 3, viral induced hepatic steatosis is likely to be due to direct cytopathic effect of HCV independent of IR [49]. Previous data suggest that IR at baseline has been linked to treatment failure [2,50,51]. Thus further studies assessing baseline and follow up score of HOMA-IR might be of great value to monitor the effect of IR on treatment outcome.

6. Conclusions

Patients with chronic HCV infection exhibit insulin resistance and low levels of serum IGF-1 following combined IFN/ribavirin therapy irrespective of treatment response. Low IGF-1 levels might reflect the severity of chronic liver disease in addition to an increased HOMA-IR and thus can be postulated to be of pathogenic etiology in the development of IR in patients with CHC.

7. Acknowledgements

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8. References


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Insulin resistance (IR), chronic hepatitis C (CHC), hepatitis C virus (HCV), growth hormone (GH), insulin resistance index (HOMA-IR: homeostasis model of assessment), body mass index (BMI), tumor necrosis factor α (TNF-α), insulin-like growth factor 1 (IGF-1), diabetes mellitus (DM), interferon (IFN), radioimmunoassay (RIA).

Abbreviations

Insulin resistance (IR), chronic hepatitis C (CHC), hepatitis C virus (HCV), growth hormone (GH), insulin resistance index (HOMA-IR: homeostasis model of assessment), body mass index (BMI), tumor necrosis factor α (TNF-α), insulin-like growth factor 1 (IGF-1), diabetes mellitus (DM), interferon (IFN), radioimmunoassay (RIA).