

The Role of Zinc, Manganse and Copper in Rumen Metabolism and Immune Function: A Review Article

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Abstract

Trace minerals are dietary elements required by the body in minute amounts, ranging from 0.10 to 50.0 mg/kg dry matter in beef cattle diets [1]. These trace minerals are all necessary for the biochemical processes of the body that support proper growth and maintenance. For example, Cu is necessary for the function of superoxide dismutase and its removal of toxic byproducts from metabolic pathways [2]. The removal of these toxic byproducts allows for metabolism to proceed efficiently, uninhibited by damaging oxygen free radicals. Zinc, an important trace mineral for enzyme function, aides in the regulation of nucleic acid production, carbohydrate metabolism, and protein synthesis, thus providing a stable framework for development [3]. The immune system is part of the host's defense against destructive forces from outside the body, such as bacteria, viruses, and parasites, or from within, such as malignant cells or those that produce autoantibodies [4]. This system is composed of two branches: the innate or non-specific immune system, and the adaptive or specific immune system [5]. In this review paper, an attempt has been made to review effects of mineral supplements in Rumen Metabolism, effects on Immune Function in different species of animals.

Keywords

Dietary Trace Mineral, Rumen Metabolism, Immune Function, Performance, Reproductive and Rumen Fermentation

1. Introduction

At present time, 111 chemical elements are officially accepted by the International Un-

ion of Pure and Applied Chemistry [6]. Out of them, 93 are classified as naturally occurring, and approximately 50 have been identified as useful to sustain a normal health status in mammals. In addition to the six core elements (C, H, N, O, S and P) that make up nucleic acids, proteins, carbohydrates, and lipids and thus the bulk of living matter [7], many other elements are essential to the nutritional requirements of superior animals. Minerals required in relatively large quantities (g/d) are named "macro" minerals. In opposition, minerals required in mg or µg amounts are referred to as "micro" or "trace" minerals [1]. Minerals are usually classified into four broad groups according to their physiological roles:

- Structural: minerals forming structural components of the body organs and tissues which include Ca, P, Mg, Fl and Si in bones and teeth and P and S in muscle proteins.
- Physiological: minerals occurring on body fluids and tissues as electrolytes, involved in the maintenance of osmotic pressure, acid base-equilibrium, membrane permeability and tissue stimuli (e.g. Na, K, Cl, Ca and Mg in blood and cerebrospinal fluid).
- Catalytic: minerals acting as catalysts in enzymatic systems, as integral compounds in the structure of metalloenzymes or as cofactors required for enzymatic activation (e.g. Fe, Cu, Zn, Mn, and Se involved in cytochromes, ceruloplasmin, carbonic anhydrase, pyruvate carboxylase, and glutathione peroxidase, respectively).
- Regulatory: minerals involved in regulation of cell replication and differentiation (e.g. Ca, in signaling transduction; Zn, in DNA transcription).

However, this classification is arbitrary and not exclusive since the same element can fulfill more than a single function [8].

Ideally, dietary supply of minerals to livestock must be sufficient to ensure the maintenance of body reserves, and to provide adequate concentrations in edible products. However, in some occasions, drinking water can be an exceptionally rich source of minerals, sometimes responsible for mineral toxicity [9]. The accomplishment of an adequate supply of minerals is especially challenging since requirements for most minerals are not constant, but affected by physiological factors including genetics, age, sex, type of production (maintenance, growth, reproduction, and lactation), and level of production [10].

2. Availability of Minerals to Animals

The evaluation of feedstuffs and mineral supplements for the host animal depends not only on the mineral content in the sources but also on the potential availability and absorption of the minerals from the gastrointestinal tract and the mineral's utilization by the animal tissues [11]. However the potentially absorbable fraction of a mineral is greatly influenced by several factor including age and species of animal, intake of mineral relative to amount required, chemical form of the mineral, and the amount and proportions of other dietary compounds involved in specific interactions with the mineral [12]. Livestock diets are often supplemented with trace minerals in the form of inorganic salts, usually oxides, chlorides, sulfates and carbonates. In recent years there has been considerable interest in the use of organic trace minerals in ruminant diets [13]. According to the Association of American Feed Control Officials [14], organic trace minerals in the United States are available in one of the following forms:

- Metal Proteinate: the product resulting from the chelation of a soluble salt with amino acids and/or partially hydrolyzed protein, e.g. Cu proteinate, Zn proteinate, Co proteinate and Mn proteinate.
- Metal Amino Acid Chelate: the product resulting from the reaction of a metal ion from a soluble metal salt with amino acids with a mole ratio of one mole of metal to one to three (preferably two) moles of amino acids forming coordinate covalent bonds. For example, Cu-lysine-sulfate is the sulfate salt of 2:1 molar ratio of L-lysine and Cu. Similarly Cu methionine-bisulfate is the bisulfate salt of a 1:1 molar ratio of DL-methionine and Cu. Other commercially available metal amino acids are Cu amino acid chelate, Zn amino acid chelate and Mn amino acid chelate in addition to chelates for macro minerals like Ca and Mg.
- Metal Amino Acid Complex: the product resulting from complexing a soluble metal salt with an amino acid. Commercially available metal complexes are Zn-methionine, Zn-lysine, Mn-methionine, Fe-methionine and Cu-lysine.
- Metal Polysaccharide Complex: the product resulting from complexing a soluble salt with a polysaccharide solution declared as an ingredient of the formulation (e.g. Cu polysaccharide complex, Zn polysaccharide complex, etc).

While the effectiveness of organic minerals for ruminants has been strongly criticized [9] [15] established that for chelation to be effective, the chelating agent should have a stronger stability for the metal than the metal binding substances in feed, but smaller stability constant than the tissue system where the metal is required. In addition, other factors including metal ion equilibria, kinetic factors, pH gradients and redox equilibrium (in case of redox active metals such as Cu²⁺), may also affect the uptake mechanism of metal ions [16]. Several studies have been conducted to address the benefits of including organic sources of minerals on the productive performance of ruminants. However, in many cases a combination of several organic elements has been used, making difficult to assess which one is responsible for the effects noticed, as discussed later.

The aim of this review is to explore the factors involved in bioavailability of Zn, Mn and Cu at the gastrointestinal tract, affecting the mineral status of the host animal. In addition, the role of Zn, Mn and Cu on ruminal physiology is presented.

3. Zinc

3.1. Interaction of Zinc with Dietary Factors

Early studies from [17] showed that in ruminants, the percentage of dietary Zn absorbed decreases as dietary Zn increases. [18] found a linear increment in endogenous fecal loss of Zn when rats were feed increasing levels of dietary Zn ranging from 0 to 8400 ppm. Similarly, in a study conducted in growing pigs receiving 67 Zn, the addition of phytase increased Zn absorption but also resulted in higher endogenous fecal loss [19]. Concurrently, reductions of urinary and fecal Zn excretion by 48% and 46%, respectively were found in humans when dietary Zn was reduced from 85 to 12 µmol/d. Despite the influence of dietary levels, the requirement of Zn appears to be affected by other dietary factors. However, all of the factors and interactions that play a role on Zn bioavailability are not clearly defined [20]. According to [21], the two major dietary factors affecting Zn bioavailability are the presence of organic chelating agents and the interaction with metallic ions, with Cu and Ca being the most important antagonists.

3.2. Role of Zinc on Rumen Fermentation

Early studies from [22] showed that *in vitro* microbial protein synthesis was increased together with a reduction in NH₃-N concentration when rumen fluid was incubated with additional Zn as $ZnCl_2$ or $ZnSO_4$. According to the authors, this response is due to an effect of Zn in increasing microbial enzymatic activity. However, further studies from [23] showed differences in the behavior of rumen microbial populations in the presence of Zn. While protozoa easily incorporated Zn and were tolerant to high Zn concentrations (25 µg/mL), cellulose degradation from rumen bacteria was deeply depressed, along with bacterial urease activity. In partial agreement, [24] found that addition of 5 μ g Zn/mL of rumen fluid increased cellulose digestion by 24%, but addition of $20 \ \mu\text{g/mL}$ of Zn depressed it by 31%. [25] found that low supplementation levels (10 to 15 ppm Zn in incubation fluid) inhibited *in vitro* urea hydrolysis and retarded ammonia accumulation. Similarly, when Zn was added in vivo at 250 ppm Zn on DM basis, molar proportion of propionate was increased with the consequent decrease in acetate: propionate ratio, and rumen ammonia was decreased due to microbial urease inhibition. However, when Zn was added to achieve a level of 470 ppm on DM basis, a tendency for depressed DM digestibility was noticed. In addition, [26] found a decrease in total VFA when Zn was provided to steers as Zn methionine or Zn glycine compared with $ZnSO_4$ at concentrations closer to physiological levels (20 ppm). In the same study, molar proportion of propionate was increased by Zn methionine and that of butyrate was decreased, with the consequent reduction in the acetate: propionate ratio. The authors conclude that supplementation of Zn methionine may alter ruminal fermentation.

While these differences can be partially explained by the dose of Zn and the fermentation substrate used by the different authors, an alternative mechanism is proposed by [27], who found that addition of 50 μ g/mL of Zn to *in vitro* incubations decreased cellulose digestion at 24 h, but not at 48 h, resulting in an overall decrease in the rate, but not the extent of digestion. The authors conclude that the initial decrease in cellulose digestion might be related to a direct effect of Zn on inactivation of bacterial cellulase, since heavy metal salts can precipitate and denature soluble proteins and enzymes. However, enough cellulase activity may be present to overcome those negative effects of high Zn concentrations. In addition, the accumulation of Zn in bacterial wall [28] might be affecting the adhesion of microbial cells to cellulose particles, a limiting step in cellulose fermentation as previously established [29].

3.3. Zinc Availability from Organic and Inorganic Sources

There is scientific evidence showing that organic Zn is metabolized differently than in-

organic sources. In four consecutive studies conducted by [30], Zn was better retained when added as Zn methionine than ZnO in lambs and heifers. However, the observed improvement was not due to higher absorption but to a lower urinary Zn excretion in animals receiving Zn methionine, and only minor changes in blood parameters were noticed. Similarly, in a study conducted by [31] in calves, organic or inorganic Zn supplementation did not affect concentration of serum enzymes (alkaline phosphatase, glutamate oxaloacetate transaminase, glutamate pyruvate transaminase and super oxide dismutase) or mean concentrations of different serum vitamins (retinol, β -carotene, a-tocopherol) and hormones (triiodothyronine, thyroxin, insulin and testosterone). Furthermore, blood parameters from supplemented groups did not differ from unsupplemented controls. In a comparative study, [32] found no differences in liver Zn and plasma Zn concentration of steers receiving ZnSO₄ or Zn amino acid complex. However, confounding effects may account for these findings, given the differences in mineral status of the animals at the beginning of the study. In partial agreement, no differences in plasma Zn concentration of steers receiving ZnSO₄, Zn methionine complex or Zn glycine were found by [26]. Although large variations prevented from obtaining differences in absorbed or retained Zn, Zn glycine resulted in larger liver Zn concentrations. Conversely, using organic and inorganic Zn. [33] found higher concentrations of Zn in plasma of supplemented beef steers than in controls, despite the Zn source. Interestingly, in animals receiving an implant containing estradiol benzoate and testosterone propionate, weight gain resulted higher when Zn was supplemented as ZnSO₄ that of Zn propionate [33]. In a large production study including 250 dairy cows, only a tendency for improvement in milk production was obtained when Cu, Zn, Mn and Co as sulfates, were replaced by organic forms of the minerals. Liver concentration of minerals was unaffected by mineral source, but supplementation with organic minerals resulted in increased milk solids and a decreased incidence of sole ulcers [34]. In another large scale study conducted on 573 dairy cows [35], supplementing 75% of the requirement of Zn as Zn methionine achieved the same hepatic Zn concentration than supplementing 100% of the requirement as $ZnSO_4$. Based on the lack of differences in health and productive performance the authors suggest that mineral content of liver is not an accurate predictor of cow's response to different sources and levels of trace minerals. In agreement, [34] suggest that Zn, Mn, and Cu content of liver is a poor indicator of trace mineral status. According to [2], the supposed benefits of organic sources of Zn on Zn availability claimed in studies conducted in monogastrics cannot be adopted in ruminants since phytic acid, a major antagonist of Zn absorption, is largely hydrolyzed in the rumen. An additional difficulty in the evaluation of Zn availability from different sources relies on the fact that Zn is absorbed according to the animal needs and homoeostasis in ruminants is achieved primarily by control of intestinal absorption [36].

4. Manganese

4.1. Interaction of Manganese with Dietary Factors

According to the [1], there is no exact data regarding the maintenance requirements of

Mn in dairy cattle. However, the coefficient of intestinal absorption for Mn in adult cattle is known to be as low as 1% of ingested Mn or even lower (37; 20), but the absorption in young calves is considerably higher [37]. Despite the generalized idea of poor Mn absorption, [8] suggested that this situation is partly a reflection of the substantial surplus of Mn provided by most practical rations, since higher coefficients of absorption were obtained when animals received diets marginal in Mn [38]. For this reason, the [1] adopted a conservative coefficient of 0.75% for Mn absorption. In agreement, [39] reported a coefficient of 0.54% for Mn absorption in dairy cows. The study of dietary factors influencing Mn bioavailability has received little attention, probably because Mn deficiency is not considered to be a major problem in ruminants [20]. In addition, most of the information available has been generated in monogastric models. [40] observed a 10% and 13% increase in kidney and bone Mn, respectively from chicks receiving 12 ppm of virginiamycin in the diet. In a further study [41], the addition of 4 ppm lincomycin resulted in higher concentrations of Mn in bone. However, while virginiamycin and other antibiotics are currently used as feed additives for ruminants [42], their role in Mn absorption in cattle remains unknown. According to [43], the intestinal absorption of Mn is negatively affected by dietary levels of Ca and P. Similar results were found by [44], who found a 45% reduction in Mn in the tibia of chicks fed excess Ca and P. However, further evidence provided by [45] indicated that while P has a negative effect on Mn absorption, no deleterious effects on Mn metabolism are obtained with excess of dietary Ca, but effects of Ca and P are difficult to distinguish since diets are usually enriched with both minerals to maintain a Ca:P physiological ratio [9]. There have been no reports relating Mn absorption with forage composition in ruminants, but phytate and fiber are known to be the main antagonists in monogastrics species including human [46], swine [47]. Through microbial activity, both phytates and fiber are broken down in rumen [48]. For that reason, [9] suggests that Mn absorption in ruminants may not be affected by the presence of phytates, being higher than that usually reported for monogastrics.

4.2. Role of Manganese on Rumen Fermentation

Limited information is available regarding the role of Mn in rumen fermentation. According to [49], Mn has a slight stimulatory effect on urease activity. [25] reported a 6% increment in IVDMD when Mn was added to incubations at a dose of 100 ppm. In a previous study, [50] found a reduction in cellulose digestion when Mn was omitted from *in vitro* incubations. However, [24] reported that cellulose digestion peaked at Mn concentrations of 10 to 20 ppm but was completely abolished when Mn was added at 300 ppm. Using 54 Mn, [51] [52] observed that Mn accumulation was higher in rumen bacterial cell walls than in cytoplasm, and that the uptake was similar in bacteria and protozoa, but the biological implications of this fact are not established. [53] fed ram lambs diets that contained from 13 to 45 mg of Mn/kg DM during 84 days. While the number of rumen bacteria was not affected by Mn, the large rumen bacteria (identified as those with a diameter of 12.9 to 16.2 μ m) resulted lower with the lowest Mn intake

and highest with dietary Mn provided at 30 mg/kg. This may be particularly relevant since large rumen bacteria contain more protein than small rumen bacteria [54]. However, despite this variation in microbial populations, no effect of Mn on DM digestibility was observed. [55] suggested that sheep consuming a diet high in fiber and low in protein may respond to Mn supplementation in excess of 36 μ g/g DM, but Mn requirements of the rumen microbes may be increased by the consumption of low quality roughages. In agreement, [56] suggest that the optimum content of Mn in the diet may be as high as 120 μ g/g DM on the basis of results from *in vitro* studies.

4.3. Manganese Availability from Organic and Inorganic Sources

The aim of mineral supplementation is to increase the biological availability of the target mineral(s), defined as the degree to which an ingested element is absorbed and can be utilized in metabolism by the animal [57]. Different sources of Mn are currently available as supplements for animal diets. Among the inorganic sources, the most commonly used include manganese carbonate (MnCO₃), hausmannite (Mn₃O₄), manganese oxide (MnO), manganese dioxide (MnO₂), manganite (Mn₂O₃), manganous chloride (MnCl₂·4H₂O) and manganese sulfate (MnSO₄) [8] [58] [59] [60]. Conversely, sources of Mn classified as "organic" include Mn-methionine, Mn-proteinate and Mn-polysaccharide [42] [61] [62] [63]. Unfortunately, only few studies have been conducted comparing the relative bioavailability of Mn sources in ruminants fed physiological concentrations of Mn [64]. According to [65], some chelates and complexes may improve the mineral bioavailability above that of soluble inorganic forms, as later shown by [42] in lambs comparing Mn-methionine with MnO. However, no differences were obtained in the same study when Mn-methionine was compared with MnSO₄. Similarly, [66] compared the bioavailability of Mn from different organic sources and MnSO₄ in broilers. The authors concluded that only organic Mn sources with moderate or strong chelation strength can provide higher relative bioavailabilities due to their ability to resist Ca antagonisms during the digestion process. In addition, studies conducted on female chicks showed an increase in Mn retention from a Mn-methionine chelate compared with MnO [67].

5. Copper

5.1. Interaction of Copper with Dietary Factors

The amount of dietary Cu needed to supply Cu requirements for maintenance, growth and lactation varies with the age of the animal, the chemical form of dietary Cu and the presence of dietary substances interfering with Cu absorption [1]. Compared with monogastrics where Cu is fairly well absorbed (30% - 75%), absorption in adult ruminants is low, ranging from 1% to 10% of dietary Cu [8] [68]. However, before developing a functional rumen, Cu absorption in lambs can be as high as 70-85% of the dietary supply [69]. The reason for this decrease in Cu absorption appears to be related to interactions occurring at the rumen environment, including the Cu-S-Mo [9] [70] [71], Cu-S [72] [73], and Cu-Fe [74] [75] antagonisms. More recently, results relating high

levels of dietary Mn with Cu deficiency have been reported [76] [77].

5.2. Copper-Molybdenum-Sulfur Interaction

In the presence of ruminal H^+ ions, dietary S is reduced to sulfide, which then reacts with Mo [78] to form different thiomolybdates (mono-, di-, tri-, tetra-thiomolybdates) according to the following reactions:

$$MoO_{4}^{2^{-}} + H^{+} + HS^{-} \leftrightarrow H_{2}O + MoO_{3}S^{2^{-}} \quad (monothiomolybdate)$$
$$MoO_{3}S^{2^{-}} + H^{+} + HS^{-} \leftrightarrow H_{2}O + MoO_{3}S_{2}^{2^{-}} \quad (dithiomolybdate)$$
$$MoO_{2}S_{2}^{-} + H^{+} + HS^{-} \leftrightarrow H_{2}O + MoO_{3}S_{3}^{2^{-}} \quad (trithiomolybdate)$$
$$MoO_{3}^{2^{-}} + H^{+} + HS^{-} \leftrightarrow H_{2}O + MoS_{4}^{2^{-}} \quad (tetrathiomolybdate)$$

In the gastrointestinal tract, thiomolybdates have been shown to bind Cu preventing its absorption, while increasing the Cu fraction associated with the solid phase of the rumen content at expense of a reduction in the fluid phase. Thiomolybdates associated with solid rumen digesta (bacteria, protozoa, and indigested feed particles) form insoluble complexes that do not release Cu even under acidic conditions like the abomasal environment [79]. In addition, absorbed thiomolybdates have also been shown to cause systemic effects on Cu metabolism including increased biliary excretion of Cu from liver stores, strong binding of Cu to plasma albumin resulting in reduced availability for biochemical processes, and inhibition of Cu dependent metalloenzymes such as ceruloplasmin, diamine oxidase, cytochrome oxidase, ascorbate oxidase and tyrosine oxidase. [20] [78] [80]. According to [20], when rumen sulfide concentrations are low, Mo has little effect on the formation of thiomolybdates. However, Cu bioavailability is deeply reduced (up to 70%) when Mo levels are not modified but sulfide concentrations are increased [81].

5.3. Copper-Sulfur Interaction

In addition to its role in the Cu-Mo-S interaction, organic or inorganic S can also reduce Cu bioavailability per-se [21]. [82] found a 55% reduction in hepatic Cu when sheep were fed high levels of S (2 g/kg DM). According to the authors, this reduction was due to formation of CuS in the digestive tract, since the diet was very low in Mo. Similar results were previously observed by [72] who found a 39% - 56% reduction in Cu bioavailability when S was provided to ewes as methionine or as NaSO₄ under low Mo dietary levels, possibly through the formation of insoluble CuS at sites beyond the rumen. However, [83] postulates that the formation of insoluble CuS and Cu₂S in the rumen is exacerbated by the digestion of insoluble proteins by protozoa, with the consequent increase in available S. Despite the effect of dietary S previously mentioned, other S sources have been also responsible for decreasing Cu bioavailability in ruminants. Molasses, a by-product from the sugarcane and beet industry, is a source of dietary sugars feed to dairy cows [1]. Benefits of adding molasses to diets include increasing palatability, acting as a binder, and reducing dust in fine-particle feeds [84]. However, due to its high content of S, the liberal use of molasses can result in dietary S levels considerably in excess of requirements [85]. [73] evidenced a decrease in liver Cu at 29, 56 or 84 d after feeding heifers with a molasses based supplement. According to the authors, this observation was the result of high concentrations of S naturally found in molasses.

In a review of Cu antagonists in cattle, [86] describes other sources of S implicated in the Cu-Mo-S and Cu-S interactions, included fertilizers, high S water, and S containing supplements. Cows grazing bahiagrass pastures fertilized with ammonium sulfate showed lower liver Cu concentrations compared with cows on non fertilized pastures, or fertilized with ammonium nitrate [86]. A previous study from [87] showed that gypsum fertilization (132 kg S/ha) increased S from 0.33% to 0.40% and from 0.29% to 0.37% of DM in tall fescue grass and in orchadgrass, respectively. However, feeding those pastures to steers resulted in no changes in Cu bioavailability, probably due to the high S content of the non fertilized pastures. For this reason [73] suggest that the choice of fertilizer source can be critical in areas where grazing cattle may be prone to Cu deficiency Sulfur levels in drinking water can also be detrimental for Cu bioavailability. [88] reported a decrease in plasma and hepatic Cu of yearling steers provided with high-S water (3651 mg of SO₄/L) compared with those receiving low-S water (566 mg of SO₄/L). Similarly, a decrease in hepatic Cu of growing steers was found by [89] when S content in the drinking water was increased from 404 to 4654 mg of SO₄/L. While the mentioned S concentrations are in excess of that commonly found in water for animals, high S-containing water has been reported in USA and Canada [90] [91].

5.4. Copper-Iron Interaction

Ruminants consuming forage-based diets are often exposed to high levels of Fe through water, forage, and unusually high amounts of soil ingestion [69] [92]. Supplementing 800 mg of Fe/kg of DM as FeO or $FeSO_4$, decreased Cu absorption from 0.06 to 0.04 in sheep [93]. In agreement, a previous study of (94) found a rapid decrease in liver and plasma Cu concentrations, activities of erythrocyte superoxide dismutase and plasma ceruloplasmin of young heifers receiving 800 mg of Fe/kg of DM. However, according to [94] the role of Fe on Cu absorption is partially dependent of S. Indeed, [81] suggest that the formation of FeS in the rumen is a critical step for Fe to antagonize Cu absorption. An alternative explanation is provided by [95] who demonstrate that excess Fe can compete with Cu for its absorption at intestinal level, by saturating the DMT-1 Cu transporter.

5.5. Role of Copper on Rumen Fermentation

In a production study conducted on beef steers [96], the addition of 20 or 40 mg of Cu/kg of DM decreased animal performance, compared with animals receiving a basal diet containing 10.2 mg of Cu/kg of DM, suggesting that high dietary Cu may inhibit ruminal fermentation. Previously, [97] found a decrease in post feeding total VFA concentration and VFA molar proportions of yearling steers receiving a high dose of sup-

plemented Cu (57.3 mg/kg of DM), but average daily gain, feed efficiency, and carcass yield and quality grade were not affected. In agreement, [98] found an in vitro depression in rumen fermentation of concentrates following the addition of high doses of $CuSO_4$. Similarly, [99] found a reduction in propionate molar proportion when high doses of Cu were added to in vitro rumen incubations. A dose-response study conducted by [100] determined that 21 µg of Cu/mL incubation fluid was required to obtain a 50% inhibition of gas production. However, a large disparity was obtained regarding the susceptibility of the different bacterial populations to Cu. While the growth of Bacteroides succinogenes, Ruminococcus albus and Butyrivibrio fibrisolvens was inhibited by 10, 20 and 30 µg Cu/mL of incubation fluid, respectively, higher concentrations were required to inhibit Megasphaera elsdenii, Selenomonas ruminantium, and Streptococcus bovis (100, 100 and 250 µg Cu/mL, respectively). Opposite results were reported by [101] who found a reduction in rumen pH and an increase in total VFA concentrations when Cashmere wether goats received supplementary Cu in the diet. According to the authors, an increase in NDF digestion may be responsible for these findings. Conversely, later studies conducted by [102] [103] found that NDF digestion was unchanged or maximized by the addition of 10 mg of Cu/kg of DM, but depressed when of 30 mg of Cu/kg of DM were added. In addition, no differences were obtained for rumen pH, IVDMD and VFA molar proportions when in vitro studies were conducted using rumen fluid donors receiving 0, 10 or 20 mg of Cu/kg of DM [104]. Based on previous observations showing that 20 or 40 mg of Cu/kg of DM increased unsaturated fatty acids in adipose tissue of steers [104], a possible role of Cu as an inhibitor of ruminal lipids biohydrogenation has been suggested. [105] However, no studies have been conducted to assess the validity of this hypothesis.

5.6. Copper Availability from Organic and Inorganic Sources

The effectiveness of organic sources of Cu to promote animal benefits has been a subject of several controversies. [106] reported a higher retention of Cu in steers supplemented with Cu-lysine compared with supplementation with CuSO₄. In another study [107], the use of organic Cu (as Cu proteinate) increased hepatic retention of Cu in multiparous beef cows, compared with inorganic Cu. However, no benefits on cow and calf performance were obtained. Conversely, [108] found an increase in body weight gain in goat kids supplemented with organic Cu, compared with inorganic Cu. In a recent meta-analysis [13] assessing the benefits of organic trace minerals, only marginal improvements in milk production, milk fat and milk protein were found. In opposition, organic trace minerals did not affect somatic cell count, interval from calving to first service, and 21-d pregnancy rate. Similarly, no differences in 60-d pregnancy rate, health or performance were found by [109] in 2-year old cows receiving Cu as $CuSO_4$ or as amino acid complex. In addition, [110] found a decrease in pregnancy rates of primiparous cows receiving organic and inorganic minerals (Cu, Co, Mn and Zn) compared with non-supplemented cows. According to the authors, excessive supplementation beyond requirements reduced reproductive performance. In a previous study conducted on steers [71], growth rate was higher when animals received $CuSO_4$ than when Cu-lysine was provided during the initial 21 d, but no differences were obtained after 98 d. Other parameters, including feed efficiency, feed intake, humoral and cellular immune response and ceruloplasmin activity, were not affected by Cu source [71]. Organic sources of Cu have been seriously criticized by [8] who consider that technologies for protection against rumen antagonisms are extravagant and provide no additional benefits than conventional CuSO₄. In addition the authors questioned the scientific legitimacy of some *in vivo* studies where no proper covariance analyses were conducted to account for initial differences between groups of animals [8]. According to [9] commercially driven pursuit of trivial advantages over cheap and effective inorganic sources of Cu should cease, and attention should focused on predicting when supplementation is needed and to the usual problem of over, rather than under-provision of Cu and its environmental impact. In conclusion, Cu, Zn and Mn are required to maintain health and production status of livestock, but their functions at the gastrointestinal tract of ruminants are not totally elucidated. Like many other minerals, Cu, Zn and Mn have the ability to interact with organic compounds of the diet, macro minerals and micro minerals, usually resulting in decreased availability for the host. Different technologies, including proteinates, amino acid chelates, amino acid complex, and polysaccharide complex are currently available for mineral protection However, these technologies appear to be more effective in monogastrics than in ruminants. Mechanisms to guarantee optimal levels of ruminally available minerals, and to optimize mineral supply to the lower gastrointestinal tract without compromising postruminal absorption require further research.

6. Trace Minerals Cu, Zn, Mn and Their Effect on Immune Function

Trace minerals are those needed by the body in minute amounts (generally included in the diet in parts per million quantities). Several of these trace minerals such as zinc, manganese, copper, and cobalt are required for the functionality of numerous structural proteins, enzymes, and cellular proteins [35]. Trace minerals may function as cofactors, activators of enzymes, or stabilizers of secondary molecular structure and serve essential functions in cell metabolism [111]. Numerous studies have shown that feeding amino acid complexes of Zn, Mn, and Cu have improved the performance of dairy cattle through improving fertility rates and reducing the incidence of disease [112] [113] [114]. These improvements in animal performance appear to be related to increased availability of trace minerals for metabolism [115] [116].

Ruminants are frequently subjected to severe dietary deficiencies of trace elements such as copper, cobalt, selenium, iodine, manganese, and zinc [37]. These deficiencies have been linked to a decline in fertility from enzymatic dysfunctions. Hypocuprosis in dairy cattle and sheep has been linked to female reproductive disorders such as prevention of embryo implantation and high prenatal mortality, particularly early embryonic loss [37]. Several studies in rats and mice have shown that both cell-mediated and hu-

moral immunity are greatly depressed by copper deficiency [37] [117]. [118] showed that marginal copper deficiency in dairy heifers reduced the capacity of neutrophils to kill S. aureus. Animals deficient in copper also show an increased susceptibility to bacterial pathogens. This has been attributed to the role of copper in superoxide dismutase and cytochrome c oxidase enzyme systems [119]. [4] reported that copper deficiency impaired the ability of macrophages to kill yeast cells. [120] demonstrated that copper-depleted calves exhibited impaired phagocytic killing activity that was reversed by copper supplementation. In another study, low copper status was associated with a reduced response of peripheral-blood lymphocytes to stimulation with T-cell mitogens [121]. Despite these studies, the overall effect of copper deficiency on macrophage function in cattle has not been studied extensively [122].

Extensive research conducted on human subjects and laboratory animals suggests that zinc deficiency reduces immune responses and disease resistance [123]. In children, zinc deficiency has been shown to affect T-lymphocyte and neutrophil function along with reduced proliferation of lymphocytes in the presence of mitogens and slower neutrophil chemotaxis [119]. Zinc deficiency also produces atrophy of the lymphoid tissues such as the thymus. Zinc deficiency also negatively impacts phagocyte function resulting in decreased ingestion and phagocytosis [124]. A study conducted in laboratory animals fed a moderately zinc deficient diet showed that the differentiation and function of B-cells may be impaired [125]. In cattle, surprisingly little research has been conducted to examine the relationship between dietary zinc and immune function. Marginal zinc deficiency appears to have marginal effects on immune function in ruminants, but research also suggests that the addition of zinc to practical diets may affect disease resistance [22]. Zinc deficiency is most deleterious to the reproductive function of male animals; however, the administration of a zinc supplement to cattle was shown to increase conception rate by 23% compared to controls, and discontinuation of this supplement resulted in decreased conception rate [126].

Manganese deficiency has been linked to suppression of estrus, reduction in conception rates, increased incidence of abortions, and low birth weights. In dairy cattle, the main clinical sign of restricted manganese intake is anestrus or irregular return to estrus sometimes with extended periods of anestrus [127]. This leads to depressed conception rates [37]. Experimental animals fed a manganese deficient diet have been shown to have deficient antibody synthesis and secretion [128]. After adding manganese to the diet, antibody production improved. The mechanism(s) by which manganese affects antibody synthesis or release has not been clearly elucidated and further study in this area is needed [128].

7. Conclusions

In this review paper, we conclude the following:

1) Trace elements are essential for health, growth, production and reproduction. They are essential for functioning of a number of components of the immune system. Thus, they contribute to maintaining proper health and immunity. They are important for functioning of a number of enzymes and proteins which are involved in many physiological and biochemical processes.

2) Essential trace minerals such as zinc, copper and manganese play a wide variety of biological and physiological roles in animal development and health. These minerals take part in the antioxidant defense, tissue development, and immune function.

3) There is almost a consensus that organic trace minerals have higher bioavailability, resulting in better animal performance, health, production immune response and stress alleviation than their inorganic salts.

4) Quantifying the requirements for trace minerals of dairy cows is extremely difficult and the methods and models currently used may not be appropriate because of the different metabolic functions of trace minerals. There is also lack of proper acts and regulations for monitoring their quality for commercial marketing.

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