Comparison of the Insecticidal Activities of Three Plants against Two Devastating Insects: *Callosobruchus maculatus* and *Sitophilus zeamais*

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**Abstract**

To find the alternate ones with insecticides of synthesis, the extracts of plants are more and more used by the peasants to protect stocks from harvest against the devastating insects. Small fireclay cups phytochimic and insecticidal are made on three plants (*Crataeva religiosa*, *Ficus thonningii* and *Tapinanthus bangwensis*). The identification of the active ingredients present in the three plants show that they contain polyphenols, flavonoides, alkaloids, tannins, saponosides, etc. The insecticidal tests showed that the extract with the chloroform (HEY) of *Crataeva religiosa* is effective on *Callosobruchus maculatus* and *Sitophilus zeamais* while extracts with cyclohexan (EC) and methanolic (EM) are effective only on *Callosobruchus maculatus*. It sucked shown that the extract with chloroform gives a better mortality rate and that *Callosobruchus maculatus* is more sensitive to the treatments the strong amount (50 mg/mL). For the study of emergences, the methanolic extract of the sheets of *Crataeva religiosa* ensures a better protection of the medium on *Sitophilus zeamais*. It was also shown that the extracts with the cyclohexane and methanolic of *Ficus thonningii* are more effective on *Callosobruchus maculatus*. Thus, the methanolic extract (EM) of *Ficus thonningii* reduces considerably the number of insects emerged in the short run on *Callosobruchus maculatus*. The tests relating to *Tapinanthus bangwensis* showed that the extracts cyclohexanic, chloroformic and methanolic are effective on *Sitophilus zeamais*. The extracts with the cyclohexan and the methanol of *Tapinanthus bangwensis* reduce considerably the number of insects emerged on *Sitophilus zeamais* and long-term. Broadly *Tapinanthus bangwensis* is more effective on the two studied insects and *Callosobruchus maculatus* is the species most sensitive to the treatments. It is thus the most promising plant.
1. Introduction

The zones of food insecurity in the world are essentially concentrated inside the tropics. However, these zones neither are exploited nor most fertile of the sphere. To cope with the increasing food request of the populations living in these areas, there exist three possibilities: to slow down the population growth; to increase the agricultural production by increasing the cultivable outputs or surfaces; to reduce the losses before and after harvests [1].

Between harvest and consumption, more than 30% of the production is lost. This proportion is stronger in sahelian area because of long period of storage [2].

If no protection is made, after seven months of storage, the loss of the food products can be total [3].

To fight against the devastating insects of stocks, the most used method is the use of insecticides of synthesis. According to Isman [4], several developing countries still make recourse to insecticide DDT and other persistent organic pollutants (POP). These pesticides of long persistence ensure the seed protection since the storerooms until in the fields after sowings as well as young seedlings against the insects and the diseases. This causes not only problems of resistance in the devastating insects but, would involve also adverse effects on the environment and the human health.

In the search for alternative methods of fight, the vegetable kingdom offers many possibilities. Many studies currently develop to insulate or identify secondary metabolites extracted from plants which have an insecticidal activity, repulsive or no appetizing with respect to the insects [5].

It is accordingly that our work is registered. Indeed, the insecticidal effect of nine organic extracts (cyclohexanic, chloroformic and methanolic), resulting from three plants, (Crataeva religiosa, Ficus thonnigii and Tapinanthus bangwensis) is tested under the conditions of laboratory on Callosobruchus maculatus (devastating insect of stocks of niebe) and Sitophilus zeamais (devastating insect of corn stocks). These three plants are listed at the time of an ethnobotanic investigation carried out in the department of Mbour. They are used by the local populations in the use and crop protection other.

2. Materials and Methods

2.1. Material

2.1.1. Plant Material

Vegetable material was collected in the rural community of Keur Balla in the depart-
ment of Mbour, dried in the shade and safe from the light before being conveyed in
Dakar for the study. It comprises various types of drugs (barks of stem, sheets) which
were crushed and used for the extractions with solvents of gradient of increasing polar-
ities. Vegetable material consists of sheets of Crataeva religiosa, Tapinanthus bangwen-
sis and Ficus thonningii.

2.1.2. Biological Material
Biological material relates to Sitophilus zeamaïs and Callosobruchus maculatus ob-
tained by breeding of mass.

2.2. Methods
Extraction and Breeding of Mass
1) Extraction
The technique used for the extractions is the maceration during 24 hours with sol-
vents of gradient of increasing polarity (Cyclohexan, administers chloroform to and
Methanol). Indeed, the samples are impregnated in solvents (1 g/10mL) of increasing
polarity during 72 hours. The extract obtained is concentrated using a rotary evapora-
tor before being dried with the room temperature (25˚C) and safe from the light during
24 to 48 hours. Thus, the diagrams of extraction for the various plants are represented
below.
2) Breeding of mass
The breedings of mass relate to Callosobruchus maculatus and Sitophilus zeamaïs.
Niebe infested was bought at the market on which sorting were operated to recover
insects and to launch the breedings of mass. The breedings are launched in jars out of
glass of 500 ml of volume approximately. Inside each jar, 20 to 25 insects are intro-
duced and of the impregnated absorbent water cotton to create the conditions of mois-
ture necessary for a good reproduction of the insects. These jars are perforated and
covered with fabric of mosquito net to make it possible the insects to breathe. The
breedings are made in the shade and the room temperature. Emergences were observed
between 17 to 20 days with C. maculatus against 28 days with S. zeamaïs. The tests of
insecticidal activity are carried out on insects of first generation constituted by old in-
ssects between 0 and 24 hours.

2.3. Identification of the Chemical Groups
2.3.1. Thin Layer Chromatography of Alkaloids
For the identification of alkaloids, the silica gel is used like stationary phase. Eluant is a
mixture of Chloroform and diethylamin (45 V/5 V). The witness used is Cinchonin.
The revelation is made with the reagent of Dragendorf. Red or red colouring orange
brand the presence of alkaloids in the extracts. The development is carried out with the
room temperature and the atmospheric pressure.

2.3.2. Thin Layer Chromatography of Tannins
For the identification of tannins, we used like eluant a mixture of water and methanol,
ethyl acetate in the respective proportions of 40 ml, 8 ml and 5 ml. For that, plates out of glass covered with silica gel are used like stationary phase. The revelation is made by a ferric chloride solution after drying. Colouring chestnut of the spots is synonymous with the presence of tannins in the extracts. The chromatography is carried out at a temperature of approximately 25°C and the atmospheric pressure.

2.3.3. Thin Layer Chromatography of the Flavonoids

For the flavonoids, eluant it used is a mixture of water and ethyl Acetate (15%). The stationary phase consists of cellulose. The revelation is made with aluminium chloride and the observation under UV with 254 Nm. Yellow colouring indicates the presence of flavonoids. In margin of the identification of the flavonoids, could be done that of polyphenols with a direct revelation under UV without use of reagents. Thus, several luminescences are observed with various colourings with the room temperature and safe from the light which show the presence of flavonoids.

2.3.4. Identification of the Saponosids

To search the saponosids, 10 ml of the aqueous total extract are versed in a test tube. The tube is agitated during 15 s then left at rest lasting 15 mn. A height of persistent foam, higher than 1cm would indicate the presence of saponosids.

2.4. Biological Test

The tests relate to three plants (Crataeva religiosa, Ficus thonningii and Tapinanthus bangwensis) and two types of insects (Callosobrucus maculatus, ravageur of niebe and Sitophilus zeamais, ravageur of corn) and two speculations (niebe and maize). Three extracts of each plant are tested on the two types of insect. Starting from each dry extract, we prepared five solutions with increasing different amounts (100 mg/ml, 2: 50 mg/ml, 3: 25 mg/mL, 4: 12.5 mg/mL and 5: 6.25 mg/mL). Solution 5 is obtained by taking 1 g of dry extract which one dissolves in 10 ml of solvent. Solution 4 is obtained by piping 5 ml of the solution 5 which one supplements to 10 ml with solvent. With the same process, we obtained solution 3 starting from solution 4; 2 from the 3 and 1 from the 2.

The biological tests are carried out in limp of Petri of diameter 90mm. In each limps, one put 20 g speculation (niebe or maize). The tests are carried out by pulverizing 500 µl of each solution in the limps of petri thanks to a Pasteur pipette. The test is repeated five times for Sitophilus zeamais and four times for Callosobruchus maculatus. Le The whole is then left with the free air during 20mn to allow the evaporation of solvent. The insects are introduced thereafter into each limp. On the whole, we used 1215 experimental units to evaluate the toxicity of all the extracts on the insects is 135 units per extract (135 × 9 = 1215). The dead insects are sorted and recovered using flexible grips. The number of died, alive and emerged insects are then counted. The formula of Abboth: Mc = (Mo − MT)/(100 − MT) × 100; (with Mc: calculated mortality, Mo: mortality observed and MT: mortality in the pilot batches) is used to correct mortality observed.
2.5. Statistical Analysis

For the data of the biological tests with the extracts of the plant, the measured variables are the number of died insects, the number of surviving insects and the number of emerged insects. Calculated mortality was obtained by applying the formula of Aboth (1925): \( \text{Mc} = \frac{(\text{Mo} - \text{MT})}{(100 - \text{MT})} \times 100; \) (where \( \text{Mo} \) = mortality in the treated batches, \( \text{MT} \) = mortality in the witness and \( \text{Mc} \) = calculated mortality). The variables many died insects, number of the surviving insects and number of the emerged insects are subjected to a variance analysis, model fixed with three factors (extracted, amounts and time). Variable mortality rate underwent a transformation arcsin (\( X = \text{mortality rate} \), \( N = \text{size of the population} \); \( n = 1999 \)) in order to standardize the population and to stabilize the variance. The general method linear model in Minitab 17 was used for the statistical analysis of the collected data. The variables many surviving insects and many insects emerged as for them underwent a transformation square root in order to standardize the population and to stabilize the variance. The curves and the tables are used to have the result of the analysis.

3. Results

3.1. Extraction Results

The results of the extractions of the various plants are in Table 1 following. Table 1 shows that the outputs of the extractions are better for the extracts with the methanol (from 4382 to 11.118%) which is a polar solvent. That shows the preponderance of the polar compounds in the studied plants.

3.2. Phytochemical Study

The results of the tests of identification, by Thin layer chromatography, of the various chemical groups present in the organic extracts of the three plants (Crataeva religiosa, Ficus thonningii, Tapinanthis bangwensis) are in Table 2.

Table 1. Extraction results.

<table>
<thead>
<tr>
<th>Plantes Extracts</th>
<th>Initial mass (g)</th>
<th>Mass of the extract (g)</th>
<th>Yield (%)</th>
<th>Aspect of the extract</th>
</tr>
</thead>
<tbody>
<tr>
<td>( \text{Crataeva religiosa} ) Hexanic</td>
<td>195</td>
<td>2.8</td>
<td>1.436</td>
<td>Powder</td>
</tr>
<tr>
<td>Chloroform</td>
<td>210</td>
<td>2.6</td>
<td>1.238</td>
<td>Powder</td>
</tr>
<tr>
<td>Méthanolic</td>
<td>195.5</td>
<td>18</td>
<td>9.208</td>
<td>Pasty</td>
</tr>
<tr>
<td>( \text{Ficus thonningii} ) Cyclohexanic</td>
<td>64.352</td>
<td>2.733</td>
<td>4.247</td>
<td>Pasty</td>
</tr>
<tr>
<td>Chloroformic</td>
<td>61.619</td>
<td>1.391</td>
<td>2.257</td>
<td>Powder</td>
</tr>
<tr>
<td>Méthanolic</td>
<td>60.228</td>
<td>2.639</td>
<td>4.382</td>
<td>Pasty</td>
</tr>
<tr>
<td>( \text{Tapinanthis bangwensis} ) Cyclohexanic</td>
<td>64.154</td>
<td>2.097</td>
<td>3.269</td>
<td>Powder</td>
</tr>
<tr>
<td>Chloroformic</td>
<td>62.057</td>
<td>1.414</td>
<td>1.279</td>
<td>Powder</td>
</tr>
<tr>
<td>Méthanolic</td>
<td>60.643</td>
<td>6.742</td>
<td>11.118</td>
<td>Pasty</td>
</tr>
</tbody>
</table>
### Table 2. Results of phytochemical tests.

<table>
<thead>
<tr>
<th>Plants</th>
<th>Extrats</th>
<th>Alcaloids</th>
<th>Flavonoids</th>
<th>Polyphenols</th>
<th>Tannins</th>
<th>Saponosids</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Ficus thonningii</em></td>
<td>Cyclohexanic</td>
<td>+</td>
<td>−</td>
<td>+</td>
<td>−</td>
<td>−</td>
</tr>
<tr>
<td></td>
<td>Chloroformic</td>
<td>+</td>
<td>−</td>
<td>+</td>
<td>−</td>
<td>−</td>
</tr>
<tr>
<td></td>
<td>Méthanolic</td>
<td>−</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>−</td>
</tr>
<tr>
<td></td>
<td>Aqueous</td>
<td>−</td>
<td>−</td>
<td>−</td>
<td>−</td>
<td>+</td>
</tr>
<tr>
<td><em>Crataeva religiosa</em></td>
<td>Cyclohexanic</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>−</td>
<td>−</td>
</tr>
<tr>
<td></td>
<td>Chloroformic</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>−</td>
</tr>
<tr>
<td></td>
<td>Méthanolic</td>
<td>−</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>−</td>
</tr>
<tr>
<td></td>
<td>Aqueous</td>
<td>−</td>
<td>−</td>
<td>−</td>
<td>−</td>
<td>+</td>
</tr>
<tr>
<td><em>Tapinanthus bangwensis</em></td>
<td>Cyclohexanic</td>
<td>+</td>
<td>−</td>
<td>+</td>
<td>−</td>
<td>−</td>
</tr>
<tr>
<td></td>
<td>Chloroformic</td>
<td>+</td>
<td>−</td>
<td>+</td>
<td>+</td>
<td>−</td>
</tr>
<tr>
<td></td>
<td>Méthanolic</td>
<td>−</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>−</td>
</tr>
<tr>
<td></td>
<td>Aqueous</td>
<td>−</td>
<td>−</td>
<td>−</td>
<td>−</td>
<td>+</td>
</tr>
</tbody>
</table>

+: présence −: absence.

### 3.3. Insecticidal Effects of the Three Plants on *Sitophilus zeamais* and *Callosobruchus maculatus*

Analysis of the variance of the combined insecticidal effects ([Table 3](#)) watch which the parameter plant is highly significant ($P < 0.001$). In the same way, the interactions plant-insects are highly significant ($P < 0.001$). The factor insect and the interactions plant-extracts, extract-insects and plant-extract-insects are significant ($P < 0.05$). That implies that mortality depends on the plant, the insects and the extracts. [Table 1](#) and [Table 2](#), [Figures 1-4](#) have the results of the tests combined against *S. zeamais* and *C. maculatus*.

**Figure 1** and **Figure 2** indicate the curve of mortality according to the plants, the extracts, the amounts and the insects. The plant *Tapinanthus bangwensis* (P1) is effective on *Sitophilus zeamais* for the extracts with chloroform and methanol. The plants *Crataeva religiosa* (P2) and *Ficus thonningii* (P3) are effective on *Callosobruchus maculatus* and *Sitophilus zeamais* for the extracts with cyclohexane, administers chloroform to and methanol with P2 and for the extract methanolic with P3.

**Figure 3** and **Figure 4** give the curve amongst emerged insects according to the plants, of the extracts, the amounts and the insects. The plant *Tapinanthus bangwensis* largely reduces the number of emerged insects of *S. zeamais* and *C. maculatus* for the cyclohexanic (EC), chloroformic (EH) and methanolic (EM) extracts. The plant *Crataeva religiosa* protects the medium well from treatment against *Sitophilus zeamais* for the methanolic extract.

### 4. Discussion

The results of the tests phytochimic highlight the presence of alkaloids in the hexanic
**Figure 1.** Curve of mortality according to the plants, extracts, amounts and insects.

**Figure 2.** Interaction between plants, extracts, amounts and insects according to mortality.
Figure 3. Curve of insects emerged according to the plants, extracts, amounts and insects.

Figure 4. Interaction between plants, extracts, amounts and insects according to the number of emerged insects.
and chloroformic extracts of *Crataeva religiosa*, *Ficus thonningii* and *Tapinanthus bangwensis*. Flavonoids, polyphenols, tannins and saponosides are also highlighted in the various organic extracts of the various studied plants. Also let us note that the saponosides are only highlighted that in the aqueous extracts.

Several similar studies are corroborated by these results. Among this work, one can quote those of Gowsalya and Saravanababu [6]. They showed the presence of tannins, saponins, flavonoïdes, steroids, glycosides, and alkaloids in the extracts ethanolic, chloroformic and hexanic of *Crataeva religiosa*.

For *Ficus thonningii*, several studies carried out by various authors are confirmed by the got results. Ndukwe et al., [7] showed the presence of carbohydrates, glycosides, saponins and alkaloids in the extract methanolic of the sheets, the stems and the roots of *Ficus thonningii* The studies of Onwkaeme and Udoh [8] highlight the presence of tannins, flavonoïdes, saponins, anthraquinone and glycosides in the sheets of *Ficus thonningii*.

The results of the tests phytochimic carried out on the species *Tapinanthus bangwensis* confirmed the work carried out by several authors. Efuntoye et al., [9] identified in the extracts methanolic and chloroformic sheets of *Tapinanthus bangwensis* the presence of tannins, of alcaloids, saponins and flavonoids. Of the same Ekhaise et al. [10] identified, in the extract methanolic of the sheets of *Tapinanthus bangwensis*, various chemical groups such as saponins, the flavonoïdes, of tannins, the steroids and the glycosides.

**Table 3.** Results of the treatments of the three plants on *Sitophilus zeamais* and *Callosobruchus maculates*.

<table>
<thead>
<tr>
<th>Source de variation</th>
<th>Mortality DL</th>
<th>F</th>
<th>P</th>
<th>Emergency DL</th>
<th>F</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Plants</td>
<td>2</td>
<td>54.60</td>
<td>0.000</td>
<td>1</td>
<td>88.15</td>
<td>0.000</td>
</tr>
<tr>
<td>Extracts</td>
<td>2</td>
<td>0.55</td>
<td>0.576</td>
<td>2</td>
<td>11.51</td>
<td>0.000</td>
</tr>
<tr>
<td>Plants doses</td>
<td>4</td>
<td>0.22</td>
<td>0.923</td>
<td>4</td>
<td>3.84</td>
<td>0.004</td>
</tr>
<tr>
<td>Insects</td>
<td>1</td>
<td>5.16</td>
<td>0.023</td>
<td>1</td>
<td>363.09</td>
<td>0.000</td>
</tr>
<tr>
<td>Plants extracts</td>
<td>4</td>
<td>3.06</td>
<td>0.016</td>
<td>2</td>
<td>24.73</td>
<td>0.000</td>
</tr>
<tr>
<td>Plants doses</td>
<td>8</td>
<td>0.42</td>
<td>0.912</td>
<td>4</td>
<td>1.71</td>
<td>0.145</td>
</tr>
<tr>
<td>Plants insects</td>
<td>2</td>
<td>149.67</td>
<td>0.000</td>
<td>1</td>
<td>58.25</td>
<td>0.000</td>
</tr>
<tr>
<td>Extracts doses</td>
<td>8</td>
<td>0.23</td>
<td>0.985</td>
<td>8</td>
<td>3.05</td>
<td>0.002</td>
</tr>
<tr>
<td>Extracts insects</td>
<td>2</td>
<td>0.97</td>
<td>0.381</td>
<td>2</td>
<td>7.81</td>
<td>0.000</td>
</tr>
<tr>
<td>Doses insects</td>
<td>4</td>
<td>0.42</td>
<td>0.794</td>
<td>4</td>
<td>3.68</td>
<td>0.006</td>
</tr>
<tr>
<td>Plantes extracts doses</td>
<td>16</td>
<td>0.25</td>
<td>0.999</td>
<td>8</td>
<td>3.94</td>
<td>0.000</td>
</tr>
<tr>
<td>Plantes extraits insects</td>
<td>4</td>
<td>2.52</td>
<td>0.039</td>
<td>2</td>
<td>10.62</td>
<td>0.000</td>
</tr>
<tr>
<td>Extracts doses insects</td>
<td>8</td>
<td>0.27</td>
<td>0.976</td>
<td>8</td>
<td>1.57</td>
<td>0.128</td>
</tr>
<tr>
<td>Erreur</td>
<td>4134</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>4199</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
The presence of these various families of compounds is likely to give an explanation of the biological activities of the studied plants.

The insecticidal tests showed that the chloroformic extract of *Crataeva religiosa* is effective on *Callosobruchus maculatus* and *Sitophilus zeamais* while extracts with cyclohexan (EC) and methanolic (EM) are effective only on *Callosobruchus maculatus*. The results also show that the extract with chloroform gives a higher mortality rate and that *Callosobruchus maculatus* is more sensitive to the treatments. Mortality is higher with the 4j, 5j, 6, and 7j. For the study of emergences, the extract methanolic of the sheets of *Crataeva religiosa* ensures a better protection on *Sitophilus zeamais* and long-term (33j, 39j, 44j).

This study also shows that the extracts with the cyclohexan and methanolic of *Ficus thonningii* are more effective on *Callosobruchus maculatus* with 3rd, 4th, 5th, 6th, 7th and 8ej i.e. in the short run.

Thus, the extract methanolic (EM) of *Ficus thonningii* reduces considerably the number of insects emerged in the short run on *Callosobruchus maculatus*. For the tests concerning the insecticidal activities of the organic extracts of the sheets of *Tapinanthus bangwensis*, the extracts cyclohexanic, chloroformic and methanolic of *Tapinanthus bangwensis* are effective on *Sitophilus zeamais*. The curve of mortality according to time shows that mortality rate is high with short (1e, 4th, 5th and 7ej) and with long terms (17th and 18ej). We also note that the extracts with the Cyclohexane and the methanol of *Tapinanthus bangwensis* reduce considerably the number of insects emerged on *Sitophilus zeamais* and long-term.

In addition, it was shown that the studied plants (*Crataeva religiosa, Ficus thonningii* and *Tapinanthus bangwensis*) contain alkaloids, tannins, flavonoids, carbohydrates, glucosides, saponosids and others polyphenols...

The phenolic alkaloids, terpenoids, compounds or the flavonoids are photosynthetats of the plants which variously affect the life or the development cycle of the enemies of the cultures [11].

The higher plants largely produce tannins [12]. Tannins present a toxic direct effect for certain species of insects [13]. Tannins influence the growth, the development and the fertility of several devastating insects. The reduced growth caused by tannins has major drawbacks for the insect, with a lower number of eggs and more a small of eggs, which would affect the survival and the health of the individuals of the subsequent generation [14].

The polyphenols ubiquitaires, in nature, cause a disturbance of the natural motricity of the insect. This one can be fast: as of the first day for quercetin, or later, the fourth day, for the narangine, syringaldéhyde or acid vanillic. It is accompanied in certain cases (cafeic acid and ferulic, vanillin, lutéoline 7-glucoside) by an effect knock down. At the end of eight days all the insects are in a state of coma or dead.

The toxicity of polyphenols is correlated positively by the gravitational power of the compound [15].

Thus the aromatic plants and their allelochimic molecules carry on a double activity:
- on the adults: by a fast toxic action of type inhalatoire (monoterpens) on the one hand, and on the other hand by action, which contributes to the insecticidal activity of the aromatic plant of a less intensity but which is exerted in the duration (polyphenols);
- on the various phases of the reproductive cycle: inhibition of the fertility, ovicide activity and larvicide at the stages neonatal and later.

The identified plants are Angiosperms. The Angiosperms contain alkaloids which are secondary metabolites made up by secondary nitrogen atoms, tertiary or quaternary in their structures [16]. They are métaboliquement active and play a significant role in the physiology of the plants or the organizations. The alkaloids have repulsive properties or anti appétantes with regard to the devastating insects [17].

*Crataeva religiosa* is of the family of Capparidaceae. Several studies showed that species of the family of Capparidaceae showed the insecticidal effect of the organic extracts on the devastating insects of stocks of harvest. Among this work, one can *et al.* quote those of Gueye [18] [19] which showed the insecticidal activity of *Boscia senegalensis* on *Carvedon serratus* (groundnut beetle). It is interesting to mention work of Seck *et al.* [20] which obtained in direct application of fruits and sheets of *B. senegalensis* to amounts from 2% to 4% (P/P) 80 to 100% of mortality of the adults of *C. maculatus* and significantly reduces at the same time the emergence and the damage of F1. Many work also showed that the organic extracts of plants give insecticidal effects on the devastating insects of stored food products. The toxicity of the extracts with organic solvents of *Aforostyrax lepidophilus, Trichilia gilgiana, Drypetes gossweileri* and *Zanha golungensis* with regard to *Sitophilus zeamais, Tribolium castaneum* and *Rhyzopertha dominica* is shown by work of Toumnou [21].

5. Conclusion

The results showed that the species *Tapinanthus bangwensis* is most effective on the two studied insects and that *Callosobruchus maculatus* is the species most sensitive to the treatments. It is thus the most promising plant. In prospect, the extracts from these plants will be subjected to fractionations and purification in order to isolate to its active principle(s).

References


