Comparative Studies on Mineral and Scavenging Ability of Edible and Some Underexploited Wild Beans in Nigeria

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Received 6 January 2016; accepted 22 January 2016; published 27 January 2016

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Abstract

This work was set out to assay for some minerals essential for healthy state and biochemical indices that underlined degenerative diseases in some edible bean cultivar and nearly extinct local wild bean. Against this backdrop, ash composition was determined before Na+, K+, Ca2+, Mg2+, Zn2+, Fe2+, Pb2+ and Cd were determined by Flame Photometer and Atomic Absorption Spectroscopy (AAS) respectively. 2,2-diphenyl-1-picrylhydrazyl (DPPH), Vitamin E and Vitamin C were also assayed to determine the scavenging ability of the bean samples. The proximate ash composition result of unprocessed and malted edible bean IT99K-573-2-1 had the highest ash content value of 6.90 ± 0.01 and 6.92 ± 0.01 respectively. In the bean samples Pb2+ and Cd2+ were not detected. The empirical mineral composition varies across both the wild and edible bean without significant difference (P ≤ 0.05) except IT07K-243-1-10 that had Ca2+ to be significantly higher than other bean samples. K+ was found to be significantly higher in Feregede and IT07K-243-1-10 compared to other bean samples. Changes in the radical scavenging ability of the various sample in this study after malting, showed a slight reduction in DPPH content except for the edible beans—IT04K-333-2 and IT845-2246-4. Well, there was slight reduction of Vitamin E only in Otili, Mucuna and IT99K-573-1-1. Compared to others only Otili and IT845-2246-4 had slight reduction in Vitamin C after malting.

Keywords
Beans, Minerals, 2,2-Diphenyl-1-Picrylhydrazyl (DPPH), Vitamin E, Vitamin C

Subject Areas: Biochemistry

http://dx.doi.org/10.4236/oalib.1102318
1. Introduction

Bean seeds are important staple foods, particularly in developing countries, due to their relatively very low cost, long conservation time, and high nutritional value [1]-[3]. They are second only to grains in supplying calories and protein to the world’s population. They belong to an extremely large category of vegetables, containing more than thirteen thousand species [4][5]. Most of the species are nearly extinct due to their under-exploitation for human use. Despite their small size, beans pack a surprisingly rich and varied array of substances that are vital for good health. Their nutritional contents contribute to many health benefits as well as some factors that can reduce the bioavailability of nutrients to human beings [6][7]. Much interest has been generated in examining some of these compounds with respect to chronic disease prevention such as diabetes and cancer [8]-[11].

Dry beans are an excellent source of the water-soluble vitamins; thiamine and folic acid and a good source of riboflavin and vitamin B6. They are rich source of iron, which makes them ideal for vegetarians who do not get an animal source of iron [12]-[14]. Most of the research on dry beans has been related to varietal selection. The criteria for selection have always been resistance to diseases or yields but nutritional quality. For example, Ba-logun and Fetuga [15] evaluated 16 wild underexploited legume seeds (family: Mimosoideae, Caesalpinoideae, and Papilionoideae) for their proximate composition and mineral content. All the leguminous seeds studied were rich in minerals like calcium, phosphorus, and potassium, with low levels of sodium. However, Ajayi et al., [16] investigated the composition, physicochemical properties, oil, and fatty acid content of 3 underutilized legumes from Nigeria, B. eurycoma, T. indica, and Mucuna flagellipes. Fasoyiro et al., [17] undertook a comparative study on the proximate, mineral, and antinutritional qualities of some underutilized grain legumes of south-western (from Oyo and Ondo States) Nigeria. The legumes included African yam bean (2 types of Sphenotylis stenocarpa), bambara groundnut (Vigna subterranean), pigeon pea (Cajanus cajan), and lima bean (Phaseolus lunatus). However, Nwosu [18] assessed the nutritional composition of Sphenotylis stenocarpa an underutilized legumes, using malting treatment.

This present study on the mineral composition and scavenging ability of dry beans (unprocessed) and malted beans (processed) would therefore be of great interest. The knowledge provided would help to orient the work of investigators involved in varietal selection and also reduce or eliminate anti-nutritional factors to make edible and non-edible legume seeds more acceptable as an inexpensive source of protein.

2. Materials and Method

2.1. Collection of Cultivar

Dry, wild bean—Sphenostyles stenocarpa (Otili), Cajanus cajan (Feregede), Phaseolus lunatus (Pakala), were sourced from the bush of Ado Ekiti environ while the wild type—Mucuna cochinindum (Mucuna), and edible type bean—Phaseolus vulgaris tagged, ITB45-2246-4 were brought from International Institute of Tropical Agriculture (IITA), Ibadan Nigeria. They were collected whole, dry and naked (i.e. without pods), with divers colors (off white, gold, navy blue, black and brown respectively). Authentication (Table 1) was carried out at the herbarium of the Plant Science Department, Ekiti State University Ado Ekiti, Nigeria.

2.2. Proximate Ash Composition

2 g of the sample is weighed into a pre-heated crucible. The crucible is placed into muffle furnace at 400˚C - 600˚C for 4 hrs until whitish-grey ash is obtained. The crucible is then placed in the desiccator and weighed:

\[
\text{%Ash} = \frac{\text{wt. of crucible + ash - wt. of crucible}}{\text{wt. of sample}} \cdot 100\%
\]

2.3. Analysis of Minerals

Flame Photometer was used for analysis of Na and K while Atomic Absorption Spectroscopy was used for analysis of Cl, Fe, Pb, Zn, Mg and Ca.

2.4. Determination of Vitamin E

The method of Pearson as modified by Maciej and Krzysztof [19] was used 1.0 g of bean sample with 10 ml of
Table 1. Identification of beans used in this study and their identification.

<table>
<thead>
<tr>
<th>Samples</th>
<th>Local/cultivar name</th>
<th>Botanical name</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>Otili</td>
<td>Sphenostyles stenocarpa</td>
</tr>
<tr>
<td>B</td>
<td>Feregede</td>
<td>Cajanus cajan</td>
</tr>
<tr>
<td>C</td>
<td>Pakala</td>
<td>Phaseolus lunatus</td>
</tr>
<tr>
<td>D</td>
<td>Mucuna</td>
<td>Mucuna cochinchinum</td>
</tr>
<tr>
<td>E</td>
<td>IT99K-573-2-1</td>
<td>Phaseolus vulgaris</td>
</tr>
<tr>
<td>F</td>
<td>IT99K-573-1-1</td>
<td>Phaseolus vulgaris</td>
</tr>
<tr>
<td>G</td>
<td>IT97K-499-35</td>
<td>Phaseolus vulgaris</td>
</tr>
<tr>
<td>H</td>
<td>IT07K-243-1-10</td>
<td>Phaseolus vulgaris</td>
</tr>
<tr>
<td>I</td>
<td>IT04K-333-2</td>
<td>Phaseolus vulgaris</td>
</tr>
<tr>
<td>J</td>
<td>IT845-2246-4</td>
<td>Phaseolus vulgaris</td>
</tr>
</tbody>
</table>

Absolute alcohol and 20 ml of 1 M alcoholic sulphuric acid was reflux for 45 mins. After cooling, 50 ml of water was added before extraction of the unsaponifiable matter with 5 × 30 ml diethyl ether and dry over anhydrous sodium sulphate. The residue re-dissolved in 10 ml absolute alcohol before taking absorbance at 470 nm against blank containing absolute alcohol.

\[
\text{Concentration of sample (mg/ml)} = \frac{\text{Abs. of sample} \times \text{concentration of standard}}{\text{Abs. of standard}}
\]

2.5. Determination of Vitamin C

The vitamin C content was determined by Benderitter et al., [20] using the ascorbic acid as the reference compound. 200 μl of the extract was pipetted and mixed with 300 μl of 13.3% of TCA (Tri-chloroacetic acid) and 75 μl of DNPH (Di-nitrophenylhydrazine). The mixture was incubated at 37°C for 3 hrs and 500 μl of 65% H₂SO₄ was added. The absorbance was read at 520 nm.

\[
\text{Vit. C mg/ml} = \frac{\text{absorbance of sample read on spectrophotometer}}{\text{Slope of the standard (0.888)}}
\]

2.6. DPPH Assay

The method described by Garcia et al., [21] was modified, 500 microlitres of prepared samples were respectively mixed with 500 microlitres of methanolic solution containing 0.3 mmol·L⁻¹ DPPH radicals. The mixture was shaken vigorously and left to stand for 30 min in the dark, and the absorbance was then measured at 516 nm against blank (methanol). The percentage scavenging effect was calculated according to the equation:

\[
\% \text{ DPPH Scavenged} = \left( \frac{A_{\text{reference}} - A_{\text{sample}}}{A_{\text{reference}}} \right) \times 100
\]

3. Results and Discussion

The consumption of beans has been associated to several health benefits like reduction of cholesterol level [22] and coronary heart diseases [23] [24], favorable effects against cancer [9], decrease of diabetes and obesity [25], high antioxidant capacity [26], antimitogenic [27] [28], and antiproliferative effects [29] [30] because they are good source of protein, essential vitamins and minerals, soluble-fiber starch, phytochemicals, and it is a low fat food [26]. The quality of bean seed is determined by the protein content, amino acid composition, digestibility, presence of anti-nutritional factors and mineral content [25] [31].

This study involves several analyses that comparatively studied the mineral content and scavenging ability of some wild and edible bean in relation to healthy living in humans. Ash content which represents the inorganic component (minerals) of the sample after all moisture and organic material has been removed, as well as the degree of purity of the beans was carried out on unprocessed and malted bean samples (Table 2). The result
Table 2. Ash content of unprocessed bean sample and malted bean sample.

<table>
<thead>
<tr>
<th>Samples</th>
<th>Local name</th>
<th>Ash content for dry bean</th>
<th>Ash content for malted bean sample</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>Otili</td>
<td>4.20 ± 0.13</td>
<td>4.47 ± 0.01</td>
</tr>
<tr>
<td>B</td>
<td>Feregede</td>
<td>3.90 ± 0.02</td>
<td>43.35 ± 0.02</td>
</tr>
<tr>
<td>C</td>
<td>Pakala</td>
<td>5.10 ± 0.15</td>
<td>5.60 ± 0.08</td>
</tr>
<tr>
<td>D</td>
<td>Mucuna</td>
<td>3.42 ± 0.02</td>
<td>3.91 ± 0.05</td>
</tr>
<tr>
<td>E</td>
<td>IT99K-573-2-1</td>
<td>6.91 ± 0.01</td>
<td>6.92 ± 0.01</td>
</tr>
<tr>
<td>F</td>
<td>IT99K-573-1-1</td>
<td>5.68 ± 0.01</td>
<td>5.94 ± 0.01</td>
</tr>
<tr>
<td>G</td>
<td>IT97K-499-35</td>
<td>3.46 ± 0.01</td>
<td>3.18 ± 0.01</td>
</tr>
</tbody>
</table>

showed that edible bean IT99K-573-2-1 had the highest ash content value of 6.90 ± 0.01 in normal sample and 6.92 ± 0.01 in malted samples, subsequently followed by edible bean IT99K-573-1-1 with a value of 5.68 ± 0.01 in ordinary samples and 5.94 ± 0.01 in malted samples. Edible bean IT04K-333-2 had the lowest ash content with the value of 3.14 ± 0.01 in typical bean sample and 3.98 ± 0.01 in malted bean sample, followed by wild type bean mucuna with the value of 3.42 ± 0.02 in common bean sample and 3.91 ± 0.51 in malted samples. The relatively higher ash content of edible bean IT99K-573-2-1 (6.90 ± 0.01) generally indicates higher concentration of minerals than the other varieties. Significant difference (p < 0.05) was observed between the malted and dried ash content for each samples implying that malting increases mineral content of the beans.

In all, eight metals were analysed out of which six were essential though in various amount i.e. micro and macro element (Na⁺, Ca²⁺, K⁺, Mg²⁺, Zn²⁺ and Fe²⁺) while the rest two which were heavy metals (Cd and Pb) were equally analysed. From the result (Table 3), Edible bean—IT07K-243-1-10 had the highest sodium content of 12.40 while wild bean—Otili had the least Na²⁺ content of 10.60 but generally, virtually all the samples have a range of 10.0 - 12.40 ppm. Sodium helps our body retaining the body’s water and pH. It enables cell to draw in nutrients [32]. Winham et al., [24] reported the association of high sodium content with hypertension. Higher sodium intake leads to the retention of more water including water in the blood vessels. This raises the blood pressure, if it raises the blood pressure too high, this can cause health problems such as hypertension.

Edible bean—IT07K-243-1-10 has the highest calcium (Ca²⁺) content of 79.10 with a wide difference from Edible bean—IT04K-333-2 that has higher Ca²⁺ as 29.00 ppm followed by wild bean—Pakala and Mucuna at 27.70 and 27.00 ppm respectively. Edible bean—IT97K-499-35 has the least Ca²⁺ content of 10.80 ppm. Edible bean IT99K-573-1-1 and IT845-2246-4 share equal amount of Ca²⁺ as 19.30. Edible bean—IT99K-573-2-1 has 23.70 ppm of Ca²⁺ while wild bean—Mucuna and Otili has almost equal amount of Ca²⁺ as 11.90 and 11.40 respectively. Calcium deficiency has been epidemiologically linked to several chronic diseases, including osteoporosis, hypertension, and colon cancer [7].

Edible bean—IT07K-243-1-10 has highest amount of K⁺ as 163 ppm followed by samples B and I that share equal amount at 162.00 ppm. At concentration 147.00 wild bean—pokala took the fourth position while wild bean—Mucuna is fifth concentrated at 122.00 ppm, Edible bean—IT99K-573-1-1 has 118.00 ppm while Edible bean—IT99K-573-2-1 is 5 units above a hundred. Three samples have their concentrations less than a hundred as wild bean—Otili has 82.70 ppm, Edible bean—IT985-2246-4 has penultimate least at 79.80 ppm while Edible bean—IT97K-499-35 among others has least quantity of Ca²⁺ as 66.30 ppm. Potassium is nutritionally important in the maintenance of cellular water balance, pH regulation in the body and it is also associated with protein and carbohydrate metabolism [33]. A diet rich in potassium seems to lower blood pressure, getting enough potassium from diet may benefit bones.

The metals Cadmium and Lead could not even be traced in any amount for any of the samples. Lead and cadmium are non-essential metals as they are toxic, even in trace [34]. They are best known for their toxicological properties. Pb and Cd can be accumulated in biological systems, becoming potential contaminants along the alimentary chain. This implies that the Bean samples are completely free from heavy metals that can harm the living system.

All the samples has presence of Mg²⁺ at a close range with Edible bean—IT04K-333-2 having the greatest amount as 6.02 ppm while wild bean—Otili has 4.86 ppm as the least and the penultimate least is Edible bean—IT97K-499-35 at concentration 4.99 ppm. The rest samples generally have some decimals above 5 ppm for presence of Mg²⁺.
Legumes are known as zinc accumulators [17] [35]. Generally, all the samples have the trace amount of Zn$^{2+}$ because Zn$^{2+}$ is a trace element. Edible bean—IT04K-333-2 has the greatest amount as 1.87 ppm while wild bean—otili has the least amount as 0.59 ppm. The penultimate least quantified is found in Edible bean—IT97K-499-35 as 0.74 ppm. The rest of the samples apart from the highest have some decimals above 1.00 ppm Zn has been recognized as a co-factor of the superoxide dismutase enzyme, which is involved in protection against oxidative processes. The main symptoms of Zn deficiency include pregnancy complications, low birth weight, maternal and infant mortality and reduction of growth in infancy and childhood [36].

Fe$^{2+}$ is found in all the bean samples in a trace amount. The sample with highest amount of Fe$^{2+}$ is Edible bean—IT04K-333-2 as 1.87 ppm while wild bean—mucuna is greater at 2.96 ppm followed by Edible bean—IT07K-243-1-10 at 2.87 ppm and wild bean—pakala at 2.81 ppm. The rest samples were few decimals above 2.0 ppm. Iron deficiency is the common nutrition disorder worldwide and affects a large proportion of women and children in developing countries. Indeed, an estimated 3.5 to 5 billion people are Fe deficient in the world [37] [38].

Further study to assess the changes in the radical scavenging ability of the various sample in this study before and after malting (Table 4 & Table 5) showed that all the bean species exhibited antioxidant efficacy assessed by DPPH assay. DPPH content were lower in all bean samples after malting except in edible bean—IT04K-333-2 and IT845-2246-4. Thus, indicating malting decreases DPPH content. It is shown that wild bean species consist of good antioxidant activity as compared to edible bean species.

Wild type bean—mucuna had the highest Vitamin E content in it with a value of 2.6791 ± 0.0002 in the dried bean, and 2.2371 ± 0.000294 in the malted sample while edible bean—IT07K-243-1-10 had the lowest vitamin E content with the value of 0.2578 ± 0.001 and 0.3101 ± 0.000574 in dry and malted bean respectively. Vitamin E is the most powerful natural antioxidant. It is involved in removal of free radicals and prevents their peroxidative effects on unsaturated lipids of membranes and thus helps maintain the integrity of cell membrane and reducing the risk of atherosclerosis [39] [40]. Vitamin E also protects red blood cells from hemolysis by preventing peroxidation. Vit E (α-tocopherol) reacts with the lipid peroxide radicals formed by peroxidation of polyunsaturated fatty acids before they can establish a chain reaction, acting as free radical trapping antioxidant [41]. The tocopheroxy-free radical product, formed in the process, is relatively unreactive and ultimately forms non-radical compounds. Usually the tocopheroxy radical is reduced back to a-tocopherol again by reaction with vitamin C from plasma or reduced glutathione (G-SH).

Vitamin C content was highest in Otili and brown edible bean—IT845-2246-4 with the value of 6.83 ± 0.508 and 4.22 ± 0.046 in dry samples and 4.90 ± 0.228619 and 3.08 ± 0.32. Thus dried pulverised Otili has more radical scavenging ability than other bean in this study. The fact that vitamin C is very sensitive to reversible oxidation, Ascorbic acid to Dehydroascorbic acid, suggests that it may be involved in cellular oxidation reduction reactions, perhaps serving as hydrogen transport agent. Ascorbic acid is required for functional activities of fibroblasts, and osteoblasts, and consequently for formation of MPS of connective tissues, osteoid tissues, dentine and intercellular cement substance of capillaries [42].
Table 4. Table showing DPPH, Vitamin E and Vitamin C content of dry pulverized samples.

<table>
<thead>
<tr>
<th>Samples</th>
<th>DPPH</th>
<th>Vitamin E</th>
<th>Vitamin C</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>56.04 ± 2.15</td>
<td>0.9850 ± 0.001</td>
<td>6.83 ± 0.508</td>
</tr>
<tr>
<td>B</td>
<td>77.05 ± 0.35</td>
<td>0.3148 ± 0.001</td>
<td>3.85 ± 0.338</td>
</tr>
<tr>
<td>C</td>
<td>17.98 ± 1.43</td>
<td>1.4199 ± 0.001</td>
<td>2.09 ± 0.374</td>
</tr>
<tr>
<td>D</td>
<td>23.43 ± 1.61</td>
<td>2.6791 ± 0.0002</td>
<td>8.22 ± 0.184</td>
</tr>
<tr>
<td>E</td>
<td>63.03 ± 0.33</td>
<td>1.5900 ± 0.001</td>
<td>3.74 ± 0.275</td>
</tr>
<tr>
<td>F</td>
<td>69.52 ± 0.08</td>
<td>1.4787 ± 0.003</td>
<td>3.09 ± 0.041</td>
</tr>
<tr>
<td>G</td>
<td>72.15 ± 0.09</td>
<td>0.5486 ± 0.001</td>
<td>3.33 ± 0.012</td>
</tr>
<tr>
<td>H</td>
<td>64.89 ± 0.60</td>
<td>0.2578 ± 0.001</td>
<td>3.84 ± 0.186</td>
</tr>
<tr>
<td>I</td>
<td>64.87 ± 0.05</td>
<td>0.5040 ± 0.001</td>
<td>2.99 ± 0.046</td>
</tr>
<tr>
<td>J</td>
<td>62.59 ± 0.33</td>
<td>0.2942 ± 0.001</td>
<td>4.22 ± 0.045</td>
</tr>
</tbody>
</table>

Table 5. Table showing DPPH, Vitamin E and Vitamin C of malted pulverized sample.

<table>
<thead>
<tr>
<th>Sample</th>
<th>DPPH</th>
<th>Vitamin E</th>
<th>Vitamin C</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>53.18 ± 0.76</td>
<td>0.0302 ± 0.000648</td>
<td>4.90 ± 0.228619</td>
</tr>
<tr>
<td>B</td>
<td>58.04 ± 0.25</td>
<td>0.4452 ± 0.000748</td>
<td>9.84 ± 0.465689</td>
</tr>
<tr>
<td>C</td>
<td>15.85 ± 0.35</td>
<td>0.2035 ± 0.000206</td>
<td>14.99 ± 0.009574</td>
</tr>
<tr>
<td>D</td>
<td>19.28 ± 1.06</td>
<td>2.2371 ± 0.000294</td>
<td>11.49 ± 0.273785</td>
</tr>
<tr>
<td>E</td>
<td>63.54 ± 0.18</td>
<td>1.9271 ± 0.000572</td>
<td>6.88 ± 0.423507</td>
</tr>
<tr>
<td>F</td>
<td>41.66 ± 0.63</td>
<td>0.9333 ± 0.00045</td>
<td>21.63 ± 0.183734</td>
</tr>
<tr>
<td>G</td>
<td>63.14 ± 1.35</td>
<td>0.8236 ± 0.00042</td>
<td>13.57 ± 0.977493</td>
</tr>
<tr>
<td>H</td>
<td>54.34 ± 0.71</td>
<td>0.3101 ± 0.000574</td>
<td>5.12 ± 0.222809</td>
</tr>
<tr>
<td>I</td>
<td>76.00 ± 0.38</td>
<td>0.5788 ± 0.000492</td>
<td>7.65 ± 0.183757</td>
</tr>
<tr>
<td>J</td>
<td>72.50 ± 0.86</td>
<td>0.8236 ± 0.00042</td>
<td>3.08 ± 0.323213</td>
</tr>
</tbody>
</table>

4. Conclusion

Arising from the comparative analysis carried out in this study, edible bean—IT07K-243-1-10 among others appears to have the greatest quantity of mineral elements followed by edible bean—IT04K-333-2 and wild bean—feregede. Edible bean—IT97K-499-35 has the least mineral content compared to other bean samples followed closely is the wild bean—Otili with least mineral content. Wild and near extinct bean feregede is recommended for human consumption because it contains appreciable amount of minerals and antioxidants compare to other cultivated edible beans.

Acknowledgements

The present project was partly funded by Tet Fund Research Grant 2015.

References


