Cancer chemoprevention through the induction of apoptosis by natural compounds

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ABSTRACT

As cell and tissue homeostasis are mediated by the balance between proliferation and apoptosis, controlling this balance is important for cancer chemoprevention. Cancer chemoprevention can be achieved by the use of natural, synthetic or biologic compounds that reverse, suppress or prevent the development of epithelial malignancies. Natural compounds including flavonoids are able to reduce oxidative stress, which is the most likely mechanism mediating the protective effects against cancer development. In addition, in vitro and in vivo studies have suggested that flavonoids, such as (-)-epigallocatechin-3-gallate (EGCG), quercetin, and curcumin, act by induction of apoptosis. Several natural compounds inhibit cell proliferation and angiogenesis. Certain natural products have been shown to inhibit the activation of nuclear factor kappa B (NF-κB) and Akt signaling pathways, both of which are known to maintain a homeostatic balance between cell survival and apoptosis. Understanding the mechanism of these natural products will contribute to the development of more specific preventive strategies against cancer development. Here we focus on the ability of natural cancer chemopreventive agents to induce apoptosis, and attempt to provide evidence for the preventive and therapeutic effects of natural compounds, EGCG, quercetin, and curcumin, in a succinct manner highlighting both NF-κB and Akt signaling pathways in vivo.

Keywords: Cancer Chemoprevention; Apoptosis; Natural Compounds

1. INTRODUCTION

Epidemiological studies have shown that diet consisting of a high rich in fruit and vegetable reduces the risk of several types of cancer [1]. Intake of fruit and vegetables has been successfully used in the prevention of chronic diseases associated with oxidative stress conditions, including cancer [2,3]. The cancer preventing properties of fruit and vegetables have been ascribed, at least in part, to their high content of polyphenols [4]. The majority of polyphenols present in food are flavonoids and phenolic acids that are an integral part of the human diet. Laboratory rodent studies have shown that polyphenols have cancer-preventing properties and considered to be potential chemopreventive agents [5-7]. They can influence important cellular and molecular mechanisms associated with multiple carcinogenic steps, such as expression of key proteins in signal transduction pathways (e.g., mitogen activated protein kinases (MAPKs) or activator protein (AP)-1), the transcription factor nuclear factor-kappa B (NF-κB) and its downstream gene products, modulation of cell-cycle regulation and induction of apoptosis [7], which affect cell differentiation, proliferation and apoptosis, immune responses and metabolism of carcinogens [4].

Apoptosis is conceivably the most potent defense against cancer development since it is the mechanism used by metazoans to eliminate deleterious cells. Many chemopreventive agents have been shown to induce apoptosis in transformed cells both in vitro and in vivo. Induction of apoptosis appears to be associated with their effectiveness in modulating carcinogenic processes [8]. Since apoptosis provides a physiologic mechanism for eliminating initiated or abnormal cells, dietary factors affecting apoptosis can influence carcinogenesis. In fact, activation of apoptosis in pre-cancerous cells is one of the most important mechanisms of cancer chemoprevention by dietary factors [9].

In this review, we focus on the apoptosis-inducing properties of several natural compounds present in the human diet and describe their beneficial effects against cancer development. Understanding the mechanism of these natural products will contribute to the development...
of more specific preventive strategies against cancer.

2. CHEMOPREVENTIVE COMPOUNDS IN FOOD

An effective chemopreventive agent should preferably intervene early in the process of carcinogenesis to eliminate pre-malignant cells before they acquire malignant character. Many chemopreventive agents are able to block or delay the promotion and/or progression of pre-malignant or malignant cells by modulating cell proliferation and/or differentiation [8,10] and, therefore, should be chronically administered to individuals with a higher risk of cancer development. However, using this approach, even minor adverse effects would be unacceptable: obstacles to the use of chemoprevention for many cancers include long-term toxicity and the development of chemoresistance [8]. These issues can limit the feasibility and success of conventional forms of chemoprevention for many cancers. Alternative approaches involve the use of agents that eliminate cells expeditiously. Targeted delivery of apoptosis-inducing agents to tumor cells would also prevent the need for chronic exposure, limiting the risk of long term toxicity and/or the development of chemoresistance [8].

An ideal chemopreventive agent should be selective for damaged or transformed cells, display a significant bioavailability in the target lesion and have more than one mechanism of action. Moreover, it should be highly effective, easy to administer, and inexpensive. Dietary compounds are particularly attractive because of long-standing exposure to them by humans, their relative lack of toxicity, and encouraging indications from epidemiological studies [11]. Indeed, numerous dietary compounds and micronutrients are emerging that have considerable potential for hindering in vivo deleterious oxidative processes and inducing apoptosis in cancer cells. One potentially important drawback of dietary compounds is their possible low bioavailability after ingestion [12,13].

Food contains several promising chemopreventive compounds [14-16]. Plant polyphenols, ubiquitous in a diet rich in fruit and vegetables, are thought to be responsible for the cancer protective effects ascribed to this type of diet [4,5,13]. In fact, numerous phenolic compounds have been shown to display anti-proliferative and cytotoxic effects towards several tumor cells, presenting toxic effects that specifically target cancer cells rather than normal cells [17-20]. Dietary polyphenols are predominantly consumed through fruit and beverages (juice, wine, tea, coffee, chocolate and beer), with the exception of vegetables, cereals and olive derivatives that are mainly associated with the Mediterranean diet [21-23]. Their average daily intake has been reported to be approximately 1 g [13,24], which is much greater than the intake of all other classes of dietary anti-oxidants. For example, this value is approximately 10-times higher than vitamin C intake and 100-times greater than the intake of both vitamin E and carotenoids [24].

In addition to the anti-oxidant properties, polyphenols exert interesting biological abilities in animal experiments and in vitro systems. The compounds are able to trap and scavenge free radicals, decrease leukocyte immobilization, induce apoptosis, inhibit cell proliferation and angiogenesis, and exhibit phytoestrogenic activity [25-27]. Dietary polyphenols interfere with signal transduction pathways related to the carcinogenesis process, thereby acting as chemopreventive agents. They include the suppression of NF-κB and activating protein (AP-1) activation, inhibition of the mitogen-activated proteins (MAPKs)-, protein kinases- and growth-factor receptor-mediated pathways, cell cycle arrest, induction of apoptosis, anti-oxidant and anti-inflammatory effects, and suppression of angiogenesis [7].

The chemopreventive properties of a variety of compounds can be directly related to their pro-apoptotic properties, most probably exerted via the intrinsic (mitochondrial) apoptotic pathway [8,10] (Table 1).

Pro-apoptotic diet-derived compounds can conceivably protect against cancer by enhancing elimination of initiated, precancerous cells. Polyphenols have been shown to induce pro-apoptotic responses in malignant or pre-malignant cells. Many studies have described the pro-apoptotic properties of dietary polyphenols in a variety of human cancer cell lines derived from colon, prostate, lung, breast cancers, and leukemia [28]. A major catechin, EGCG, among green tea catechins is the most potent polyphenolic compound with respect to inducing apoptosis and inhibiting proliferation in cancer cells: it induces apoptosis in and suppresses growth of human cancer cell lines, such as breast adenocarcinoma (MCF-7 and MDA-MB-231 cells) [29], oral squamous carcinoma [30], leukemia [31], breast [32], lung [33], prostate [34] and colon [35] cancers.

Several studies have indicated that phenolic concentrations leading to apoptosis are within the micromolar range. For example, quercetin reportedly induces apoptosis at concentrations ranging from 29 M to 150 M [36,37]. In general, the effective concentrations required for induction of apoptosis are higher than those leading to growth inhibition. However, some studies have reported the induction of apoptosis without inhibition of cell proliferation (e.g. EGCG effect on H661 lung cancer cells) [38]. Further increases in polyphenol concentration may cause necrosis of all cell lines tested [39].

Recent studies have proposed that different dietary phenolic compounds may act synergistically, together with established anti-cancer agents and act as enhancers of anti-carcin drugs [40,41] (Table 2). For example, the synergistic effects of EGCG and curcumin in pre-ma-
lignant and malignant human oral epithelial cells [42], resveratrol and quercetin in human pancreatic cancer cells [36], quercetin and cisplatin in human laryngeal Hep2 cells [43] and HeLa cells [44], and EGCG and sulindac or tamoxifen against human PC-9 lung cancer cells [45] have been described. This recognized synergy among dietary phenolics and conventional synthetic drugs provides an interesting approach to combination therapy as well as for the pre-treatment of neoplastic cells with polyphenolic agents. This strategy has been able, in some cases, to even overcome chemoresistance [46].

The anti-carcinogenic and cytotoxic activities of polyphenols are largely determined by structural parameters, as much as their anti-oxidant potencies [20,47-50]. Despite their close resemblance, their bioactivity varies considerably upon minor structural modifications, since such modifications often induce significant conformational changes [7,51,52]. This implies an important drawback in the understanding of the effects of polyphenols in human health, when considering the huge number of different compounds (>8000) [12].

### 3. APOPTOSIS

Although the term apoptosis was first used in 1972 [53] to describe a morphologically distinct form of cell death, several components of this concept have only recently been described in detail. The process of programmed cell death is characterized by distinct morphological characteristics and energy-dependent biochemical mechanisms. Apoptotic cells show morphological changes such as cell shrinkage, pyknosis and extensive plasma membrane blebbing leading to the formation of apoptotic bodies, which are subsequently phagocytosed by macrophages.

<table>
<thead>
<tr>
<th>Compounds</th>
<th>Plant</th>
<th>Target organs</th>
<th>Carcinogens/cell line</th>
<th>Animal</th>
<th>Effects/molecular targets</th>
<th>Ref No.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Intestine</td>
<td>None</td>
<td>ApcMin+/– mice</td>
<td>c57bl/kj-db/db mice</td>
<td>Attenuates aberrant nuclear β-catenin and activated Akt and ERK signaling</td>
<td>115</td>
<td></td>
</tr>
<tr>
<td>Intestine</td>
<td>Azoxymethane</td>
<td></td>
<td>c57bl/kj-db/db mice</td>
<td>Overcomes the activation of the IGF/IGF-IR axis, improving hyperlipidemia, hyperinsulinemia, and hyperleptinemia</td>
<td>116</td>
<td></td>
</tr>
<tr>
<td>(-)-Epigallocatechin Green gallocate (EGCG) tea</td>
<td>Liver</td>
<td>Diethylnitrosamine</td>
<td>c57bl/kj-db/db mice</td>
<td>Inhibits IGF/IGF-IR axis, improving hyperinsulinemia, and attenuating chronic inflammation</td>
<td>117</td>
<td></td>
</tr>
<tr>
<td>Tumor-associated endothelial cells and endothelial progenitor cells</td>
<td>A375SM (melanoma), xenograft</td>
<td></td>
<td></td>
<td>Selective anti-angiogenic effects, inhibits the phosphorylation of Akt in tumor-associated endothelial cells, MMP-9 mRNA expression level and vascular endothelial growth factor in endothelial progenitor cells</td>
<td>118</td>
<td></td>
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<tr>
<td>Urinary bladder</td>
<td>UM-UC-3, xenograft</td>
<td></td>
<td></td>
<td>Down-regulates N-cadherin and inactivation of Akt signaling</td>
<td>119</td>
<td></td>
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<tr>
<td>Head and neck</td>
<td>CAL27 (squamous cell carcinoma), xenograft</td>
<td></td>
<td></td>
<td>Suppresses the activation of NF-κB without affecting the expression of pAkt</td>
<td>124</td>
<td></td>
</tr>
<tr>
<td>Breast</td>
<td>MDA-MB23 (adenocarcinoma), xenograft</td>
<td></td>
<td></td>
<td>Inhibits survivin, NF-κB and its downstream effectors cyclin D1 and Bcl-2, and strongly up-regulated p21WAF1</td>
<td>125</td>
<td></td>
</tr>
<tr>
<td>Curcumin</td>
<td>Curcuma longa</td>
<td>Head and neck</td>
<td>SCC40 (squamous cell carcinoma), xenograft</td>
<td></td>
<td>Blocks nicotine-induced activation of the AKT/MTOR pathway in HNSCC, which retards cell migration, reduced MMP-9 expression</td>
<td>128</td>
</tr>
<tr>
<td>Curcumin</td>
<td>Curcuma longa</td>
<td>Head and neck</td>
<td>4-Nitroquinoline 1-oxide, xenograft</td>
<td>CBA/CaJ mice</td>
<td>Decreases COX-2, IL-8, and VEGF mRNA and protein expression, decreased AKT and extracellular signal-regulated kinase activation</td>
<td>133</td>
</tr>
<tr>
<td>Colon</td>
<td>HCT-116 (adenocarcinoma), xenograft</td>
<td></td>
<td></td>
<td>B16F10 (mouse melanoma), xenograft, c57bl mice</td>
<td>Suppresses Cyclin D1, p-NF-κB, BclXL, p-Akt, and VEGF</td>
<td>134</td>
</tr>
<tr>
<td>Brain</td>
<td>ACC-2 and ACC-M (adenoid cystic carcinoma), xenograft</td>
<td></td>
<td></td>
<td>Down-regulates the PI3K/Akt/IKK-α/NF-κB signaling pathway</td>
<td>147</td>
<td></td>
</tr>
</tbody>
</table>
Table 2. Synergistic induction of apoptosis by combining natural compounds and anticancer drugs/radiation through NF-κB or PI3K/Akt pathway in vivo.

<table>
<thead>
<tr>
<th>Compounds</th>
<th>Combination agent</th>
<th>Target organ</th>
<th>Carcinogen/cell line</th>
<th>Animal</th>
<th>Effects/molecular targets</th>
<th>Ref. No.</th>
</tr>
</thead>
<tbody>
<tr>
<td>EGCG</td>
<td>Tamoxifen</td>
<td>Breast</td>
<td>Xenograft, MDA-MB-231 (estrogen receptor-negative breast cancer)</td>
<td>Nude mice</td>
<td>Decreases the tumor protein expression of mTOR and decreases of the expression of EGFR, NF-κB, b-Raf, p-MEK, S6K, 4EBP1, Akt, vascular EGFR-1 (VEGFR-1) and VEGF</td>
<td>148</td>
</tr>
<tr>
<td>Sulforaphane</td>
<td>Pancreas</td>
<td>Xenograft, MIA-PaCa2 (pancreatic cancer)</td>
<td>Nude mice</td>
<td>Affects the self-renewal potential, ALDH1 activity, apoptosis induction, inhibition of angiogenesis, NF-κB and epithelial-mesenchymal transition processes</td>
<td>153</td>
<td></td>
</tr>
<tr>
<td>Quercetin trans-Pterostilbene (trans-3, 5-dimethoxy-4'-hydroxystilbene, t-PTER), FOLFOX6, radiation</td>
<td>Colo-rectum</td>
<td>Xenograft, HT-29 (colorectal cancer)</td>
<td>Nude mice</td>
<td>Over-expresses superoxide dismutase 2 and down-regulates of bcl-2 expression by inhibiting NF-κB activation</td>
<td>155</td>
<td></td>
</tr>
<tr>
<td>Radiation</td>
<td>Colo-rectum</td>
<td>Xenograft, HCT-116 (colorectal cancer)</td>
<td>Nude mice</td>
<td>Potentiates the anti-tumor effects of adiation therapy by suppressing NF-κB and NF-κB-regulated gene products, leading to inhibition of proliferation and angiogenesis</td>
<td>123</td>
<td></td>
</tr>
<tr>
<td>EGCG</td>
<td></td>
<td>Breast</td>
<td>Xenograft, MDA-MB-231 (breast cancer)</td>
<td>Nude mice</td>
<td>Decreases the level of VEGFR-1 protein expression, decreases the tumor protein levels of EGFR and Akt</td>
<td>150</td>
</tr>
<tr>
<td>Gemcitabine</td>
<td>Pancreas</td>
<td>Xenograft, MIA PaCa2 (pancreatic cancer)</td>
<td>Nude mice</td>
<td>Potentiates the anti-tumor effects of gemcitabine by suppressing proliferation, angiogenesis, NF-κB, and NF-κB-regulated gene products</td>
<td>151</td>
<td></td>
</tr>
<tr>
<td>Curcumin</td>
<td>resveratrol</td>
<td>Prostate</td>
<td>None</td>
<td>Prostate-specific PTEN knockout mice</td>
<td>Negatively regulates of the activated p-Akt, cyclin D1, AR and mTOR</td>
<td>154</td>
</tr>
<tr>
<td>Paclitaxel</td>
<td>Uterine cervix</td>
<td>Uterine cervical cells (uterine cervical cancer), 3-methylcholanthrene</td>
<td>NOD-SCID mice</td>
<td>Augments the anti-tumor action of paclitaxel by down-regulating the activation and down-stream signaling of anti-apoptotic factors and survival signals such as NF-κB, Akt and mitogen-activated protein kinases</td>
<td>156</td>
<td></td>
</tr>
<tr>
<td>Cisplatin</td>
<td>Head and neck</td>
<td>Xenograft, CAL27 (SCC)</td>
<td>Nude mice</td>
<td>Inhibits cytoplasmic and nuclear IKKβ, resulting in inhibition of NF-κB activity</td>
<td>157</td>
<td></td>
</tr>
<tr>
<td>Dasatinib</td>
<td>Colo-rectum</td>
<td>None</td>
<td>ApcMin/+ mice</td>
<td>Suppresses EGFRs, IGF-R and c-Src signaling pathway, decreases the activation of down-stream signaling pathways, Akt and Erk(s), associated with decreased NF-κB activity</td>
<td>158</td>
<td></td>
</tr>
</tbody>
</table>

Parenchymal or neoplastic cells. Apoptosis rarely causes an inflammatory response, since 1) apoptotic cells do not release their constituents into the surrounding interstitial tissue, 2) apoptotic cells are quickly phagocytosed by cells in the surrounding tissue, preventing secondary necrosis, and 3) the engulfing cells do not produce inflammatory cytokines [54].

Apoptosis can be initiated by receiving extracellular or intracellular signals including growth factor withdrawal, UV- or gamma-irradiation, chemotherapeutic agents, heat shock, nutrient deprivation, and by a family of transmembrane proteins called death receptors. These signals are transduced to adapter proteins and transmitted to specific cysteine proteases called “initiator caspases”. At this point the cell is committed to undergo apoptosis, followed by “execution of cells” (mediated by sequential activation of the so-called “executioner caspases”), systematic disintegration of cell structure and phagocytosis of the cell corpses [55].

Caspases (cysteine-dependent aspartate-specific prote-
Apoptosis is typically activated during the early stages of apoptosis. This family of proteins is synthesized as inactivezymogens but, once activated, can begin a proteolyticcascade, which results in the cleavage of key cellularcomponents required for normal cellular function,including structural proteins in the cytoskeleton and nu-clear proteins such as DNA repair enzymes. Caspasescan also activate other degradative enzymes such asDNAases, which begin to cleave the DNA in the nucleus.

To date, 14 different members of the caspase-family (Table 3) have been described in mammals [55-57]. The tenmajor pro-apoptotic caspases can be classified as initiators(caspase-2, -8, -9, -10), effectors or executioners(caspase-3, -6, -7) and inflammatory caspases (caspase-1,-4, -5) [55,58]. Other caspases that have been identifiedto date (caspase-11, -12, -13 and -14) are involved in-specific apoptotic processes or expressed solely in spe-cific types of tissue [55].

It is currently accepted that apoptosis can occur via two main pathways: the extrinsic or death receptor (Figure 1) and intrinsic or mitochondrial (Figure 2) pathways [59]. The extrinsic pathway initiated extracellularly via activation of cell surface receptors by specific mole-cules known as pro-apoptotic ligands including CD95L/ FasL (receptor CD95/FasR), and Apo2L/TRAIL (receptorsDR4, DR5) [53]. The Apo2 ligand has sparked growinginterest within the oncology field due to its reported abil-it to selectively trigger cancer cell death [60,61]. Onceactivated, the death domains of these receptors, bind totheadaptor protein Fas-associated death domain (FADD),resulting in the assembly of the death-inducing signalingcomplex (DISC), and recruitment and assembly of the ini-tiator caspase-8 and -10 [62]. Once activated, caspase-8directly activates caspase-3 to initiate degradation of thecell. Active caspase-8 can also cleave Bid (pro-apoptotic)to tBid, which binds to the mitochondrial membrane tofacilitate the release of cytochrome c and initiate the intrin-sic pathways. This allows “cross-talk” between the two mainpathways and amplifies the apoptotic signaling from deathreceptors.

The extrinsic and intrinsic pathways both end at thepoint of the execution phase. Execution caspases activateacytoplasmic endonuclease, which degrades nuclear ma-terial, and proteases that degrade nuclear and cytoskeletalproteins. Such executioners including caspase-3 (considered to be the most important executioner caspase) canbe activated by any of the initiator caspases (caspase-8,-9 or -10).

In addition to p53, the extrinsic and intrinsic pathwaysare also regulated by NF-κB, the ubiquitin proteosome system and the phosphatidylinositol-3-kinase (PI3K) path-way [63]. NF-κB is one of the most studied transcriptionfactors in mammalian cells. Its function has been impli-cated in inflammation, cell proliferation, differentia-

Table 3. Caspases involved in apoptosis or programmed cell death.

<table>
<thead>
<tr>
<th>Type</th>
<th>Name</th>
<th>Synonyms</th>
</tr>
</thead>
<tbody>
<tr>
<td>Initiator or apical</td>
<td>Caspase-2</td>
<td>ICH1, Nedd2</td>
</tr>
<tr>
<td></td>
<td>Caspase-8</td>
<td>FLICE, MACH1, MCH5, FADD-like Ice</td>
</tr>
<tr>
<td></td>
<td>Caspase-9</td>
<td>MCH6, ICELAP6</td>
</tr>
<tr>
<td></td>
<td>Caspase-10</td>
<td>FLICE2, MCH4</td>
</tr>
<tr>
<td>Effectors or executioner</td>
<td>Caspase-3</td>
<td>CPP32, YAMA</td>
</tr>
<tr>
<td></td>
<td>Caspase-6</td>
<td>MCH2</td>
</tr>
<tr>
<td></td>
<td>Caspase-7</td>
<td>MCH3, CMH, ICELAP3</td>
</tr>
<tr>
<td>Inflammatory</td>
<td>Caspase-11</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>Caspase-12</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>Caspase-13</td>
<td>ERICE</td>
</tr>
<tr>
<td></td>
<td>Caspase-14</td>
<td>MICE</td>
</tr>
</tbody>
</table>

a: Caspase = cysteinyl aspartic acid-protease.

Figure 1. Extrinsic pathway.
anti-apoptotic and pro-apoptotic functions. Physiologically, NF-κB induces resistance to apoptosis through activation of inhibitor of apoptosis (IAP) and X-linked IAP. In addition, NF-κB activation has been shown to inhibit p53-dependent apoptosis following expression of the oncogene AP12/MALT1 [66]. This NF-κB directed survival response is associated with increased expression of anti-apoptotic proteins. Thus, it is not surprising that NF-κB expression is dysregulated in various disease states including chronic inflammation and cancer. In contrast, some stimuli that lead to activation of NF-κB may induce apoptosis, probably via the activation of pro-apoptotic proteins such as c-myc, p53 and caspase-1 [67].

The PI3K signaling pathway is also crucial for many aspects of cell growth, survival and tissue neo-vascularization and is frequently up-regulated in many cancers [68]. The PI3Ks are a family of related enzymes that are capable of phosphorylating the 3 position hydroxyl group of the inositol ring of PI(4,5)P2 to generate PI(3,4,5)P3 [69]. Upon activation of the PI3K pathway by many growth factors such as epidermal growth factor (EGF), PI(3,4,5) P3 is produced on the inner side of the plasma membrane and binds to Akt. Akt inactivates pro-apoptotic factors such as Bad, which controls the release of cytochrome c [70,71], procaspase-9 and Forkhead transcription factors (such as FOXO). Akt also activates anti-apoptotic genes including the cyclic-AMP response element-binding protein and IKB kinase leading to NF-κB nuclear localization and the subsequent transcription of pro-survival genes such as Bcl-xL, caspase inhibitors and c-Myb [72,73]. Over-expression of Akt has anti-apoptotic effects in various cell types resulting in resistance to cell death [74].

4. CANCER CHEMOPREVENTION AND APOPTOSIS

Cancer is a pathologic condition where the normal mechanisms of cell cycle regulation are dysfunctional either due to excessive cell proliferation, insufficient apoptosis or both [75-77]. Suppression/inhibition of apoptosis during
cancerogenesis is known to play a role in the development and progression of cancers [77-79]. At present, it is accepted that cell populations are tightly regulated by their rates of proliferation, differentiation and death. When the homeostatic balance is disrupted in such a way that clonal outgrowth of mutated cell populations occurs, the development of a tumor will followed [77-79].

In simple terms, one can define carcinogenesis as a multistage process where a normal cell becomes transformed into one with a malignant phenotype. Cells become initiated by the acquisition of an activating mutation in an oncogene or an inactivating mutation in a tumor suppressor gene (initiation). Several additional factors confer these cells a growth advantage (promotion), which allow the cell to survive while accumulating abnormal characteristics and ultimately progressing to a metastatic tumor (progression) [80]. Carcinogenesis is a complex process driven by tight interactions between oncogene activation, tumor suppressor inactivation and cell death machinery. Early in transformation, activated oncogenes that drive the cell to uncontrolled proliferation simultaneously trigger apoptosis, probably as a safety mechanism to remove cells carrying oncogenic mutations [81]. Later in tumorigenesis, the supply of nutrients and oxygen becomes limited, with the tumor cells undergoing hypoxia-induced apoptosis [82]. In order to survive, tumor cells acquire apoptotic-inhibiting mutations (reduced apoptosis) [82]. Failures in normal apoptotic pathways contribute to carcinogenesis by creating a permissive environment for genetic instability and accumulation of mutations, promoting resistance to immune-based destruction, overriding cell-cycle checkpoints (that would normally induce apoptosis), facilitating growth factor/hormone-independent cell survival, supporting anchorage-independent survival during metastasis, reducing dependence of oxygen and nutrients, and conferring resistance to cytotoxic anticancer drugs and radiation [77]. Thus, inhibition of apoptosis can lead to tumor development.

In animal models, most chemical initiators are unable to initiate tumor growth unless a tumor promoter is subsequently applied. Many tumor promoters inhibit apoptosis in vitro [83]. Activation of apoptosis is thus being considered to be one of the most promising therapeutic approaches in cancer therapy [63,77,84]. Tumor cells can acquire resistance to apoptosis, for instance, by overexpressing anti-apoptotic proteins such as Bcl-2 or down-regulating/mutating pro-apoptotic proteins such as Bax, the expression of both being regulated by the p53 tumor suppressor gene [85,86]. p53 is a transcription factor essential for the prevention of cancer formation, which can be damaged by radiation, several chemicals and viruses such as human papillomavirus (HPV). The p53 pathway is ubiquitously lost in human cancer either by p53 gene mutation or by loss of cell signaling upstream and downstream of p53 in cancers that express the WT p53 gene [87]. Therefore, despite the enthusiasm towards apoptosis based-drugs, possible difficulties are also being anticipated such as selection of apoptosis-resistant tumor cells and systemic toxicity [84].

Several epidemiological studies, later evaluated by meta-analysis, have identified associations between certain dietary factors and cancer that either increase or decrease cancer risk [15,88,89]. It is currently accepted that diet can affect the overall process of carcinogenesis by different mechanisms: its constituents may contain cancer-causing substances as well as many cancer preventive agents. These dietary agents can retard or prevent the process of carcinogenesis by multiple mechanisms, namely 1) enhanced detoxification of the carcinogenic intermediates through induction of phase 2 drug metabolizing enzymes, 2) reduced carcinogenic activation due to suppression of cytochrome P450-dependent monoxygenases, 3) perturbations in cell cycle progression, 4) selective promotion of apoptosis in cancerous or precancerous cells, and 5) inhibition of angiogenesis and metastasis formation [16]. Since apoptosis provides a physiologic mechanism for eliminating abnormal cells, dietary factors affecting apoptosis can have an important effect on carcinogenesis. Conceivably, dietary factors that activate apoptosis in pre-cancerous cells offer a cancer preventive mechanism. In fact, most initiated cells are destroyed by apoptosis before they become malignant and develop into a tumor [79]. Increased understanding in the field of cancer has led to the conviction that most human malignancies should be fought on multiple fronts: in addition to cancer therapy, cancer prevention has become an important means of controlling cancer [8]. Common prevention strategies include avoiding exposure to known cancer-causing agents, enhancement of host-defense mechanisms against cancer, life style modifications and chemoprevention [8].

The term chemoprevention refers to the use of agents to slow the progression of, reverse or inhibit carcinogenesis, and was first introduced by Sporn and co-workers in the mid-1970’s [90]. Animal studies, clinical trials and in vitro studies have examined the anticancer activity of numerous putative chemopreventive agents. These studies strongly suggest that the anti-cancer activities of many of these compounds involve the induction of apoptosis, and support the notion that apoptosis is a novel target for cancer chemoprevention [8,10]. Moreover, the pro-apoptotic properties of a variety of chemopreventive agents, like those of many conventional and experimental cancer chemotherapeutic agents, appear to be related to mitochondrial alterations in tumor cells [10,91]. In fact, several classes of chemopreventive agents contain members that trigger mitochondrial disruption and/or mitochondrial-mediated apoptosis (intrinsic pathway) in tumor cells in
vitro, although other agents may induce apoptosis via the death receptor pathway [9].

Chemotherapy aims to kill cancer cells, in the hope of preventing further cancer progression. Chemoprevention, on the other hand, involves administering non-toxic agents to individuals who may be at an increased risk for cancer. Moreover, surgical and traditional therapeutic approaches (chemotherapy and radiation) are, at present, unable to control most cancer types. Thus, the development of new chemopreventive strategies is required [10,11,92,93]. Chemopreventive compounds can be classified into two major groups: 1) blocking agents, which prevent carcinogens from reaching or reacting with critical target sites, and 2) suppressing agents, which stop the evolution of the pre-neoplastic process. Given that the initiation and progression phases are relatively transient and irreversible events, it seems logical that chemopreventive agents should intervene at the prodomal promotion phase. Three decades of research suggest that chemoprevention is a promising strategy to reduce the incidence of cancer, both in well-defined high-risk groups and in the general population [10-12,92,93].

Of great importance, aberrant NF-κB regulation and Akt activation has been observed in many cancers. To prevent the development and progression of cancers, the strategy should target the cell signaling pathways that are deregulated in malignant tumors. Aberrant regulation of NF-κB and the signaling pathways that control its activity are involved in cancer development and progression, as well as in drug resistance, especially during chemotherapy and radiotherapy [94]. Blocking NF-κB can cause tumor cells to cease proliferation or become more sensitive to the action of antitumor agents [95]. Changes in Akt activator expression observed in human precancerous tissues that might be targeted for chemoprevention [96]. Inhibition of Akt signaling has been associated with the biological actions of numerous chemopreventive compounds. Thus far, several chemopreventive agents including, green tea polyphenols, curcumin, and quercetin have shown their various activities in the inhibition of carcinogenesis through the regulation of major cell signaling pathways such as Akt and NF-κB. Therefore, those are the subject of intense study. Agents capable of suppressing Akt and/or NF-κB activation have therapeutic promise and potential to inhibit carcinogenesis. We currently review dietary cancer chemopreventive compounds including EGCG, curcumin, and quercetin, capable of functioning in this capacity.

Representative dietary cancer chemopreventive compounds include EGCG, curcumin, and quercetin.

4.1. EGCG

Cancer prevention by green tea and its constituents has been studied in different animal models of carcinogenesis [97]. The major catechins (a group of polyphenols) in green tea are EGCG, (-)-epigallocatechin (EGC), (-)-epicatechin-3-gallate (ECG) and (-)-epicatechin. EGCG, the most abundant and most studied catechin, exhibits significant growth inhibitory effects in cancer cells. More importantly, after treatment of a primary cell line such as normal epithelial cells with EGCG, there is no observable toxicity at doses that are used for cancer inhibition studies [98,99]. A collection of reviews have commented on the possible mechanistic effects of EGCG in multiple cell lines and have noted similarities in regard to growth inhibition and cell cycle arrest [100-102].

EGCG reportedly affects the transcription factors p53 and NF-κB leading to a change in the ratio of Bax/Bcl-2 in a manner that favors apoptosis [103]. The induction of apoptosis by other green tea catechins has been evaluated in a dose dependent manner (i.e. ECG > EGCG > EGC > EC) [104].

EGCG treatment may lead to a significant dose- and time-dependent inhibition of activation and translocation of NF-κB to the nucleus by suppressing the degradation of IκBα in the cytoplasm [105,106]. EGCG may also inhibit the ATP- and IL-1β-induced activation of NF-κB [107].

EGCG has been found to inhibit PI3K/Akt activation, resulting in the modulation of Bcl-2 family proteins and leading to the enhanced apoptosis of bladder cancer cells [108]. EGCG has also been shown to inhibit vascular endothelial growth factor (VEGF)-induced angiogenesis in vitro through suppression of VE-cadherin phosphorylation and inactivation of Akt, suggesting that EGCG has an inhibitory effect on the Akt signaling pathway [109, 110]. Further studies have also demonstrated that constitutive activation of Akt, EGFR and Stat3 was inhibited in both YCU-H891 head and neck squamous cell carcinoma and MDA-MB-231 breast carcinoma cell lines treated with EGCG [111].

A dose-dependent increase in p53 was observed after EGCG treatment of LNCaP cells, which carry WT p53, but not in DU145 cells carrying mutant p53 [112]. EGCG was also shown to stabilize p53 and cause up-regulation of its transcriptional activity, resulting in the activation of its downstream targets such as p21WAF1 and Bax, and the induction of apoptosis [113]. In human liver cancer cells, a significant increase in the expression of p53 and p21WAF1 protein that lead to cell cycle arrest was reported after EGCG treatment [114]. Table 1 summarizes the action/mechanisms of three selected compounds that affect NF-κB and Akt-phosphatidylinositol 3-kinase (PI3K) activity in vivo system.

Administration of EGCG to Apcmin/+ mice, an animal model of human intestinal carcinogenesis, via their drinking fluid was found to significantly decrease small intestinal tumor formation [115]. Shimizu et al. have recently re-
ported that EGCG prevents obesity-related colonic and liver tumorigenesis by inhibiting the phosphorylation of the insulin like growth factor-1 receptor (IGF-1R), extracellular signal-regulated kinase (ERK), Akt, glycogen synthase kinase-3beta (GSK-3β), signal transducers and activators of transcription 3 (Stat3), and c-Jun NH2-terminal kinase (JNK) proteins, and improving hyperinsulinemia in carcinogen-induced mouse model [116,117]. EGCG suppressed the growth of melanoma cells in nude mice with impaired angiogenesis by inhibiting PI3K/Akt signaling specifically in tumor-associated endothelial cells and peripheral blood-derived endothelial cells [118]. Continuous feeding with EGCG to mice prior to and during establishment of bladder carcinoma xenografts in vivo revealed >50% reduction in mean tumor volume without detectable toxicity [119].

4.2. Curcumin

It is widely accepted that curcumin, a yellow pigment found in the rhizome of the spice turmeric, has potent cancer chemopreventive activity in various animal carcinogenesis models [120]. Curcumin is known to be a strong inhibitor of NF-κB. Curcumin has been shown to inhibit IKK, suppress both constitutive and inducible NF-κB activation, and potentiate tumor necrosis factor (TNF)-induced apoptosis [121]. Recent studies have shown that curcumin suppresses the constitutive activation of NF-κB [122] and sensitizes human colorectal cancer xenografts in nude mice to gamma-radiation by targeting NF-κB regulated gene products [123]. Treatment with a liposomal formulation of curcumin resulted in a dose-dependent growth suppression of cancer cells and decreased activation of NF-κB [124]. The findings that expression of NF-κB target genes, including cyclin D1, cyclooxygenase (COX)-2, matrix metalloproteinase-9, Bcl-2, Bcl-xL, Mcl-1L and Mcl-1S, was reduced by the treatment with a liposomal formulation of curcumin indicate that curcumin acts the NF-κB pathway which is involved in carcinogenesis. Al-Hujaily et al. have demonstrated that a novel curcumin analogue, PAC, significantly reduced tumour size, and triggered apoptosis in breast cancer tumor xenografts by inhibiting expression of survivin, NF-κB and its downstream effectors, and strongly up-regulated p21 (WAF1) [125]. Moreover, clinical trials have shown that curcumin down-regulates the expression of NF-κB and COX-2 in peripheral blood mononuclear cells from patients with pancreatic cancer [126]. These results clearly indicate that curcumin inhibits tumor growth by affecting the NF-κB signaling in vitro and in vivo.

Curcumin also exhibits an inhibitory effect on Akt signaling. Recent studies have shown that curcumin dose- and time-dependently inhibits the phosphorylation of Akt and mammalian target of rapamycin (mTOR), and their downstream targets in prostate cancer cells [127]. Inhibition of the Akt/mTOR pathway by curcumin results in suppression of the growth of SCC40 xenografts, and curcumin at 15 mg significantly increases in survival in the 4-nitroquinoline 1-oxide--induced head and neck squamous cell carcinoma survival study [128].

Curcumin has also been shown to inhibit the proliferation of cisplatin-resistant ovarian cancer cells via the inhibition of Akt activation [129]. It has also been reported that an analog of curcumin, 4-hydroxy-3-methoxybenzoic acid methyl ester (HMBME), targets the Akt signaling pathway, inhibits the proliferation of cancer cells and induces apoptosis [130]. Likewise, HMBME was shown to decrease the level of phosphorylated Akt, inhibit Akt kinase activity, and reduce the DNA-binding activity of NF-κB [130]. Several reports also suggest that curcumin has molecular targets within the Akt signaling pathways and that the inhibition of Akt activity may facilitate inhibition of proliferation and induction of apoptosis in cancer cells [131,132]. A curcumin derivative, diphenyl difluoroketone, significantly inhibited the colon cancer xenograft Akt and ERK phosphorylation in mice [133]. In other study, solubilized curcumin effectively blocked brain tumor formation in the mice that had already received an intracerebral bolus of mouse melanoma cells (B16F10) by suppressing p-Akt, Cyclin D1, p-NF-κB, Bcl-xL and VEGF [134].

4.3. Quercetin

Quercetin, a powerful anti-oxidant, is consumed by humans as part of their diet [135]. Although the quercetin content of foods has not been systematically analyzed, it is found in many fruits, vegetables and beverages. The chemopreventive effects of quercetin against DNA damage and precancerous changes in cells were recently demonstrated both in vitro and in vivo. Quercetin was found to arrest the progression of cervical neoplasia in Swiss albino mice [136].

Quercetin may offer a defense against the detrimental effects of carcinogenic chemicals and can induce apoptosis via the mitochondrial pathway [137]. Although the anti-carcinogenic mechanisms of quercetin are not well known, quercetin specifically inhibits p21-Ras expression in human colon cancer cell lines and in primary colorectal cancers [138]. Reports suggest that quercetin has DNA-damaging and pro-oxidant property in cells [139]. One proposed mechanism involves the inhibition of Akt/protein kinase B (PKB) phosphorylation, an upstream kinase of the pro-survival protein kinase cascade involving PI3K. Significant down-regulation of Bcl-2 and Bcl-xL along with Cu-Zn SOD, which could lead to an increase in ROS, has been reported after quercetin treatment in certain human cancer cell lines [43,140].

Survivin, which binds directly to and inactivates caspases
was inhibited by quercetin, resulting in the activation of caspases [43]. One promising therapeutic approach for the induction of apoptosis in cancer cells is using TNF-related apoptosis-including ligand (TRAIL) [141]. Quercetin has been shown to potentially arrest human prostate cancer cells and mediate activation of caspase and poly(ADP-ribose)polymerase (PARP) cleavage [142, 143]. A combined treatment of TRAIL and quercetin was found to enhance TRAIL-induced cytotoxicity by activating caspases and inhibiting phosphorylation of Akt [144].

A principal approach for treatment of tumors is based on the fact that cancer cells are resistant to CD95-mediated apoptosis. Alterations in the CD95 system result in the escape of tumor cells from this defense system. Sensitivity to CD95 can be restored by treating the cells with quercetin, thus making the cancer cells susceptible to apoptosis [145]. One study has demonstrated the inhibitory effects of quercetin on H2O2-induced apoptosis via mediation of the AP-1-mediated apoptotic pathway. The mechanistic action of quercetin as an anti-apoptotic compound is attributed to its ability to inhibit MAPK pathways and reduce expression of genes participating in the JNK-c-JUN/AP-1 [146].

In an azoxymethane (AOM)-induced rat colon cancer model, dietary administration with quercetin and curcumin decreased the number of aberrant crypt foci (ACF), putative precursor lesions for colonic adenocarcinoma, by 4- and 2-fold, respectively, compared with the controls [137]. Western blot analyses of caspase-9, Bax (pro-apoptotic) and Bcl-2 (anti-apoptotic) proteins from colon scrapings suggest that quercetin and curcumin induce apoptosis via the mitochondrial pathway. Sun et al. reported that quercetin significantly prevented in vivo growth of human salivary adenoid cystic carcinoma xenografts in nude mice, accompanied by induction of tumor cell apoptosis, suppression of NF-κB nuclear translocation, as well as down-regulation of Akt and IκB kinase-α activation. Thus, quercetin would be a promising chemotherapeutic agent through its function of down-regulating the PI3K/Akt/IKK-alpha/NF-κB signaling pathway [147].

5. COMBINATION THERAPY

Some naturally occurring chemopreventive compounds are known to act in synergy with other chemopreventive or anti-cancer agents, as listed in Table 2. The anti-neoplastic agents have dose-limiting toxicity and drug resistance, thus limiting their clinical application. Development of novel strategies that overcome radio- and chemo-resistance and sensitize cancer cells to anti-neoplastic agent can enhance the therapeutic effect of these drugs. Combination treatment with EGCG and tamoxifen resulted in synergistic growth inhibition of MDA-MB-231 [150]. Curcumin and gemcitabine treatment decreased pancreatic tumor volume in vivo model [151, 152]. Furthermore, combination of quercetin with sulfuraphane, an isothiocyanate enriched in broccoli, exerted synergistic effects by improving apoptosis resistance [153]. Liposomal forms of curcumin plus resveratrol significantly decreased prostatic adenocarcinoma in prostate-specific PTEN-knockout mice by effectively inhibiting cell growth and inducing apoptosis [154].

Bcl-2 family proteins are regulators of chemoresistance and radioresistance in cancer. In vivo treatment with quercetin and trans-3, 5-dimethoxy-4'-hydroxystilbene (t-PTER) altered expression of molecules involved in regulating cancer cell resistance to drugs and radiations [155]. Combined administration with t-PTER+ quercetin, FOLFOX6 (oxaliplatin, leukovorin, and 5-fluorouracil, a first-line chemotherapy regimen), and radiotherapy eliminates colorectal cancer cells growing in vivo leading to long-term survival. Gene expression analysis of a Bcl-2 family of genes revealed that down-regulation of bcl-2 expression via inhibition of NF-κB activation. Curcumin also potentiates the anti-tumor effects of radiation therapy in colorectal cancer by suppressing NF-κB and NF-κB-regulated gene products, leading to inhibition of proliferation and angiogenesis [123].

In mouse cervical multi-stage squamous cell carcinoma model using 3-methylcholanthrene and a xenograft model of human cervical cancer in mice, the combined treatment with curcumin and paclitaxel induced a synergistic reduction in the tumor incidence as well as tumor volume compared with the individual treatment of paclitaxel or curcumin [156], suggesting that a sub-optimal concentration of curcumin augments the anti-tumor action of paclitaxel by downregulating the activation and downstream signaling of anti-apoptotic factors and survival signals such as NF-κB, Akt and MAPKs that have significant roles in proliferation, survival, angiogenesis and metastasis. By inhibition of NF-κB activity, curcumin augments the anti-tumor action of cisplatin enhancing growth suppression in vivo [157].

Recently, preclinical investigation revealed that combination therapy with curcumin and dasatinib to be highly effective causing an over 95% regression of intestinal adenomas in Apcmin/+ mice, which could be attributed to decrease proliferation and increased apoptosis [158].

6. SUMMARY AND PERSPECTIVES

The pathogenesis of many chronic diseases, including cancer, has been associated with abnormally regulated apoptosis [14, 75, 77, 78, 159]. The synergistic combination of an undesirable proliferative stimulus and an associated
defect in the apoptotic pathway seems universal in cancer [76-80]. Epidemiological studies indicate that dietary habits contribute to, at least, one third of all human cancers [160], and suggest that certain dietary components can exacerbate or interfere with carcinogenesis. Apoptosis is likely to be a crucial mechanism in the chemopreventive properties associated with such dietary factors [14].

In addition to the conventional therapeutic agents, numerous dietary components and micronutrients are emerging, which possess considerable potential for hindering in vivo deleterious oxidative processes and inducing apoptosis of cancerous or pre-cancerous cells [160,161], and are therefore being considered as promising chemopreventive agents. A range of dietary compounds can modulate apoptosis and those with pro-apoptotic properties exhibit beneficial effects in animal and in vitro studies by eliminating cancerous cells [9,14,92]. Moreover, some dietary compounds have also shown beneficial effects in clinical trials [93].

A balance of cell proliferation and apoptosis normally maintains cellular homeostasis. However, apoptosis is a very complex process with numerous specific targets within each arm of the apoptotic pathways. Nevertheless, it is very encouraging that single bioactive dietary agents can directly and indirectly influence many of the targets within the apoptotic pathway. In addition, many of these dietary agents appear to exhibit some degree of specificity for neoplastic cells. Furthermore, the protective effects of single agents can be potentiated and/or synergized by other dietary factors suggesting the possibility of combinational approaches for chemoprevention. While dietary interventions seem encouraging for devising new chemopreventive strategies, there are several issues remaining that need to be fully understood. The dose of each agent, duration of exposure, relative bioavailability of each dietary compound and potentially adverse side effects and/or interactions should be considered.

Further research is required to identify the phytochemical-specific molecular mechanisms of the huge number of already recognized bioactive dietary chemopreventive agents. The potential benefits of cancer chemoprevention appear promising given the data obtained from clinical trials and pre-clinical studies.

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