Breeding and Analysis of Two New Grapefruit-Like Varieties with Low Furanocoumarin Content

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Abstract

Furanocoumarins (FCs) are a group of related plant defense metabolites occurring in several plant families, including some species in the genus citrus, such as grapefruit and pummelo. FCs function as toxins against pathogens, insects and other plant pests and some are toxic to humans at high levels. Although the levels of FCs in grapefruits are non-toxic to humans, they inhibit the intestinal enzyme CYP3A, thus preventing degradation of medicines, such as statins, and causing dangerous overdose effects. This overdosing can cause devastating side effects, ranging from stomach bleeding to kidney problems, muscle aches and irregular heartbeats. In the present study, we utilize LC/MS to characterize the levels of FCs pathway intermediates and end products in twelve citrus cultivars, including mandarin (Citrus reticulata), orange (Citrus sinensis (L.) Osbeck), Pummelo (Citrus maxima (Burm.) Merr.), grapefruit (Citrus paradisi Macf.), and two newly selected grapefruit-like varieties [(Citrus reticulata) X (Citrus maxima) (Burm.) Merr]. The orange and mandarin varieties do not contain FCs or FCs precursor compounds suggesting that this biosynthetic pathway is absent or inactive in mandarins and oranges and therefore a good genetic source for null alleles to FCs biosynthesis. We report the selection and characterization of two new low FCs seedless grapefruit-like varieties, “Aliza” and “Coocki”, developed by a cross between pummelo and mandarin. Fruits of these varieties resemble grapefruit and contain high levels of the flavanone naringin, typical of grapefruit, but contain only trace amounts of FCs (based on LC/MS analysis). Based on the variability of FCs content and inheritance in citrus species, the results suggest that future development of new low-FCs grapefruit varieties is an achievable objective.

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1. Introduction

Furanocoumarins (FCs) are plant defense compounds that function as toxins against pathogens, insects and other plant pests [1] [2]. Insects exposed to linear FCs suffer decreased larval weight, extended generation time and increased mortality [3]. High doses of some FCs are also toxic to humans; among maladies caused in humans by FCs are “celery picker’s itch” and “bartender’s itch”, caused by contact with FCs-containing celery plants or oil of lime peels, respectively. Also, FCs can cause temporary or even permanent blindness if introduced into the eyes. Some FCs are also carcinogenic and teratogenic, i.e. they can cause cancer and birth defects, respectively [3]-[5].

FCs are found in several plant families, including edible plant species such as the citrus species pummelo and grapefruit, but the levels of mutagenic FCs (such as psoralen) in pummelo and grapefruits are rather low, and therefore there is no risk associated with consumption of their fruit and juice for healthy individuals [3]. Citrus is one of the most important fruit crops worldwide with total annual production of 115.5 million tons, to which grapefruit contributes about 5.5% [6]. However, in recent years, there has been a decline in grapefruit consumption, especially in the US, as a result of the “grapefruit juice effect”, attributed to FCs: FC-containing grapefruit juice negatively affects the body’s degradation of medicines, including statins, leading to overdose effects.

The mechanism of the “grapefruit juice effect” is based on FCs that inhibit the intestinal drug catabolic enzyme CYP3A, thereby leading to accumulation of medicines, which in turn can lead to harmful effects ranging from relatively mild hypotension and dizziness in the case of some calcium-channel blockers, to potentially severe nephrotoxicity in the case of some immunosuppressant drugs [5] [7]-[16].

Drugs that interact with FCs [17] include:

**ANTICANCER**
- Dasatinib (leukaemia)
- Erlotinib (lung cancer and pancreatic cancer)
- Everolimus (kidney cancer)
- Lapatinib (breast cancer)
- Nilotinib (leukaemia)
- Pazopanib (kidney cancer)
- Sunitinib (kidney/gastrointestinal cancer)
- Vandetanib (thyroid cancer)
- Venurafenib (skin cancer)

**ANTI-INFECTIVE**
- Erythromycin (antibiotic)
- Halofantrine (antimalaria)
- Maraviroc (antiHIV)
- Primaquine (antimalaria)
- Quinine (antimalaria)
- Rilpivirine (antiHIV)

**ANTICHOLESTEROL**
- Atorvastatin
- Lovastatin
- Simvastatin

**CARDIOVASCULAR**
- Amiodarone (heart rhythm disorders)
- Apixaban (anticlotting)
Dronedarone (heart rhythm disorders)
Eplerenone (heart failure)
Felodipine (high blood pressure/angina)
Nifedipine (high blood pressure/angina)
Quinidine (heart rhythm disorders)
Rivaroxaban (blood anticlotting)
Ticagrelor (blood anticlotting after heart attack)

CENTRAL NERVOUS SYSTEM
Oral alfentanil (painkiller)
Oral fentanyl (painkiller)
Oral ketamine (painkiller, sedative)
Lurasidone (schizophrenia/mental health problems)
Oxycodone (painkiller)
Pimozide (schizophrenia/other mental health problems)
Ziprasidone (schizophrenia, mania, bipolar disorder)

GASTROINTESTINAL
Oral alfentanil (painkiller)
Oral fentanyl (painkiller)
Oral ketamine (painkiller, sedative)
Lurasidone (schizophrenia/mental health problems)
Oxycodone (painkiller)
Pimozide (schizophrenia/other mental health problems)
Ziprasidone (schizophrenia, mania, bipolar disorder)

IMMUNOSUPPRESSANTS
Cyclosporine (post organ transplant, rheumatoid arthritis, psoriasis)
Sirolimus (post organ transplant)
Tacrolimus (post organ transplant)

URINARY TRACT
Solifenacin (frequent urination/incontinence)
Silodosin (enlarged prostate)
Tamsulosin (enlarged prostate)

The FC-biosynthesis pathway starts with the amino acid phenylalanine (Figure 1) via umbelliferone to obtain linear or angular FCs. In citrus, only the linear FC-biosynthesis pathway is present which leads to formation of demethyl suberosin by alkylation of umbelliferone (Figure 1). Three FCs—6,7-dihydroxybergamottin (6,7-DHB), bergamottin and paradisin C, appear to co-segregate genetically, which suggests that these three compounds are tightly linked metabolically as co-products of the “bergamottin pathway” [19]. Ranked in descending order of their inhibitory potency, the FCs found in grapefruit are: paradisin C > 6,7-DHB > bergamottin > isoimperatorin > bergaptol [9] [20].

In the present study, we report on variations in the biosynthesis levels of various FCs in diverse varieties of grapefruit and pummelo. Twelve citrus cultivars (Table 1), including four pummelos, two grapefruits, one mandarin, one orange and three newly selected grapefruit-like varieties, were tested for juice and leaf levels of five furanocumarins (FCs)—bergamottin, bergaptol, epoxy bergamottin, 6,7-dihydroxybergamottin and isoimperator— and two FC precursors-psoralen and umbelliferone. The results show that the orange and the mandarin do not contain FCs; moreover, the lack of FC precursor compounds suggests that the biosynthesis pathway is entirely not active in mandarins and oranges. We also observe great diversity in the accumulation of FCs and their precursor compounds in various grapefruit and pummelo cultivars. Finally, we describe two new low-FC and seedless grapefruit-like cultivars, “Aliza” and “Cookie” that were selected in our breeding program by induced mutagenesis [21].

2. Materials and Methods
2.1. Materials and Chemicals
Bergamottin (purity >99%) was purchased from Fluka (Buchs, Switzerland); bergaptol (>99%) and naringin (purity >99%) from Extrasynthese (Genay, France); epoxybergamottin (99%), 6,7-DHB (99%) and isoimperatorin (99%) from Herboreal Ltd. (Dalkeith, United Kingdom); andpsoralen (>99%) and umbelliferone (99%); from Sigma-Aldrich (Steinem, Germany).
Precursors of Furanocoumarins

phenylalanine

\[
\text{PAL} \quad \text{PAL}
\]

cinnamic acid

coumaric acid

\[
\text{C4H} \quad \text{C4H}
\]

Precursors of Furanocoumarins

phenylalanine

cinnamic acid

coumaric acid

\[
\text{PAL} \quad \text{PAL}
\]

DMAPP

prenyltransferase

\[
\text{DMAPP} \quad \text{DMAPP}
\]

marmesin synthase

marmesin

\[
\text{O}_2, \text{NADPH} \quad \text{O}_2, \text{NADPH}
\]

psoralen synthase

psoralen
Figure 1. The linear FC biosynthesis pathway from phenylalanine to 6,7-dihydroxybergamottin (6,7-DHB). The chemical structures were taken from the PubChem database [18].
<table>
<thead>
<tr>
<th>Cultivar</th>
<th>Group</th>
<th>♂️</th>
<th>♀️</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tahitian Pummelo</td>
<td>[Citrus maxima (Burm.) Merr.]</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td>Chandler Pummelo</td>
<td>[Citrus maxima (Burm.) Merr.]</td>
<td>“Siamese Pink” pummelo [Citrus maxima (Burm.) Merr.]</td>
<td>“Siamese Sweet” pummelo [Citrus maxima (Burm.) Merr.]</td>
</tr>
<tr>
<td>Marsh Grapefruit</td>
<td>[Citrus paradisi Macf.]</td>
<td>Orange [Citrus sinensis (L.) Osbeck]</td>
<td>Pummelo [Citrus maxima (Burm.) Merr.]</td>
</tr>
<tr>
<td>Duncan Grapefruit</td>
<td>[Citrus paradisi Macf.]</td>
<td>Orange [Citrus sinensis (L.) Osbeck]</td>
<td>Pummelo [Citrus maxima (Burm.) Merr.]</td>
</tr>
<tr>
<td>Valencia Orange</td>
<td>Orange [Citrus sinensis (L.) Osbeck]</td>
<td>Citrus reticulata</td>
<td>Pummelo [Citrus maxima (Burm.) Merr.]</td>
</tr>
<tr>
<td>Flamingo Pummelo</td>
<td>[Citrus maxima (Burm.) Merr.]</td>
<td>“Chandler” pummelo [Citrus maxima (Burm.) Merr.]</td>
<td>“Tahitian” pummelo [Citrus maxima (Burm.) Merr.]</td>
</tr>
<tr>
<td>Hanna Pummelo</td>
<td>[Citrus maxima (Burm.) Merr.]</td>
<td>“Chandler” pummelo [Citrus maxima (Burm.) Merr.]</td>
<td>“Tahitian” pummelo [Citrusmaxima (Burm.) Merr.]</td>
</tr>
<tr>
<td>Einat A red triploid</td>
<td>“Oroblanco”-like citrusfruit</td>
<td>Tetraploid “Hudson” [Citrus paradisi Macf.]</td>
<td>Acid-freepummelo [Citrus maxima (Burm.) Merr.]</td>
</tr>
<tr>
<td>Aliza Grapefruit-like</td>
<td>“Orah” mandarin [Citrus reticulata Blanco]</td>
<td>“Chandler” pummelo [Citrus maxima (Burm.) Merr.]</td>
<td></td>
</tr>
<tr>
<td>Dany Grapefruit-like</td>
<td>“Michal” mandarin [Citrus reticulata Blanco]</td>
<td>“Duncan” grapefruit [Citrus paradisi Macf.]</td>
<td></td>
</tr>
<tr>
<td>Cookie (seedless mandelo) Grapefruit-like</td>
<td>“Frua” mandarin [Citrus reticulata]</td>
<td>“Siamese Sweet” pummelo [Citrus maxima (Burm.) Merr.]</td>
<td></td>
</tr>
</tbody>
</table>

ND = not determined.

2.2. Extraction of Furanocoumarins

FCs were extracted from fruit juice and leaves. For the juice analysis, 10 ml of juice of each cultivar were mixed with an equal volume of ethyl acetate. For the leaf analysis, 10 g of leaf tissue were ground in liquid nitrogen and mixed with ethyl acetate at a volume equivalent to the weight, i.e. 10 ml. The extraction was performed by vortex mixing for 5 min at 25°C followed by centrifugation at 3220 g for 25 min at 25°C. The organic phase was collected and re-extracted with an equivalent volume of double-distilled water. The mixture was then mixed and centrifuged as described above. A 2-ml aliquot of the organic phase was evaporated under air flow and dissolved in 1 ml of acetonitrile for LC-MS analysis.

2.3. Extraction of Naringin

The fruit slices were blended in a blender for 1 min and the blended material was mixed with butanol (1:1; v/v), centrifuged at 4°C at 10,000 g for 15 min, and the butanol fraction was centrifuged again at 10,000 g for 10 min. A 3-ml aliquot of the upper phase was dried in a speed-vacuum centrifuge, and the precipitate was dissolved in 5 ml of 100% methanol and filtered through a 0.45-µm filter. LC-MS analyses were conducted with a UPLC-Triple Quadrupole-MS (Waters Xevo TQ MS). The samples were again filtered, through a 0.22-µm Millex-HV Durapore (PVDF) membrane, before injection into the LC-MS apparatus. Separation was performed on 2.1 × 50 mm i.d., 1.7 µm UPLC BEH C18 column. Chromatographic and MS parameters were as follows. The mobile phase consisted of water (phase A) and 0.1% formic acid in acetonitrile (phase B). The linear-gradient program was as follows: 100% to 95% A over 0.1 min, 95% to 5% A over 5 min, held at 5% A for 2 min, then back to the initial conditions (95% A) for 3 min. The flow rate was 0.3 ml·min⁻¹ and the column temperature was kept at 35°C. All of the analyses were performed with the ESI source in positive-ion mode, with a capillary voltage of 3.2 kV, a cone voltage of 30 V, a desolvation temperature of 350°C, a desolvation gas flow rate of
850 L/h and a source temperature of 150°C. Quantization was performed by MRM acquisition by monitoring the 581/152, 581/273 transitions, RT = 6.07, (dwell time of 161 ms for each transition) for naringin.

### 2.4. LC-MS Analyses

LC-MS analyses were conducted with a UPLC-Triple Quadrupole-MS (Waters Xevo TQ MS. Samples were filtered through a 0.22-µm Millex-HV Durapore (PVDF) membrane before injection into the LC-MS apparatus. Separation was performed with a 2.1 m×50 mm i.d., 1.7 µm UPLC BEH C18 column. Chromatographic and MS parameters were as follows. The mobile phase consisted of water (phase A) and 0.1% formic acid in acetonitrile (phase B). The linear-gradient programme for bergamottin, epoxy bergamottin, 6,7-dihydroxybergamottin, bergaptol and isomperatorin was as follows: 100% to 95% A over 0.1 min, 95% to 5% A over 8 min, held at 5% A for 3 min, then back to the initial conditions (95% A) for 3 min. The linear-gradient programme for psoralen and umbelliferone was as follows: 100% to 95% A over 0.1 min, 95% to 5% A over 4 min, held at 5% A for 3 min, then back to the initial conditions (95% A) for 3 min. The flow rate was 0.3 ml·min$^{-1}$ and the column temperature was kept at 35°C. All of the analyses were performed with the ESI source used in positive-ion mode, with a capillary voltage of 3.2 kV, a cone voltage of 30 V, a desolvation temperature of 350°C, a desolvation gas flow of 650 L/h and a source temperature of 150°C. Quantitation was performed with MRM acquisition by monitoring the following transitions 339/147, 339/203 (RT = 7.1, dwell time of 161 ms) for bergamottin; 355/153, 355/203 (RT = 5.6, dwell time of 78 ms) for epoxy bergamottin; 373/153, 373/203 (RT = 4.2, dwell time of 78 ms) for 6,7-DHB; 202/131, 202/146 (RT = 3.1, dwell time of 161 ms) for bergaptol; 271/146, 271/203 (RT = 5.5, dwell time of 161 ms) for isomperatorin; 187/115, 187/13 (RT = 2.4, dwell time of 78 ms) for psoralen and 163/91, 163/107 (RT = 1.8, dwell time of 78 ms) for umbelliferone.

### 2.5. Statistical Analysis

One-way ANOVA and Turkey’s HSD pair-wise comparison tests were applied by means of JMP statistical software, version 7 (SAS Institute Inc., Cary, NC, USA) and Microsoft’s Office Excel.

### 3. Results and Discussion

FCs were found at varied concentrations in different cultivars, but were completely absent from mandarins, oranges and were close to absent in the new grapefruit-like varieties “Aliza” and “Cookie” (Figure 2). In grapefruit and pummelo, Bergamottin and 6,7-DHB were found at higher concentrations than other FCs, probably because they are final products that accumulate over time. In correlation, the FC precursors are present in very low concentrations in grapefruit and pummelo (Figure 2), as they are utilized for biosynthesis of the final products. Interestingly, although the FC biosynthesis pathway of grapefruit (Figure 1) was inherited from pummelo, the accumulated concentrations of bergamottin and 6,7-DHB in the grapefruit cultivars “Duncan” and “Marsh” (a seedless mutation of “Duncan”) were much higher than those observed in the pummelo cultivars that we examined (Figure 2).

The complete absence of FCs and FC precursors, from umbelliferone to psoralen in oranges and mandarins (Figure 2), suggests that the entire pathway for biosynthesis of FC is absent or inactive in these species, and that only a null allele for FC biosynthesis can be inherited from these species. The new selection “Aliza”, a hybrid of “Chandler” pummelo and “Orah” mandarin is almost completely void of FCs, suggesting the pummelo parent is heterozygous for FC biosynthesis (i.e. contains one active copy and one inactive copy, presumably inherited by “Aliza”). Furthermore, the finding that the pummelo hybrid “Hana” (cross between “Chandler” and “Tahitian” pummelos) has no FCs (Figure 2) indicates that “Chandler” probably also contributed its inactive allele for FC biosynthesis to “Hana”. In contrast, “Chandler” contributed its active allele for FC biosynthesis to the pummelo “Flamingo” (a different cross between “Chandler” and “Tahitian” pummelos) (Figure 2); the rather higher level of 6,7-DHB in “Flamingo” might be a result of the “release” of the isomperatorin that accumulates in “Tahitian”, by the active allele of “Chandler”.

The concentrations of bergamottin and 6,7-DHB observed in 4- to 5-month-old leaves were extremely low relative to those observed in the fruits. Pummelo cultivars have much lower concentrations of epoxy bergamottin in their fruits versus other FCs compounds (Figure 2(a)), but the accumulated amounts of this FC in their leaves were equivalent to the highest concentrations observed among all of the examined cultivars (Figure 2). We note that, during commercial processing, epoxy bergamottin may be hydrolyzed to 6,7-DHB, which has been identi-
Figure 2. Analysis of FC levels in 12 citrus cultivars. FCs were analyzed in fruit (a) and leaf (b) tissues. Data are means of 3 replications. Different letters above columns indicate significant differences at $p \leq 0.05$. 
fied as a potentially important inhibitor of CYP3A4 [17] [22].

In all of the fruit juices of the examined cultivars, bergapten and isoimperatorin were found at significantly lower concentrations than the other FCs, reflecting the fact that they are intermediate products in the pathway. However, pummelo “Tahitian” contained an extraordinarily high concentration of isoimperatorin; this pummelo also had very low concentrations of bergamottin and 6,7-DHB, suggesting that the pathway in this variety is partially blocked and it accumulates isoimperatorin instead of bergamottin and 6,7-DHB.

We have selected two new seedless grapefruit-like cultivars, “Aliza” and “Cookie” (Table 1), which do not accumulate FCs. At the start of ripening “Aliza” fruits have green skin and yellow flesh, but the former turns yellow later in the season, and when fully ripe these fruits have orange skin and orange flesh (Figure 3). By inducing mutations, we selected seedless types of “Aliza” and “Cookie” (Figure 3 and Figure 4). Both of these selections have unique and favorable characteristics: they are juicy and, because they contain naringin (Table 2), they have been classified as grapefruit-like cultivars; “Aliza” and “Cookie” are also easy to peel.

4. Conclusion

FC composition and concentrations are highly variable in citrus, suggesting that selection for new grapefruit like cultivars with low FCs is a feasible objective. Indeed, we describe the selection of two new low FC grapefruit-like varieties that are potentially safe for consumption by humans using statins and other drugs. The corre-
Figure 4. Photographs of “Cookie”, a new grapefruit-like cultivar.

Table 2. Quality traits and naringin contents in juices of 12 different citrus cultivars. Data are means ± S.E. of 3 replications.

<table>
<thead>
<tr>
<th>Cultivar</th>
<th>Seeds (no. ± SE)</th>
<th>Sugar (Bx ± SE)</th>
<th>Acid (% ± SE)</th>
<th>Juice (% ± SE)</th>
<th>Size (g ± SE)</th>
<th>Naringin (ppb ± SE)</th>
<th>Ripening season</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tahitian</td>
<td>120.00 (±6.00)</td>
<td>12.00 (±0.07)</td>
<td>1.50 (±0.04)</td>
<td>30.50 (±2.50)</td>
<td>542.00 (±28.00)</td>
<td>66690.56 (B)</td>
<td>October-November</td>
</tr>
<tr>
<td>Chandler</td>
<td>84.00 (±6.00)</td>
<td>12.00 (±0.04)</td>
<td>0.80 (±0.02)</td>
<td>9.00 (±0.80)</td>
<td>684.00 (±34.00)</td>
<td>144550.72 (A)</td>
<td>October-November</td>
</tr>
<tr>
<td>Flamingo</td>
<td>0.30 (±0.20)</td>
<td>10.70 (±0.10)</td>
<td>0.90 (±0.5)</td>
<td>19.10 (±1.50)</td>
<td>1218.00 (±60.00)</td>
<td>75495.57 (B)</td>
<td>November-January</td>
</tr>
<tr>
<td>Hanna</td>
<td>26.00 (±4.00)</td>
<td>9.50 (±0.23)</td>
<td>0.60 (±0.07)</td>
<td>23.80 (±1.50)</td>
<td>917.00 (±60.00)</td>
<td>86314.11 (B)</td>
<td>November-February</td>
</tr>
<tr>
<td>Duncan</td>
<td>55.00 (±5)</td>
<td>11.80 (±0.15)</td>
<td>2.30 (±0.04)</td>
<td>33.80 (±1.60)</td>
<td>339.00 (±16.00)</td>
<td>88589.18 (B)</td>
<td>October-April</td>
</tr>
<tr>
<td>Marsh</td>
<td>3.00 (±1.00)</td>
<td>11.20 (±0.12)</td>
<td>2.00 (±0.04)</td>
<td>35.60 (±9.00)</td>
<td>372.00 (±3789.91)</td>
<td>42496.21 (C)</td>
<td>October-April</td>
</tr>
<tr>
<td>Eimat</td>
<td>2.70 (±1.30)</td>
<td>9.50 (±0.3)</td>
<td>1.80 (±0.05)</td>
<td>23.70 (±30.00)</td>
<td>687.00 (±3789.91)</td>
<td>67598.29 (B)</td>
<td>November-February</td>
</tr>
<tr>
<td>Valencia</td>
<td>4.00 (±0.70)</td>
<td>9.60 (±0.25)</td>
<td>1.40 (±0.05)</td>
<td>45.90 (±8)</td>
<td>263.00 (±8)</td>
<td>0.00 (E)</td>
<td>February-May</td>
</tr>
<tr>
<td>Dany</td>
<td>9.00 (±1.40)</td>
<td>11.00 (±0.09)</td>
<td>1.70 (±0.06)</td>
<td>22.10 (±21.00)</td>
<td>458.00 (±0.00)</td>
<td>0.00 (E)</td>
<td>January</td>
</tr>
<tr>
<td>Aliza</td>
<td>0.00 (±0.20)</td>
<td>10.40 (±0.03)</td>
<td>0.70 (±0.03)</td>
<td>42.80 (±1.60)</td>
<td>621.00 (±58.00)</td>
<td>31124.00 (CD)</td>
<td>November-February</td>
</tr>
<tr>
<td>Cookie</td>
<td>0.00 (±0.20)</td>
<td>11.00 (±0.04)</td>
<td>0.50 (±2.00)</td>
<td>38.50 (±25.00)</td>
<td>486.00 (±1839.04)</td>
<td>13865.00 (DE)</td>
<td>October-November</td>
</tr>
<tr>
<td>Orah</td>
<td>16.00 (±1.60)</td>
<td>12.10 (±0.18)</td>
<td>1.10 (±0.04)</td>
<td>35.70 (±5.00)</td>
<td>155.00 (±0.00)</td>
<td>0.00 (E)</td>
<td>January-March</td>
</tr>
</tbody>
</table>

lation of FC accumulation between leaves and fruits provides a potential marker for screening young plantlets for additional FC-free grapefruit-like varieties.

References


