Determination of Cichoric Acid as a Biomarker in Echinacea Purpurea Cultivated in Iran Using High Performance Liquid Chromatography

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Abstract

Echinacea purpurea (Purple coneflower) is an immunostimulating drug, containing multiple substances. The most important substance in activity is polysaccharide, caffeic acid derivatives (cichoric acid), alkamides and glycoproteins. It is not clear yet, which substances are responsible for activity. Cichoric acid is an appropriate marker of the quality of Echinacea purpurea containing product, because it has immune stimulatory effects and it is susceptible to degradation. In this study a TLC scanner system and HPLC method has been used for identification and determination of cichoric acid in aerial parts of Echinacea purpurea. The results showed that the cichoric acid content of Echinacea purpurea cultivated in Iran is about 1.50 ± 0.65% (w/w) which is comparable with cichoric acid content in native plants. The local conditions have no significant effect on cichoric acid content as a biomarker of Echinacea purpurea quality.

Keywords: Echinacea Purpurea, Cichoric Acid, TLC, HPLC, Iran

1. Introduction

Herbal medicine has lower side effects in comparison with chemical drugs. Attention and stimulation for using herbal drugs are increasing in recent years. Systematic study about characteristics properties of herbal drug has been increased. Echinacea purpurea Monech (purple coneflower), originated in the United States of America and was brought to Europe in the late 19th century [1]. Recently, Echinacea purpurea has been cultivated in some parts of Iran (e.g. Isfahan) and used in some herbal drugs [2]. It exhibits antioxidative, antibacterial, antiviral and antifungal activities, but most of all it affects on various immune parameters [3-21]. Because of its immuno-enhancing activity, it has been recently used in AIDS therapy [22]. It is used for treating flu and cold, against which symptomatic treatment is an established procedure.

The most important potential active compounds in purple coneflower are polysaccharides, caffeic acid derivatives (especially cichoric acid), alkamides and glycoproteins [6,7]. Cichoric acid is an appropriate marker because it has immune stimulatory effects and is probably the most effective substance in Echinacea purpurea. Climate and cultivation conditions could affect cichoric acid content in Echinacea purpurea. Recently, nevertheless, the lack of studies on the adaptability of the culture, the productivity and the quality of the product was apparent. Various methods for determination of cichoric acid such as HPLC, and capillary electrophoresis have been used [22-31].

In this study, a TLC scanner system and an HPLC method was used for identification and quantification of cichoric acid content in aerial parts of Echinacea purpurea which is cultivated in Iran. The result was compared with native plants in Europe and America.

2. Experimental

2.1. Chemicals

2-aminoethyl diphenyl borinate (Sigma), caffeic acid, chlorogenic acid and all solvents; methanol, ethyl acetate, formic acid was analytical grade and purchased from Merck. All solvents used for running HPLC, were HPLC grade. Dionized, double distilled water was used.
2.2. Samples

Several different batches of aerial parts of aerated and dried Echinacea purpurea were collected and ground with a 40 mm mesh to a fine powder. Samples were prepared by solving the powder in methanol/water (70:30) with the ratio of 1 g in 10 ml using an ultrasonic bath for approximately 5 min and then filtered. Chlorogenic acid and caffeic acid were dissolved in alcohol/water solutions which are stable at least for 30 hours if light protected.

2.3. TLC and HPLC

The CAMAG TLC system with auto sampler and scanner which is controlled by Wincats software was used for identification. The plates were prepared by TLC silica gel HF254 (5 µm). The mobile phase composed of anhydrous formic acid, ethyl acetate and water (2:1:17). Sample solution and standard solution were prepared respectively by dissolving 0.5 g powder in 5 ml solvent and 0.05 mg of caffeic acid and 0.05 mg of chlorogenic acid in 10 ml solvent 25 µl of sample solution and 10 µl of standard solutions were applied as TLC bands [31,32]. For detection of stains under UV 366 lamp, 2-amino diphenyl borinate in methanol (10 mg/ml) was used as reagent which is famous as natural product reagent.

2.4. High Performance Liquid Chromatography (HPLC)

The Cecil 1100 HPLC system was used for quantitative assay. This system equipped with a 20 µl injection loop, a C18 Lichrosorb column with 0.25 m length, and 4.6 mm diameter and UV-Vis detector which was set at 330 nm. Acetonitrile and phosphate buffer (phosphoric acid/water (1:99 V/V) were used as mobile phases in a gradient elution of solvents with 1.5 ml/min flow rate according to set programs in EP and USP [31,32]. Sample solution and standard solution were prepared by dissolving 0.25 mg of powder and 0.47 mg of chlorogenic acid in 10 ml solvent respectively. Chlorogenic acid was used as external standard which was cheaper and available in Iran [22].

The chromatograms related to standard and sample are shown in Figures 4 and 5 respectively. According to literatures the relative retention time for cichoric acid respect to chlorogenic acid is about 2.3 [31]. While the chlorogenic acid peak appears at 8 minute, the cichoric the ratio of slope of the two calibration curves prepared with standard solutions [22,31,32].

3. Results and Discussion

The chemical structures of Cichoric acid (a), Chlorogenic acid (b) and Caffeic acid (c) are shown in Figure 1. Figure 2 shows the image of TLC plate of sample and standard solutions. In the left track, the yellow fluorescent zones are related to chlorogenic acid (Rf = 0.45) and caffeic acid (Rf = 0.84) contained in the standard solutions. In the right track of TLC plate, the fluorescent zones are related to caftaric acid (Rf = 0.52) and cichoric acid (Rf = 0.86) contained in sample solution. Figure 3, clearly confirmed the presence of cichoric acid and absence of chlorogenic acid in sample solution [31,32].

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\[
C_u\% = 100 \left( \frac{A_u \times C_s \times 0.695}{A_s \times C_t} \right)
\]

Figure 1. (a) The structure of Cichoric acid, (b) Chlorogenic acid and (c) Caffeic acid.
Figure 2. TLC image of sample solution containing caftaric acid and cichoric acid (right track) and standard solution containing chlorogenic acid and caffeic acid (left track).

Figure 3. TLC schematic figure of sample containing caftaric acid and cichoric acid (right track) and standard solution containing chlorogenic acid and caffeic acid (left track).

Figure 4. HPLC chromatogram of 0.25 mg/5 ml standard solution (chlorogenic acid).

Figure 5. HPLC chromatogram of 2.5 mg/10 ml sample solution.

acid peak should appear at about 18 minute. Then the cichoric acid concentration is calculated, based on equa-
tion which was mentioned in Equation (1). The HPLC experimental data obtained is shown in Table 1.

Calculation results shows that cichoric acid concentration is about \((1.5 \pm 0.65)\%\) (w/w) in the aerial parts of Echinacea purpurea cultivated in Iran. This amount show good agreement with native plant \((1.2-3.1)\%\). This means that the local conditions of cultivation of this herbal drug have no significant effects on its medicine marker.

Table 1. The HPLC experimental data.

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<tr>
<td>(C_s)</td>
<td>0.047 mg/mL</td>
</tr>
<tr>
<td>(C_t)</td>
<td>2.5 mg/mL</td>
</tr>
<tr>
<td>(A_s)</td>
<td>4934823</td>
</tr>
<tr>
<td>(A_t)</td>
<td>5977445</td>
</tr>
<tr>
<td>(C_c)</td>
<td>1.58%</td>
</tr>
<tr>
<td>SD</td>
<td>(\pm 0.65)%</td>
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\(C_s\): concentration of standard solution (chlorogenic acid).
\(A_s\): the peak area for chlorogenic acid.
\(C_t\): concentration of sample solution containing cichoric acid.
\(A_t\): the peak area for cichoric acid.
\(C_c\): concentration of cichoric acid in sample solution.
SD: Standard deviation \((n = 3)\).
4. Conclusions

The qualitative and quantitative analysis of Cichoric acid content in Echinacea purpurea which is cultivated in Iran has been studied. Cichoric acid is probably one of the most important markers for controlling the quality of drugs containing Echinacea purpurea. The results showed that the cichoric acid content of Echinacea purpurea is appropriate for drugs in comparison with native plants.

5. References


