Increased oxidative stress and altered antioxidants status in patients with chronic allergic rhinitis

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ABSTRACT

Background: Allergic rhinitis is an inflammatory disorder of the upper airways. Although several oxidants and antioxidants are likely to be involved, alterations in only limited parameters have been studied. Objective: In this study an attempt has been made to study the oxidant-antioxidant imbalance by investigating changes in a wide range of oxidants and antioxidants in the blood. Methods: Blood samples were obtained from 39 chronic allergic rhinitis patients (males 24, females 15), aged 20 - 70 (mean age 36.33 ± 2.03) years and 53 individuals (36 males, 17 females); aged 24 to 64 (mean age 45.42 ± 1.36) years. Duration of allergic rhinitis was 1.77 ± 0.237 years. In the study group, nasal symptoms were scored and the results were recorded. The patients were classified as having perennial Allergic rhinitis (PAR) if they had had at least 2 rhinitis symptoms (sneezing, rhinorrhea, nasal obstruction, itching) for at least 6 months a year in the previous 2 years and if they had a positive skin prick test response to at least 1 clinically significant perennial allergen (e.g., house dust mites, molds, cockroach, cockroach excrement grass and tree pollen, cat and dog epithelia and molds, or animal dander). They had no other allergic diseases except persistent allergic rhinitis diagnosed by the physical and history examination. Erythrocyte lipid peroxidation, erythrocyte antioxidants viz., glutathione, glutathione reductase, superoxide dismutase, catalase and plasma antioxidants viz., ceruloplasmin, glutathione-S-transferase, vitamin C, total antioxidant activity were estimated in the above two groups. Results: Erythrocyte lipid peroxidation (0 hour, p < 0.01) and superoxide dismutase (p < 0.01) were significantly higher, whereas plasma vitamin C (p < 0.001), ceruloplasmin (p < 0.05) and total antioxidant activity (p < 0.001) were significantly lower in chronic allergic rhinitis patients when compared to controls. Plasma glutathione S-transferase and erythrocyte catalase, glutathione, and glutathione reductase remained unchanged from normal subjects. Conclusion: The changes in different parameters indicate an imbalance in the oxidant and antioxidant status in chronic allergic rhinitis patients. Further studies are required to investigate the potential for antioxidant supplements to be used as routine therapy in chronic allergic rhinitis patients. Capsule Summary: The study shows that the body is trying to cope for the oxidative stress by altering the enzyme levels. But external supplement may also be required as the total antioxidant levels are very much depleted.

Keywords: Oxidative Stress; Antioxidant Status; Chronic Allergic Rhinitis

1. INTRODUCTION

Allergic rhinitis represents a global health issue affecting between 10% and 20% of the world population, with increasing prevalence over the last decade [1]. It is a very common disorder affecting people at all ages and has been associated with significant impairments in quality of life, sleep and work performance [2]. AR is a complex disease characterized by inflammation of the nasal mucosa, along with paroxysms of sneezing, itching of the eyes, nose and palate, rhinorrhea and nasal obstruction [3]. In allergic rhinitis, numerous inflammatory cells, including mast cells, CD4-positive T cells, B cells, macrophages, and eosinophils, infiltrate the nasal lining upon exposure to an inciting allergen (most commonly airborne dust mite fecal particles, cockroach residues, animal dander, moulds, and pollens). The T cells infiltrating the nasal mucosa are predominantly T helper (Th) 2 in nature and release cytokines (e.g., interleukin [IL]-3, IL-4, IL-5, and IL-13) that promote immunoglobulin E (IgE) production by plasma cells. IgE production, in turn, triggers the release of mediators, such as histamine and leukotrienes, that are responsible for arteriolar dilation, increased vascular permeability, itching, rhinorrhea (runny
nose), mucous secretion, and smooth muscle contraction [1,2]. The mediators and cytokines released during the early phase of an immune response to an inciting allergen, trigger a further cellular inflammatory response over the next 4 to 8 hours (late-phase inflammatory response) which results in recurrent symptoms (usually nasal congestion) [1,4].

ROS generation through normal cellular metabolism and by means of exogenous insults is a constant problem for which cells have developed multiple protective mechanisms to survive. For example, cigarette smoke inhalation results in increased exposure to both superoxide and hydrogen peroxide [5,6]. To protect itself against exposure to noxious oxidants, the airway mucosa has developed an antioxidant system. Elevated levels of ROS such as hydroxyl radicals, superoxides, and peroxides may induce a variety of pathological changes that are highly relevant in nasal and airway mucosas. These include lipid peroxidation, increased airway reactivity, increased nasal mucosal sensitivity and secretions, production of chemotaxant molecules, and increased vascular permeability. The association between chronic inflammation and oxidative stress is well documented. Oxidative stress results in an imbalance between the oxidative forces and the antioxidant defense systems, which is believed to favor an oxidative injury that has been implicated in the pathogenesis of asthma and AR [7,8]. In the studies carried out thus far, the investigators have studied changes in only a few parameters. The role of oxidative stress in allergic rhinitis is not well studied but is likely to be similar to that in asthma [9]. Because several oxidants and antioxidants are likely to be involved in the pathogenesis of the inflammatory process in allergic rhinitis, a comprehensive study of several parameters of oxidative stress and antioxidant defenses is required to highlight the role of oxidant-antioxidant imbalance in allergic rhinitis. The present study was carried out with this aim.

2. MATERIALS AND METHODS

2.1. Study Design

The study plan was approved by the Ethics Committee of the Medical Faculty, and all subjects volunteered for the trial. Patient group age ranged was 20 to 70 years (mean age, 36.33 ± 2.03); the control group age range was 24 to 64 years (mean age, 45.42 ± 1.36). In the patients group, 24 patients were male and 15 patients were female. In the control group, 36 patients were male and 17 patients were female. In the patients group, the mean complaint time ranged from 1.77 ± 0.237 year. In the study group, nasal symptoms were scored and the results were recorded. The patients were classified as having perennial Allergic rhinitis (PAR) if they had had at least 2 rhinitis symptoms (sneezing, rhinorrhea, nasal obstruction, itching) for at least 6 months a year in the previous 2 years and if they had a positive skin prick test response to at least 1 clinically significant perennial allergen (e.g., house dust mites, molds, cockroach, cockroach excrement grass and tree pollen, cat and dog epithelia and molds, or animal dander). They had no other allergic diseases except persistent allergic rhinitis diagnosed by the physical and history examination.

2.2. Methods

Random blood samples were collected in heparinized bottles from normal subjects and allergic rhinitis patients. The erythrocyte suspension was prepared according to the method of Kartha and Krishnamurthy [10]. Blood was centrifuged at 3000 g for 10 minutes. Plasma and buffy coat were carefully removed and the separated cells washed thrice with cold saline phosphate buffer, pH 7.4 (sodium phosphate buffer containing 0.15 M NaCl). The RBCs were then suspended in an equal volume of physiological saline. This suspension was used for some of the assays performed. The assays performed in the erythrocytes were lipid peroxidation (LP), glutathione (GSH), glutathione reductase (GR), catalase (CT) and in plasma were glutathione-S-transferase (GST), vitamin C, ceruloplasmin, antioxidant activity (AOA).

The hemoglobin content of the erythrocytes was determined by the cyaanmethemoglobin method. Erythrocyte LP was determined by incubating RBC suspension in saline phosphate buffer containing 0.44 M H2O2 at 0 hour and 2 hours. Aliquots were drawn from the above mixture at 0 hour and 2 hours. Lipid peroxidation in RBC was determined by estimating malondialdehyde (MDA) produced using thiobarbituric acid [11]. Erythrocyte GR activity was determined by recording the decrease in absorbance due to depletion of NADPH for a period of 5 minutes at 340 nm [12]. SOD was determined according to the method of Beauchamp and Fridovich [13] based on inhibition of nitrozoium reduction. CT activity in the hemolysate was determined by adopting the method of Brannan et al. [14]. The assay is based on the disappearance of H2O2 in the presence of the enzyme source at 26°C. The GSH content of erythrocytes was determined as described by Beutler et al. [15].

Plasma ceruloplasmin was determined by p-phenylene diamine oxidase activity [16]. Plasma vitamin C was determined chemically using dinitrophenyl hydrazine as a colour compound [17]. Plasma GST was determined by incubating CDNB (1 chloro 2,4 dinitro benzene) with reduced GSH in the presence of serum containing glutathione-S-transferase. 2,4-dinitrophenylglutathione (adduct) formed was read at 340 nm [18]. AOA activity was measured by Fenton type reaction [19]. The package used for statistical analysis was SPSS/PC+ (version...
11.0).

3. RESULTS

3.1. Oxidative Changes

The mean TBARS level estimated in allergic rhinitis patients was high at 0 hour and 2 hours compared to normal controls (Table 1). The susceptibility towards lipid peroxidation, indicated by the difference of the two values, obtained at 0 hour and 2 hour was also increased compared to normal controls. But only the increase observed at 0 hour was significant (p < 0.01).

3.2. Erythrocyte Antioxidants

The mean SOD enzyme activity was increased significantly in the erythrocytes of patients with allergic rhinitis when compared to normal controls (NC, p < 0.01) (Table 2). No statistical significant variation in mean glutathione level, catalase activity, and glutathione reductase activity was observed in patients when compared with normal controls.

3.3. Plasma Antioxidants

The mean vitamin C levels decreased significantly in case of allergic rhinitis patients compared to normal controls (Table 3). There was a significant rise in mean ceruloplasmin levels in allergic rhinitis patients when compared to normal controls (Table 3).

Table 1. In vitro RBC lipid peroxidation in allergic rhinitis.

<table>
<thead>
<tr>
<th>Group</th>
<th>TBARS as nmol MDA/g Hb</th>
<th>Mean ± SEM</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0 Hour</td>
<td>2 Hours</td>
</tr>
<tr>
<td>Normal Controls (NC)</td>
<td>77.8 ± 4.46</td>
<td>384.5 ± 18.54</td>
</tr>
<tr>
<td></td>
<td>(20.8 - 181.6)</td>
<td>(102.8 - 898.7)</td>
</tr>
<tr>
<td>Allergic Rhinitis</td>
<td>100.0 ± 6.69**</td>
<td>461.8 ± 34.39</td>
</tr>
<tr>
<td></td>
<td>(37.7 - 232.6)</td>
<td>(123.6 - 1002.8)</td>
</tr>
<tr>
<td>% Change (%)</td>
<td>28.53% &gt; NC</td>
<td>20.10% &gt; NC</td>
</tr>
</tbody>
</table>

Significance of results vs. NC: **p < 0.01, NS = not significant; Ranges of TBARS levels observed are given in parentheses; n = number of cases; (Kruskal Wallis Test—Mann Whitney Test).

Table 2. Erythrocyte antioxidant levels in allergic rhinitis (Mean ± SEM).

<table>
<thead>
<tr>
<th>Diagnosis</th>
<th>GSH (µmol/g Hb)</th>
<th>SOD (units/g Hb)</th>
<th>Catalase (units/g Hb)</th>
<th>GR (units/g Hb)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0 Hour</td>
<td>2 Hours</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Normal Controls (NC)</td>
<td>4.79 ± 0.209</td>
<td>9214 ± 492.5</td>
<td>245,996 ± 10410.2</td>
<td>1.77 ± 0.153</td>
</tr>
<tr>
<td></td>
<td>(2.36 - 10.25)</td>
<td>(4046 - 21,990)</td>
<td>(27,920 - 413,385)</td>
<td>(0.10 - 4.09)</td>
</tr>
<tr>
<td></td>
<td>n = 53</td>
<td>n = 53</td>
<td>n = 53</td>
<td>n = 51</td>
</tr>
<tr>
<td>Allergic Rhinitis</td>
<td>5.33 ± 0.390</td>
<td>11,267 ± 777.9*</td>
<td>240,358 ± 18213.4</td>
<td>1.90 ± 0.391</td>
</tr>
<tr>
<td></td>
<td>(0.90 - 10.38)</td>
<td>(2826 - 23,621)</td>
<td>(73,886 - 531,720)</td>
<td>(0.00 - 12.97)</td>
</tr>
<tr>
<td></td>
<td>n = 39</td>
<td>n = 39</td>
<td>n = 39</td>
<td>n = 36</td>
</tr>
<tr>
<td>% Change (%)</td>
<td>11.27% &gt; NC</td>
<td>23.36% &gt; NC</td>
<td>2.29% &lt; NC</td>
<td>7.34% &lt; NC</td>
</tr>
</tbody>
</table>

Statistical significance of results vs. NC: *p < 0.01, NS = not significant; The figures in the parentheses indicate the ranges of antioxidant levels observed; n = number of cases; (Kruskal Wallis Test—Mann Whitney Test).
Table 3. Plasma antioxidant levels in allergic rhinitis patients (Mean ± SEM).

<table>
<thead>
<tr>
<th>Diagnosis</th>
<th>Vitamin C (µmol/L)</th>
<th>Ceruloplasmin (g/L)</th>
<th>GST (IU/L)</th>
<th>AOA (mmol/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal Controls (NC)</td>
<td>22.5 ± 1.23 (3.5 - 49.5) n = 53</td>
<td>0.479 ± 0.0268 (0.225 - 1.40) n = 53</td>
<td>4.31 ± 0.45 (0.410 - 15.41) n = 53</td>
<td>1.03 ± 0.060 (0.32 - 2.20) n = 53</td>
</tr>
<tr>
<td>Allergic Rhinitis</td>
<td>13.9 ± 1.78*** $ (1.5 - 51.0) n = 39</td>
<td>0.615 ± 0.0728* (0.112 - 2.310) n = 39</td>
<td>4.66 ± 0.522 (1.04 - 14.58) n = 39</td>
<td>0.61 ± 0.042*** # (0.12 - 1.24) n = 36</td>
</tr>
<tr>
<td>% Change</td>
<td>38.22% &lt; NC</td>
<td>28.39% &gt; NC</td>
<td>8.12% &gt; NC</td>
<td>40.70% &lt; NC</td>
</tr>
</tbody>
</table>

Statistical significance of results vs. NC: *p < 0.05, **p < 0.01, ***p < 0.001, NS = not significant; The figures in the parentheses indicate the ranges of antioxidant levels observed; n = number of cases; ($) = ANOVA- Dunnett t Test, # = Kruskal Wallis Test—Mann Whitney Test.

Ascorbic acid is an antioxidant vitamin which is phylogenetically available in respiratory tracts. As an antioxidant, it may cause a reduction of negative effects caused by oxidative attack on tissues during inflammation. Smoking is known to increase the metabolic turnover of ascorbic acid due to its oxidation by free radicals and ROS generated by cigarette smoking. Ascorbic acid has been shown to stimulate immune system by enhancing T-cell proliferation in response to infection. These cells are capable of lysing infected targets by producing large quantities of cytokines and by helping B-cells to synthesize immunoglobulins to control inflammation reaction. Plasma devoid of vitamin C has increased rate of lipid peroxidation. On the contrary, plasma in the presence of vitamin C has decreased rate lipid peroxidation. This indicates that vitamin C is an important antioxidant in the plasma. Oxidative stress in allergic rhinitis patients caused by increased oxygen free radicals might have increased consumption of vitamin C to mitigate the toxicity of free radicals. This in turn, might have resulted in decreased plasma vitamin C level. Thus, a deficiency of plasma vitamin C level must have increased the susceptibility of RBC towards lipid peroxidation in allergic rhinitis patients. In another study on children with asthma, it was shown that the reduction in vitamin C causes and increases the risk of asthma. Kalayci et al. [30] showed that children with asthma had lower level of ascorbic acid when comparing with healthy children in the cycle of remission. In our study, a significant decrease has been observed in the plasma vitamin C and total antioxidant level between the AR and control groups. Podoshin et al. [31] showed that vitamin C reinforcement reduced the symptoms in 74%. A diet rich in antioxidants has been associated with a low prevalence of allergic disease [32].

Ceruloplasmin levels, in the present study, have significantly increased in patients suffering from allergic rhinitis. It is an acute phase protein and also has antioxidant activity. Ceruloplasmin catalyzes oxidation of ferrous ions and does not release any damaging oxygen radicals. The ferroxidase activity of ceruloplasmin allows it to inhibit iron dependent lipid peroxidation or OH· formation from hydrogen peroxide and superoxide ions. Since ceruloplasmin is a copper binding enzyme, it also sequesters the copper ions in the plasma and reduces its deleterious effects on RBC. The rise in ceruloplasmin in the present study may have occurred in response to inflammation of airways as well as a part of an increased antioxidant response to cell injury.
The present study is the first to examine changes in several measures of oxidative stress and antioxidant status in allergic rhinitis. Previous studies have focused on only a few parameters, but we have investigated alterations in a wide variety of antioxidants. Taken together, these results provide new information on the role of oxidative stress in allergic rhinitis. A strategy for designing well-balanced antioxidant therapies based on both reducing endogenous ROS production and increasing the total antioxidant capacity of human cells may prove useful in the prevention of allergic rhinitis.

REFERENCES


